Internal radiation dose assessment after administration of radiopharmaceuticals prepared with cyclotron-produced ^{99m}Tc

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ABSTRACT (500 words)

Purpose: ^{99m}Tc is the radionuclide most widely used in diagnostic nuclear medicine. It is readily available from ⁹⁹Mo/^{99m}Tc generators as a β'decay product of the ⁹⁹Mo (T_{1/2}=66 h) parent nuclide. The latter is obtained as a fission product in nuclear reactors by neutron-induced reactions on highly enriched uranium. Alternative production routes, such as direct reactions using proton beams on specific target materials [e.g. ¹⁰⁰Mo(p,2n)^{99m}Tc] are a reliable and relatively cost-effective method. However, the results of the LARAMED (Laboratory of Radionuclides for Medicine) project research from the Legnaro National Laboratories of the National Institute for Nuclear Physics (LNL-INFN), showed that the extracted ^{99m}Tc from the proton-bombarded ¹⁰⁰Mo-enriched target contains small quantities of several technetium radioisotopes (^{93m}Tc, ⁹³Tc, ⁹⁴Tc, ^{94m}Tc, ⁹⁵Tc, ^{95m}Tc

The aim of this work was to determine the patient dose increase (DI) due to the contribution of Tcradioisotopes generated as impurities, after the administration of four radiopharmaceuticals prepared with cyclotron-produced ^{99m}Tc (CP-^{99m}Tc).

Methods: Four ^{99m}Tc-radiopharmaceuticals (pertechnetate, sestamibi, hexamethylpropylene-amine oxime (HMPAO) and disodium etidronate (HEDP)), were considered in this study. The biokinetic models reported by the International Commission on Radiological Protection (ICRP) for each radiopharmaceutical were used to determine the main source organs and to calculate the number of disintegrations that occurred in each source organ (N_{source}) for each Tc-radioisotope present in the CP-^{99m}Tc solution. Then, equivalent dose in the main organs and effective dose were calculated for each Tc-radioisotope with the OLINDA/EXM software versions 1.1 and 2.0, using the calculated N_{source} and the adult male phantom as program inputs. Finally, the total effective dose produced by all Tc-radioisotopes present in the CP-^{99m}Tc solution was calculated at five different times after the end of bombardment (EOB), using ¹⁰⁰Mo-enriched (i.e. 99.05 %) metallic target, and compared with the effective dose delivered by the generator-produced-^{99m}Tc.

Results: In all cases, the total effective DI of CP-^{99m}Tc-radiopharmaceuticals, calculated with both versions of the OLINDA software, remained within the 10% limit from 6 up to 12 h after EOB. ^{94m}Tc and ^{93m}Tc are the Tc-radionuclides with the highest concentration in the CP-^{99m}Tc solution at EOB. However, their contribution to DI 6 h after EOB is minimal, due to of their short half-life. Although their concentration in the CP-^{99m}Tc solution is 5 times less than ^{94m}Tc and ^{93m}Tc at the EOB, the radioisotopes with the largest contribution to the effective DI are ⁹⁶Tc, followed by ⁹⁵Tc and ⁹⁴Tc, due to the type of their emissions and relatively long half-life.

Conclusion: The increase in the patient radiation dose caused by the other technetium-nuclides contained in CP-^{99m}Tc produced as described in this paper, is quite low. Therefore, it was concluded that although in the CP-^{99m}Tc solution the concentrations of two radionuclides, ⁹⁴Tc and ⁹⁵Tc, are outside the limits established by the European Pharmacopoeia, it could be used in routine nuclear medicine diagnostic studies, but would have to be administered from 6 to 12 h after the EOB to maintain the effective DI within the 10 % limit.

Keywords:

cyclotron-produced ^{99m}Tc, ^{99m}Tc-radiopharmaceuticals effective dose, Tc-dosimetry, Dose Increase (DI).

1. INTRODUCTION

Technetium-99m is the radionuclide most widely used in diagnostic nuclear medicine. Sodium pertechnetate is used as raw material to prepare more than twenty ^{99m}Tc-radiopharmaceuticals employed for the detection of diverse physiological and pathological conditions such as myocardial perfusion, kidney and brain disorders as well as bone metastases¹. The radioisotope is readily available in hospitals from ⁹⁹Mo/^{99m}Tc generators, as β decay product of ⁹⁹Mo, which in turn, is a selected and purified fission product of weapon-grade highly enriched uranium (HEU-WG), special targets in nuclear reactors. However, the majority of ⁹⁹Mo isotope global production is the work of less than seven nuclear reactor facilities worldwide and most of them are close to being permanently shut down at the end of their running periods (more than 50 years of operation). Therefore, alternative production routes for both ⁹⁹Mo and ^{99m}Tc nuclides are currently being evaluated²⁻⁴. Among them, the use of cyclotrons to produce either ⁹⁹Mo or ^{99m}Tc is one of the most interesting and promising approaches^{5,6}.

Experimental studies on ^{99m}Tc production by cyclotrons were initiated within the framework of the TECHN_OSP/LARAMED (Laboratory of Radionuclides for Medicine) research project, by the Legnaro National Laboratories at the National Institute for Nuclear Physics, Italy (LNL-INFN)⁷. A PETtrace GE cyclotron was used to obtain cyclotron-produced ^{99m}Tc (CP-^{99m}Tc), using the ¹⁰⁰Mo(p,2n)^{99m}Tc reaction on ¹⁰⁰Mo-enriched (i.e. 99.05 %) metallic targets, the isotopic composition of the enriched molybdenum used was a good compromise between enriched and cost⁸. ^{99m}Tc was extracted and purified from the irradiated targets by solvent extraction through the Methyl-Ethyl Ketone (MEK) method, using a self-developed automated module with an efficiency greater than 90%⁹.

Results demonstrated that the direct reaction method is reliable and a relatively cost-effective option to produce ^{99m}Tc. However, gamma-ray spectrometry analyses of the obtained purified ^{99m}Tc solutions demonstrated the presence of small quantities of several technetium radioisotopes, such as ⁹³Tc, ^{93m}Tc, ^{94m}Tc, ^{95m}Tc, ^{95m}Tc, ⁹⁶Tc and ^{97m}Tc⁸. It has been reported by previous studies that the

abundance of these contaminant nuclides is basically dependent on some changing factors, such as the target's isotopic composition, beam energy level, target thickness, irradiation time and time span from irradiation to administration^{10,11}. Despite the presence of technetium isotopes in the solution, all radiopharmaceuticals labelled with CP-^{99m}Tc showed no differences in the labelling efficiency, radiochemical purity and biological behaviour when compared to the radiopharmaceuticals labelled with generator-produced ^{99m}Tc (GP-^{99m}Tc)^{12,13}. It was also demonstrated that scatter-corrected images of rats, obtained after administration of radiopharmaceuticals for myocardial perfusion and bone imaging labelled with CP-^{99m}Tc and GP-^{99m}Tc, were comparable^{12,14}. Selivanova *et al* confirmed these results demonstrating no difference in whole body biodistribution images of patients, after administration of sodium pertechnetate produced by cyclotron and generator¹⁵.

The main problem with the technetium impurities is that they contribute to the radiation dose of the patient¹⁰. Thus, the European Pharmacopoeia Commission established a monograph of sodium pertechnetate obtained by proton irradiation of highly-enriched molybdenum-100 targets, in order to set the limits of radionuclide impurities for the solutions used in clinical diagnosis¹⁶.

The aim of this work was to determine the increase of the total effective dose in patients due to the contribution of all Tc-radionuclides produced as impurities. Effective dose increase was calculated for four radiopharmaceuticals prepared with ^{99m}Tc produced by a PETtrace GE cyclotron, evaluating five times points after the end of bombardment (EOB), in order to find the best period when the CP-^{99m}Tc radiopharmaceuticals could be used.

2. MATERIALS AND METHODS

Four of the ^{99m}Tc-radiopharmaceuticals most used in the clinical diagnosis at the Nuclear Medicine Department of the Veneto Institute of Oncology IOV-IRCCS were considered for this study: sodium pertechnetate, used in clinical diagnosis of thyroid function and morphology; disodium etidronate (HEDP), a phosphonate commonly used for defining bone metastasis in cancer patients; hexamethylpropyleneamine oxime (HMPAO), used as a tracer of brain function and sestamibi (MIBI), widely used in cardiac scans for diagnosis of heart disease. The total effective dose increases

were assessed considering the administration of the four radiopharmaceuticals labelled with both GP-^{99m}Tc and CP-^{99m}Tc produced by the alterative ¹⁰⁰Mo(p,2n) reaction route, through a GE PETtrace cyclotron and using enriched (99.05%) molybdenum-100 metallic targets.

2.1 Number of disintegrations in the source organs

The biokinetic models published by the International Commission on Radiological Protection (ICRP) were used to determine the main source organs for each radiopharmaceutical. Pertechnetate and HEDP biokinetic models were obtained from ICRP publication 53: "Radiation dose to patients from radiopharmaceuticals"¹⁷; HMPAO and MIBI models were obtained from publication 80 (addendum 2 to ICRP publication 53)¹⁸. The cumulated activities, or number of disintegrations that occurred in the source regions over time (N_{source}), were calculated for each radionuclide taking into account the related biological elimination constant for each source organ reported by the ICRP radiopharmaceutical models.

The number of disintegrations in the bladder (N_b) for each radionuclide was calculated for all 4 radiopharmaceuticals using the kidney-bladder model presented in ICRP publication 106 (addendum 3 to ICRP publication 53)¹⁹, with the following formula:

$$N_b = f_s \sum_{i} a_i \left[\frac{1 - \exp(-\lambda_p t_b)}{\lambda_p} - \frac{1 - \exp(-(\lambda_i + \lambda_p) t_b)}{\lambda_i + \lambda_p} \right] \left[\frac{1}{1 - \exp(-(\lambda_i + \lambda_p) t_b)} \right]$$

where *fs* is the fractional distribution to organ or tissue, a_i is the fraction of f_s eliminated with the corresponding biological constant λ_i , λ_p is the physical decay constant, and t_b is the bladder filling and voiding interval (considered a constant of 3.5 h in adults).

Kidney and bladder are the organs that received the highest dose after administration of phosphonates such as HEDP, because they are mainly eliminated by the kidney. Thus, the number of disintegrations in the kidney (N_K) was calculated using the kidney-bladder model from ICRP 53

with the following formula (considering both kidney uptake and the activity that passes through this organ due to excretion):

$$N_{K} = f_{K} \left[\frac{1 - \exp(-\lambda_{p} t_{K})}{\lambda_{p}} \right] \sum_{i} a_{i} \frac{\lambda_{i}}{\lambda_{i} + \lambda_{p}}$$

where f_K is the kidney fractional distribution and t_K is the mean transit time of phosphonates in the kidney. The other 3 radiopharmaceuticals are eliminated predominantly by the gastro-intestinal (GI) system; therefore, the number of disintegrations that occurred in this tract were calculated using the GI model of the OLINDA/EXM 1.1 software²⁰, which is based on the ICRP 30 GI model²¹.

2.1 Dose Calculations

Each organ's absorbed dose was calculated for each Tc radioisotope with the OLINDA/EXM 1.1 and OLINDA/EXM 2.0 software codes. The OLINDA version 1.1 makes use of anthropomorphic phantoms based on the Oak Ridge models, which employs geometrical shapes to define the organs and calculates the effective dose using the tissue weighting factors recommended by ICRP 60²². In version 2.0, the geometrical models were replaced by the realistic NURBS-type models, based on the recent standardized masses defined by the ICRP in its publication 89²³, and the tissue weighting factors were implemented in accordance with the recommendations of ICRP 103²⁴.

Both versions of OLINDA/EXM software use the RADAR dose system and calculate the equivalent dose in each organ according to the general equation:

$\overline{D}_{target \leftarrow source} = N_{source} \times DF_{target \leftarrow source}$

where N_{source} is the number of disintegrations that occur in the source organ per unit of activity administered (MBq-h/MBq) and *DF* is a dose factor that depends on the radionuclide used and on the spatial relationship between the target and the source organs, and on their tissue compositions. The dose factor *DF* is given as:

$$DF = \frac{k \sum_{i} n_i E_i \phi_i w_R}{m}$$

where m is the mass of the target organ, n_i is the number of *i*-th nuclear transitions per nuclear transformation, E_i is the mean energy of the *i*-th nuclear transition, ϕ_i is the absorbed fraction in the target organ of radiation energy E_i emitted from the source organ, w_R is the radiation weighting factor assigned to the *i*-th radiation and k is a constant (which value depends on the units of the included quantities).

To calculate the equivalent dose in the main organs delivered from the radiopharmaceuticals labelled with each one of the Tc radioisotopes present in the CP-^{99m}Tc solution, the number of disintegrations of all source organs calculated for each radionuclide and the adult male phantom were used as program inputs with both versions of OLINDA/EXM software. Effective dose (ED) for each Tc radioisotope was calculated by OLINDA as the tissue-weighted sum of the equivalent doses. Total effective dose (*ED_t*) produced by the CP-^{99m}Tc was calculated at 5 different times after irradiation (6, 8, 10, 12 and 15 h) using the following equation:

$ED_t = \sum_i f_i ED_i$

where f_i is the fraction of total activity corresponding to each radioisotope *i* and *ED_i* is the effective dose contribution per unit of activity from each radioisotope *i*. The values for f_i parameters were obtained from the experimental results found in the framework of the TECHN_OSP/LARAMED research project on cyclotron-Tc production. The relative activities of ^{99m}Tc and the other technetium isotopes, obtained after irradiation of ¹⁰⁰Mo-enriched (99.05%) metallic targets with 15.7 MeV proton beam for 60 min, were used as reference data⁸. The target irradiation was carried out on the model PETtrace 16-9 medical cyclotron, located at St. Orsola-Malpighi Hospital, Nuclear Medicine Department (Bologna, Italy). Table 1 shows the average of relative activities of the technetium nuclides in the extracted solutions, obtained from 3 experiments and calculated at the EOB⁸. Table 2 shows the radionuclide's impurity limits for human use of sodium pertechnetate, reported by the European Pharmacopoeia, version 9.3, and the fraction of the total activity corresponding to each Tc-radioisotope calculated for the five time points after EOB studied.

Table 1. Technetium-nuclide relative activities in the extracted solution calculated for EOB (see text for irradiation parameters).

Radioisotope	Physical half-life (h)	Radioactive concentration (%)
^{99m} Tc	6.01	96.59 ± 0.62
^{97m} Tc	2193.6	0.001130 ± 0.000064
⁹⁶ Tc	102.72	0.0273 ± 0.0033
⁹⁵ Tc	20	0.1229 ± 0.0243
^{95m} Tc	1464	0.00063 ± 0.00013
⁹⁴ Tc	4.88	0.1577 ± 0.0367
^{94m} Tc	0.87	0.8932 ± 0.3868
⁹³ Tc	2.75	0.05429 ± 0.01214
^{93m} Tc	0.73	2.15 ± 0.09

Table 2. Radionuclide impurity limits reported by the European Pharmacopoeia, version 9.3, and the fraction of the total activity corresponding to each Tc-radioisotope in the extracted solution, calculated for five different time points after EOB.

	Fraction of technetium isotopes contribution to the total activity (%)								
Pharmacopoeia Limit	^{99m} Tc	^{97m} Tc	⁹⁶ Tc	⁹⁵ Tc	^{95m} Tc	⁹⁴ Tc	^{94m} Tc	⁹³ Tc	^{93m} Tc
		0.010	0.070	0.070	0.005	0.040	0.02	0.040	0.010
Time after EOB (h)	^{99m} Tc	^{97m} Tc	⁹⁶ Tc	⁹⁵ Tc	^{95m} Tc	⁹⁴ Tc	^{94m} Tc	⁹³ Tc	^{93m} Tc
0	96.59	0.001	0.027	0.123	0.001	0.158	0.893	0.054	2.154
6	99.54	0.002	0.054	0.206	0.001	0.139	0.0152	0.025	0.0134
8	99.53	0.003	0.067	0.242	0.002	0.131	0.0039	0.019	0.0024
10	99.49	0.004	0.083	0.284	0.002	0.125	0.0010	0.014	0.0004
12	99.43	0.005	0.104	0.334	0.003	0.118	0.0002	0.011	0.0001
15	99.31	0.007	0.143	0.425	0.004	0.109	0.0000	0.007	0.0000

3. RESULTS

The total number of disintegrations in the main source organs depends directly on the pharmacokinetics of each complex and on the half-life of the radionuclide. The calculated values, for each Tc-radioisotope after administration of Tc-pertechnetate, Tc-HEDP and Tc-HMPAO radiopharmaceuticals are summarized in Table 3. For each radiopharmaceutical, ^{97m}Tc and ^{95m}Tc

are the radionuclides that produced the highest number of the disintegrations per unit of administered activity (see table 3), due to their long half-life.

Table 3. Calculated number of disintegrations in the source organs per each MBq of ^{9x}Tc-radiopharmaceuticals administered.

	Number of disintegrations per unit of administered activity (MBq h/MBq)									
Source organ	^{99m} Tc	^{97m} Tc	⁹⁶ Tc	⁹⁵ Tc	^{95m} Tc	⁹⁴ Tc	^{94m} Tc	⁹³ Tc	^{93m} Tc	
Tc-Pertechnetate										
Thyroid	0.037	0.068	0.064	0.052	0.067	0.035	0.015	0.027	0.013	
Salivary gland	0.056	0.101	0.096	0.078	0.101	0.052	0.022	0.041	0.020	
Stomach content	0.179	0.200	0.199	0.193	0.200	0.175	0.111	0.160	0.102	
Stomach wall	0.247	0.288	0.286	0.275	0.288	0.239	0.134	0.212	0.122	
Small intestine content	0.548	0.799	0.779	0.703	0.799	0.510	0.191	0.398	0.166	
Upper large int. walls	0.379	1.506	1.341	0.878	1.501	0.308	0.027	0.156	0.020	
Upper large int. content	0.712	2.586	2.328	1.587	2.579	0.583	0.054	0.303	0.040	
Lower large int. content	0.349	4.737	3.698	1.574	4.708	0.244	0.005	0.079	0.003	
Bladder content	0.342	0.963	0.807	0.554	0.958	0.306	0.063	0.209	0.049	
Others	4.318	28.533	20.711	9.908	28.267	3.678	0.852	2.324	0.725	
Tc-HEDP										
Bone	2.953	35.453	21.619	8.119	34.904	2.443	0.425	1.423	0.346	
Kidneys	0.125	0.869	0.553	0.245	0.856	0.113	0.062	0.088	0.060	
Bladder content	1.147	2.151	1.856	1.490	2.139	1.072	0.319	0.833	0.256	
Total body	4.005	41.310	25.492	10.031	40.683	3.448	0.896	2.213	0.777	
Tc-HMPAO										
Brain	0.407	6.635	3.580	1.194	6.499	0.335	0.062	0.193	0.052	
Thyroid	0.044	0.356	0.249	0.110	0.353	0.037	0.008	0.022	0.007	
Lung	0.708	8.585	5.226	1.953	8.451	0.587	0.118	0.347	0.100	
Liver	0.505	1.372	1.243	0.890	1.369	0.446	0.131	0.306	0.114	
Gall bladder content	0.097	0.130	0.127	0.118	0.130	0.092	0.039	0.075	0.035	
Stomach wall	0.047	0.736	0.398	0.134	0.721	0.039	0.008	0.023	0.007	
Small intestine wall	0.203	3.170	1.716	0.579	3.106	0.168	0.033	0.099	0.028	
Upper large int. wall	0.066	1.030	0.558	0.188	1.009	0.055	0.011	0.032	0.009	
Lower large int. wall	0.050	0.781	0.423	0.143	0.765	0.041	0.008	0.024	0.007	
Stomach content	0.006	0.007	0.007	0.006	0.007	0.074	0.004	0.005	0.003	
Small intestine content	0.233	0.340	0.331	0.299	0.339	0.189	0.081	0.169	0.071	
Upper large int. content	0.303	1.099	0.989	0.669	1.096	0.216	0.023	0.129	0.017	
Lower large int. content	0.148	2.013	1.572	0.675	2.001	0.091	0.002	0.034	0.001	
Kidneys	0.624	3.082	2.526	1.416	3.066	0.527	0.109	0.320	0.007	
Bladder content	0.546	1.472	1.199	0.818	1.463	0.500	0.134	0.373	0.107	
Others	2.993	24.504	17.147	7.546	24.248	2.515	0.570	1.545	0.488	

It has been demonstrated that the concentration of Tc-MIBI in the source organs can change depending on whether the myocardial perfusion test is carried out in stress or rest conditions²⁵; thus, the total number of disintegrations in the main source organs after MIBI administration was calculated considering both stress and rest models. Table 4 shows in general, a higher number of disintegrations in the source organs when the test was performed in rest conditions because the Tc-MIBI-concentration in the source organs increases. Muscle and heart are the only organs that present a higher number of disintegrations during the myocardial perfusion test in stress conditions.

Table 4. Calculated number of disintegrations in the source organs per each MBq of ^{9x}Tc-MIBI administered, using both stress and rest models.

	Nu	mber of di	isintegrati	ons per u	nit of adm	ninistered	activity (MBq h/M	Bq)
Source organ	^{99m} Tc	^{97m} Tc	⁹⁶ Tc	⁹⁵ Tc	^{95m} Tc	⁹⁴ Tc	^{94m} Tc	⁹³ Tc	^{93m} Tc
Tc-MIBI (rest)									
Heart	0.069	0.227	0.195	0.126	0.226	0.061	0.016	0.041	0.014
Liver	0.674	2.452	2.057	1.264	2.440	0.596	0.192	0.420	0.168
Gall bladder	0.246	1.213	0.994	0.557	1.207	0.207	0.043	0.126	0.036
Small intestine content	0.496	0.723	0.705	0.636	0.723	0.462	0.172	0.361	0.150
Upper large int. content	0.644	2.340	2.107	1.425	2.334	0.528	0.049	0.274	0.036
Lower large int. content	0.316	4.287	3.347	1.436	4.261	0.221	0.045	0.072	0.003
Kidneys	0.653	1.409	1.324	1.047	1.407	0.581	0.156	0.399	0.133
Bladder content	0.190	0.576	0.498	0.334	0.573	0.168	0.034	0.113	0.027
Muscles	1.387	6.850	5.613	3.148	6.813	1.170	0.242	0.712	0.204
Salivary glands	0.104	0.514	0.421	0.236	0.511	0.088	0.018	0.053	0.015
Thyroids	0.006	0.009	0.008	0.008	0.009	0.006	0.003	0.005	0.002
Others	3.120	15.412	12.630	7.082	15.330	2.633	0.545	1.602	0.460
Tc-MIBI (stress)									
Heart	0.092	0.303	0.260	0.168	0.302	0.081	0.022	0.055	0.019
Liver	0.530	2.132	1.774	1.056	2.121	0.463	0.134	0.313	0.116
Gall bladder	0.202	0.998	0.818	0.459	0.993	0.170	0.035	0.104	0.030
Small int. content	0.385	0.563	0.548	0.495	0.562	0.359	0.134	0.281	0.117
Upper large int. cont.	0.501	1.821	1.639	1.108	1.816	0.410	0.038	0.213	0.028
Lower large int. cont.	0.246	3.335	2.603	1.117	3.314	0.172	0.004	0.056	0.002
Kidneys	0.466	1.007	0.945	0.748	1.005	0.415	0.111	0.285	0.095
Bladder content	0.151	0.545	0.463	0.291	0.543	0.131	0.024	0.085	0.019
Muscles	2.774	13.700	11.227	6.295	13.626	2.340	0.484	1.424	0.409
Salivary glands	0.069	0.342	0.281	0.157	0.341	0.059	0.012	0.036	0.010
Thyroids	0.004	0.006	0.006	0.005	0.006	0.004	0.002	0.003	0.002
Others	2.566	12.672	10.385	5.823	12.605	2.165	0.448	1.317	0.378

Radiopharmaceutical effective doses (ED) calculated for each Tc radioisotope are summarized in table 5. The small difference in the effective dose values, obtained with both versions of OLINDA/EXM software, is attributable to the divergence in the tissue weighting factors and the phantom models used by each version.

Table 5. Radiopharmaceuticals effective doses calculated using both versions of OLINDA/EXM

 for each Tc radioisotope

	Tc-radiopharmaceuticals effective doses (mSv/MBq)									
Nuclide	Tc-Pertechnetate	Tc-HEDP	Tc-HMPAO	Tc-MIBI rest	Tc-MIBI stress					
OLI	NDA/EXM 1.1									
^{99m} Tc	1.07E-02	6.42E-03	9.25E-03	7.94E-03	6.84E-03					
^{97m} Tc	1.44E-01	4.45E-02	1.99E-01	1.14E-01	9.01E-02					
⁹⁶ Tc	4.71E-01	3.71E-01	5.06E-01	4.21E-01	3.83E-01					
⁹⁵ Tc	8.87E-02	5.55E-02	8.01E-02	7.67E-02	6.94E-02					
^{95m} Tc	1.90E-01	1.66E-01	2.40E-01	1.63E-01	1.46E-01					
⁹⁴ Tc	1.13E-01	7.99E-02	9.55E-02	9.14E-02	8.19E-02					
^{94m} Tc	6.58E-02	3.63E-02	4.54E-02	3.70E-02	2.26E-02					
⁹³ Tc	4.23E-02	3.18E-02	3.33E-02	3.14E-02	2.78E-02					
^{93m} Tc	1.12E-02	6.73E-03	6.86E-03	5.71E-03	4.79E-03					
OLII	VDA/EXM 2.0									
^{99m} Tc	1.07E-02	5.24E-03	8.64E-03	8.09E-03	7.04E-03					
^{97m} Tc	1.50E-01	3.82E-02	1.88E-01	1.22E-01	1.00E-01					
⁹⁶ Tc	3.67E-01	3.06E-01	4.37E-01	3.44E-01	3.16E-01					
⁹⁵ Tc	7.29E-02	4.52E-02	6.97E-02	6.74E-02	6.09E-02					
^{95m} Tc	1.55E-01	1.39E-01	2.13E-01	1.38E-01	1.25E-01					
⁹⁴ Tc	1.02E-01	6.31E-02	8.68E-02	8.32E-02	7.46E-02					
^{94m} Tc	6.92E-02	2.94E-02	3.97E-02	3.54E-02	2.44E-02					
⁹³ Tc	3.95E-02	2.48E-02	3.05E-02	2.87E-02	2.55E-02					
^{93m} Tc	1.30E-02	6.09E-03	7.05E-03	6.08E-03	5.26E-03					

Table 6 shows the total effective dose (mSv/MBq) calculated with both software versions, for the radiopharmaceuticals prepared with pure ^{99m}Tc eluted from a generator (GP-^{99m}Tc) and the ^{99m}Tc produced by the cyclotron at different time points after EOB. The increase in effective dose to the patient (in percent), caused by the presence of the other Tc-radionuclides, is also reported in this table.

In general the values of effective dose calculated with version 2.0 of OLINDA/EXM decreased for Tc-HEDP and Tc-HMPAO, remained the same for Tc-pertechnetate, and increased for Tc-MIBI, compared to those obtained using version 1.1 (Table 6). The difference in effective dose after

administration of CP-^{99m}Tc and GP-^{99m}Tc labelled-radiopharmaceuticals were lower using OLINDA/EXM 2.0 except for the case of Tc-HEDP, in which the values remained invariable. All radiopharmaceuticals showed greater differences in effective doses after 15 h of EOB, because the concentration of impurities increases over the time, as shown in table 2.

Table 6. Total effective dose (ED_t) calculated for GP-^{99m}Tc and CP-^{99m}Tc radiopharmaceuticals and percent of patient dose increase (DI) after administration of CP-^{99m}Tc radiopharmaceuticals at different time points after EOB.

	Calculated total effective dose (mSv/MBq) and dose increase for each radiopharmaceutical									
OLINDA/EXM 1.1	Tc-Pertechnetate		Tc-HEDP		Tc-HMPAO		Tc-MIBI rest		Tc-MIBI stress	
Labelled with GP-99mTc	El	D_t	E	D_t	ED_t		El	D_t	ED_t	
	1.07	E-02	6.42	E-03	9.25E-03		7.94	E-03	6.84	E-03
Labelled with CP-99m Tc	ED_t	DI (%)	ED_t	DI (%)	ED_t	DI (%)	ED_t	DI (%)	ED_t	DI (%)
Time after EOB										
6 h	1.13E-02	5.34	6.83E-03	6.44	9.80E-03	5.96	8.43E-03	6.22	7.29E-03	6.53
8 h	1.13E-02	6.04	6.89E-03	7.31	9.88E-03	6.84	8.50E-03	7.11	7.35E-03	7.48
10 h	1.14E-02	6.97	6.96E-03	8.47	9.99E-03	7.99	8.59E-03	8.24	7.43E-03	8.69
12 h	1.16E-02	8.15	7.06E-03	9.92	1.01E-02	9.41	8.71E-03	9.67	7.54E-03	10.19
15 h	1.18E-02	10.47	7.24E-03	12.80	1.04E-02	12.21	8.93E-03	12.46	7.74E-03	13.14
OLINDA/EXM 2.0	_									
Labelled with GP-99mTc	El	D_t	E	D_t	El	D_t	El	D_t	E	D_t
	1.07	E-02	5.24	E-03	8.641	E-03	8.091	E-03	7.04	E-03
Labelled with CP-99m Tc	ED_t	DI (%)	ED_t	DI (%)	ED_t	DI (%)	ED_t	DI (%)	ED_t	DI (%)
Time after EOB										
6 h	1.11E-02	4.40	5.57E-03	6.41	9.12E-03	5.57	8.51E-03	5.19	7.42E-03	5.42
8 h	1.12E-02	4.92	5.62E-03	7.30	9.19E-03	6.38	8.57E-03	5.90	7.47E-03	6.18
10 h	1.13E-02	5.64	5.68E-03	8.47	9.28E-03	7.43	8.64E-03	6.83	7.54E-03	7.15
12 h	1.14E-02	6.56	5.76E-03	9.94	9.40E-03	8.74	8.74E-03	7.98	7.63E-03	8.37
15 h	1.16E-02	8.37	5.91E-03	12.85	9.62E-03	11.33	8.92E-03	10.25	7.79E-03	10.74

Figure 1 shows the contribution (in percentage) of each Tc-radionuclide to the radiopharmaceutical's effective dose at 3 different time points after EOB (8, 10 and 12h), calculated with both versions of OLINDA/EXM. Data for MIBI are reported only for stress condition because, in this case, the DI is higher than in rest conditions. As expected, ⁹⁶Tc is the radioisotope

with greatest contribution to the effective dose, followed by ⁹⁵Tc and ⁹⁴Tc; the contribution of other Tc-radioisotopes is quite low, being at least 3 orders of magnitude smaller than the contribution of ^{99m}Tc.



Figure 1. Contribution (in percentage) of each Tc-radionuclide to the effective dose of radiopharmaceuticals at 3 different time points after EOB (8, 10 and 12h) calculated with both versions of OLINDA/EXM.

4. DISCUSSION

The number of disintegrations in the main source organs calculated for each Tc-radioisotope after administration of Tc-pertechnetate, Tc-HEDP and Tc-MIBI are in general agreement with the data reported by Hou *et al*¹⁰. However, some differences were found. The authors did not consider 3 important source organs (salivary gland, stomach content and upper large intestine walls) reported in the ICRP biokinetic model to calculate the dose produced by Tc-pertechnetate. They also underestimated the number of disintegrations for the Tc-MIBI model since liver delay and gallbladder uptake were not considered for the calculated with the Tc-HEDP biokinetic model, because their calculated values range from half to 9 times less the values reported in table 3. It is not possible to determine whether these differences produced a considerable effect on the calculated total effective dose, by comparison with the results of the present work, because Hou *et al* did not report the total effective dose values, but rather, the percent of difference between cyclotron and reactor-produced technetium¹⁰.

In general, low differences between the total effective doses calculated with both versions of the OLINDA software were expected, because version 2.0 uses more anatomically-accurate NURBS phantoms. Furthermore, some tissue-weighting factor values differ from the version 1.1 (ICRP 60) to version 2.0 (ICRP 103), resulting in an increment in the tissue equivalent dose, as in the case of Tc-MIBI, where the rise of muscle-weighting factor from 0.005 to 0.01 produced higher total effective doses using OLINDA/EXM 2.0 (see Table 6). However, the DI remained within the 10 % limit from 6 to 12 h after EOB for all of the radiopharmaceuticals labelled with the CP-^{99m}Tc studied, when the more accurate version of Olinda/EXM 2.0 was used.

Table 6 shows that the percent of increase in dose is different for each radiopharmaceutical because the biological half-life in each source organ differs among compounds. Therefore, dose calculation must be done for each radiopharmaceutical in order to determine the range of time after the EOB in which the Tc-radiopharmaceutical can be administered to remain within the 10 % limit of DI. The greatest time dependence in the DI contribution from Tc-radionuclides in the studied time intervals was found for ^{94m}Tc and ^{93m}Tc, since their impact on the dose decreases quickly. Although these Tc-radionuclides present the highest concentration in the CP-^{99m}Tc solution at the EOB, their contribution to the dose 6 h later will be minimal due to their short half-life (see Table 2). Radionuclides such as ^{97m}Tc, ⁹⁶Tc, and ^{95m}Tc, having a longer half-life, should instead increase their dose contribution over time; however, due to the small concentrations in the reaction mixture, the increment is negligible (see fig 1).

Table 7 compares the increase of the total effective doses for three Tc-radiopharmaceuticals calculated from our experimental results, with the theoretical calculations from Hou *et al* ¹⁰ using the same Isoflex Mo target enriched to 99.05%, irradiated for 3h with a proton beam of 16 MeV. Dose contribution of Tc-impurities calculated by Hou also using OLINDA/EXM 1.1 software, is higher due to the longer irradiation time. Twelve hours after EOB, the increase in effective dose is greater than 10% for Tc-pertechnetate and Tc-MIBI, only Tc-HEDP DI remained under 10%. However, this is probably a consequence of the underestimation of the number of Tc-HEDP disintegrations in the whole body. Selivanova *et al* reported a 7.58 % increase in Tc-pertechnetate effective dose, assuming an irradiation at 24 MeV for two hours and injection time 6 hours after EOB, which is superior to the 5.30% obtained in this work for the same Mo target and injection time. This increment is the consequence of a greater production of ⁹⁴Tc, ⁹⁵Tc and ⁹⁶Tc-radionuclides at 24 MeV.

⁹⁶Tc was the radioisotope with the highest contribution to the effective dose and is potentially detrimental to the image quality, due to its energetic γ rays ⁸. Thus, the best CP-^{99m}Tc method should be one that produces the least amount of ⁹⁶Tc. Using a proton beam of 15.7 MeV and an enriched Isoflex Mo target (99.05 %) it was found that just 0.0267% of the total activity is related to ⁹⁶Tc (see Table 1). Selivanova *et al*²⁶ reported a ⁹⁶Tc concentration in the Tc-final product of 0.0695%, 0.1478% and 0.1614% using the same type of Mo target and proton beams of 20, 22 and 24 MeV, respectively. These results demonstrate that the low-energy conventional medical

cyclotron is preferable, since it produces 99m Tc with less concentration of 96 Tc than medium-energy cyclotrons, as preliminary reported in the theoretical study by Esposito *et al*⁶.

Table 7. Comparison of the increase of total effective dose calculated in this study (15.7 MeV and 1h irradiation) for three Tc-radiopharmaceuticals with the values reported in the literature by Hou *et al* (16 MeV and 3 h irradiation).

Irradiation data	Ra	diopharmaceutic	als
15.7 MeV (1 h)	Tc-pertechnetate	Tc-HEDP	Tc-MIBI rest
Time after EOB			
8 h	5.99 %	7.25 %	7.05 %
12 h	8.07 %	9.82 %	9.57 %
16 MeV (3 h)			
Time after EOB			
8 h	8.28 %	7.49 %	9.23 %
12 h	10.99 %	9.59 %	12.22 %

To calculate the maximum patient radiation dose increase after the administration of the CP-^{99m}Tcradiopharmaceuticals, the total effective dose values were multiplied by the maximum activity used in clinical practice for each type of diagnostic radiopharmaceutical. Table 8 shows the maximum radiation dose that a patient could receive after the administration of radiopharmaceuticals labelled with GP-^{99m}Tc and CP-^{99m}Tc, assuming the Tc impurity levels in the limits established by the European Pharmacopoeia (see table 2) and the Tc impurities obtained in our experimental studies. Analysing these values, it is clear that the increment of the dose due to the impurities is minimal. In all cases, patient radiation dose is lower than the maximum dose accepted for an imaging study (10 mSv) and also, than the dose received by patients after the administration of 370 MBq of ¹⁸FDG (7.03 mSv)¹⁷.

Table 8. Maximum patient radiation dose after the administration of radiopharmaceuticals labelled

 with technetium from different sources.

	Patient radiation dose after ^{99m} Tc-radiopharmaceutical administration (ntion (mSy	V)			
^{99m} Tc Source	Tc-pertechnetate (250 MBq)		Tc-pertechnetate (250 MBq)		ate Tc-HEDP (1000 MBq)		<i>Tc-HMPAO</i> (185 MBq)		<i>Tc-MIBI rest</i> (740 MBq)		<i>Tc-MIBI stress</i> (740 MBq)	
				C	LINDA/E	XM versio	n					
GP - ^{99m}Tc	1.1	2.0	1.1	2.0	1.1	2.0	1.1	2.0	1.1	2.0		
	2.68	2.68	6.42	5.24	1.71	1.60	5.88	5.99	5.06	5.21		
CP- ^{99m} Tc with th	e pharma	conoeial li	mits									
	2.79	2.76	6.77	5.52	1.80	1.68	6.17	6.24	5.33	5.43		
CP - ^{99m}Tc												
Time after EOB (h)												
6	2.82	2.78	6.83	5.57	1.81	1.69	6.24	6.30	5.39	5.49		
8	2.84	2.80	6.89	5.62	1.83	1.70	6.29	6.34	5.44	5.53		
10	2.86	2.82	6.96	5.68	1.85	1.72	6.36	6.39	5.50	5.58		
12	2.89	2.84	7.06	5.76	1.87	1.74	6.44	6.46	5.58	5.64		

CONCLUSIONS

The percent of increase in dose is different for each radiopharmaceutical because the biological half-life in each source organ differs among compounds. Therefore, to determine the range of time after EOB in which the Tc-solution can be used, it is necessary to perform dosimetric studies with the most representative radiopharmaceuticals, not just with Tc-pertechnetate.

Comparing the results obtained using the data of our target irradiation experiments with the data reported by other research groups, it was concluded that the CP-^{99m}Tc, produced at lower energy (16 MeV) for shorter irradiation time (60 min), shows some advantages. Because, although the quantity of ^{99m}Tc produced in this way is lower, its radionuclidic purity is higher, and the solution can be used for a longer period of time without increasing the patient absorbed dose substantially.

The patient radiation absorbed dose increase by the contribution of Tc contaminant activity present in the cyclotron-^{99m}Tc solution, produced by the beam irradiation conditions reported in this work, is relatively low. However, its impact should not be underestimated. Based on this data, it was concluded that although the concentrations of two radionuclides ⁹⁴Tc and ⁹⁵Tc in the CP-^{99m}Tc, is outside the limits established by the European Pharmacopoeia for all the time points studied, radiopharmaceuticals labelled with this CP-^{99m}Tc solution administered from 6 to 12 h after the end of bombardment could be used in routine nuclear medicine diagnostic studies because, the dose increment remained within the 10% limit.

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6. REFERENCES

- 1. Arano Y. Recent advances in Tc-99m radio pharmaceuticals. *Annals of Nuclear Medicine*. 2002;16(2):79-93.
- 2. Pupillo G, Esposito J, Haddad F, Michel N, Gambaccini M. Accelerator-based production of Mo-99: a comparison between the Mo-100(p,x) and Zr-96(alpha,n) reactions. *Journal of Radioanalytical and Nuclear Chemistry*. 2015;305(1):73-78.
- 3. Pupillo G, Esposito J, Gambaccini M, Haddad F, Michel N. Experimental cross section evaluation for innovative Mo-99 production via the (alpha,n) reaction on Zr-96 target. *Journal of Radioanalytical and Nuclear Chemistry*. 2014;302(2):911-917.
- 4. Pillai MRA, Dash A, Knapp FF, Jr. Sustained Availability of Tc-99m: Possible Paths Forward. *Journal of Nuclear Medicine*. 2013;54(2):313-323.
- 5. Boschi A, Martini P, Pasquali M, Uccelli L. Recent achievements in Tc-99m radiopharmaceutical direct production by medical cyclotrons. *Drug Dev Ind Pharm.* 2017;43(9):1402-1412.
- 6. Esposito J, Vecchi G, Pupillo G, et al. Evaluation of Mo-99 and Tc-99m Productions Based on a High-Performance Cyclotron. *Science and Technology of Nuclear Installations*. 2013. doi: 10.1155/2013/972381:14.
- 7. Martini P, Boschi A, Cicoria G, et al. In-house cyclotron production of high-purity Tc-99m and Tc-99m radiopharmaceuticals. *Applied Radiation and Isotopes*. 2018;139:325-331.
- 8. Uzunov NM, Melendez-Alafort L, Bello M, et al. Radioisotopic purity and imaging properties of cyclotron-produced Tc-99m using direct Mo-100(p,2n) reaction. *Physics in Medicine and Biology*. 2018;63(18).
- 9. Martini P, Boschi A, Cicoria G, et al. A solvent-extraction module for cyclotron production of high-purity technetium-99m. *Applied Radiation and Isotopes*. 2016;118:302-307.

- 10. Hou X, Celler A, Grimes J, Benard F, Ruth T. Theoretical dosimetry estimations for radioisotopes produced by proton-induced reactions on natural and enriched molybdenum targets. *Physics in Medicine and Biology*. 2012;57(6):1499-1515.
- 11. Lebeda O, van Lier EJ, Stursa J, Ralis J, Zyuzin A. Assessment of radionuclidic impurities in cyclotron produced Tc-99m. *Nucl Med Biol.* 2012;39(8):1286-1291.
- 12. Galea R, Wells RG, Ross CK, et al. A comparison of rat SPECT images obtained using Tc-99m derived from Mo-99 produced by an electron accelerator with that from a reactor. *Physics in Medicine and Biology*. 2013;58(9):2737-2750.
- 13. Uccelli L, Boschi A, Pasquali M, et al. Influence of the Generator in-Growth Time on the Final Radiochemical Purity and Stability of Tc-99m Radiopharmaceuticals. *Science and Technology of Nuclear Installations*. 2013. doi: 10.1155/2013/379283.
- 14. Hou X, Tanguay J, Vuckovic M, et al. Imaging study of using radiopharmaceuticals labeled with cyclotron-produced Tc-99m. *Physics in Medicine and Biology*. 2016;61(23):8199-8213.
- Selivanova SV, Lavallee E, Senta H, et al. Clinical Trial with Sodium Tc-99m-Pertechnetate Produced by a Medium-Energy Cyclotron: Biodistribution and Safety Assessment in Patients with Abnormal Thyroid Function. *Journal of Nuclear Medicine*. 2017;58(5):791-798.
- 16. European Pharmacopoeia 9.3 Sodium pertechnetate (99mTc) injection (accelerator produced). (published online at <u>http://online6.edqm.eu/ep903/)</u>. *Monograph No 2891*. 2018;01:4801–4803.
- 17. ICRP. Radiation Dose to Patients from Radiopharmaceuticals. *CRP Publication 53*. 1988;Ann. ICRP 18:(1-4).
- 18. ICRP. Radiation dose to patients from radiopharmaceuticals (addendum 2 to ICRP publication 53). *ICRP Publication 80*. 1998;Ann. ICRP 28:(3).
- 19. ICRP. Radiation dose to patients from radiopharmaceuticals (addendum 3 to ICRP publication 53). *ICRP Publication 106*. 2008;Ann. ICRP 38((1-2)):(1-2).
- 20. Stabin MG, Sparks RB, Crowe E. OLINDA/EXM: The second-generation personal computer software for internal dose assessment in nuclear medicine. *Journal of Nuclear Medicine*. 2005;46(6):1023-1027.
- 21. ICRP. Limits for Intakes of Radionuclides by Workers. *ICRP Publication 30 (Part 1)*. 1979;Ann. ICRP 2:(3-4).
- 22. ICRP. 1990 Recommendations of the International Commission on Radiological Protection. *ICRP Publication 60* 1991;Ann. ICRP 21:(1–3).
- 23. ICRP. Basic Anatomical and Physiological Data for Use in Radiological Protection Reference Values. *ICRP Publication 89.* 2002;Ann. ICRP 32:(3-4).
- 24. ICRP. The 2007 Recommendations of the International Commission on Radiological Protection. *ICRP Publication 103*. 2007;Ann. ICRP 37((2-4)):(2-4).
- 25. Richter WS, Cordes M, Calder D, Eichstaedt H, Felix R. Washout and redistribution between immediate and 2 hour myocardial images using 99mTc-sestamibi *European Journal of Nuclear Medicine*. 1995;22(1):49-55.
- 26. Selivanova SV, Lavallee E, Senta H, et al. Radioisotopic Purity of Sodium Pertechnetate Tc-99m Produced with a Medium-Energy Cyclotron: Implications for Internal Radiation Dose, Image Quality, and Release Specifications. *Journal of Nuclear Medicine*. 2015;56(10):1600-1608.