

COMMENTARY

RECOMBINANT PROBIOTICS AND THEIR POTENTIAL IN HUMAN HEALTH

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ABSTRACT: *The use of probiotics in the form of whole bacteria to achieve health benefits in humans is evolving by the development of recombinant probiotic bacteria as carriers of specific genes achieving a probiotic effect. Studies have demonstrated that bacteria carrying either the gene for IL-10 or for trefoil factors when given in an animal model or as part of a human trial can ameliorate inflammatory conditions of the colon. Such an approach has the advantages of a long-term delivery, the potential for fewer side effects and utility in many other conditions including other autoimmune diseases, dental caries, candidiasis and allergies. Other health benefits are also under investigation in relation to altering the fatty acid composition of adipose tissue by colonizing the intestine with a bacterium carrying the gene involved in fatty acid metabolism. Not only is the lean body mass advantageously affected but the enzyme is active in inducing apoptosis in cancer cells. Benefits may also be obtained not only by adding a gene but deletion of a gene from a bacterium which modulates the production of pro-inflammatory cytokines or may make the probiotic bacterium act as a more effective delivery system by enhancing colonization.*

KEY WORDS: Probiotic Bacteria, Recombinant Probiotics, Recombination

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The effects of probiotic bacteria on human health have been investigated mainly during the last two decades (Seegers, 2002; Wells and Mercenier, 2003; Hanniffy et al., 2004),

but the potential for future developments in this field remains impressive. As a corollary of the investigations on probiotics, two very promising areas of study have emerged in recent years: i) bacterial derived molecules having probiotic properties (Caselli et al., 2011) and ii) the generation of recombinant probiotic strains able to express molecules with favourable pharmacological properties (Blaise Corthesy et al., 2007). In the present article we will focus our attention on this last issue. We will take into consideration findings related to bacteria stably transfected with genes coding for molecules able to prevent or cure different pathological conditions. Colonizing (e.g. *Streptococcus gondii*, *Lactobacillus plantarum*, *Lactobacillus paracasei*, *Lactobacillus casei*, *Lactobacillus acidophilus*) as well as non-colonizing (e.g. *Lactobacillus lactis*) bacterial species have been investigated both as live vaccine vehicles (acting as carriers for protective antigens) and as active producers of molecules with known pharmacological properties.

In respect to the use of these microorganisms as carriers for antigens, the most complete studies have been carried out with the 50kDa carboxyl-terminal fragment of tetanus toxin (TTFC) (Grangette et al., 2002); this approach has now been extended to additional antigens e.g. the B subunit of cholera toxin (Seegers, 2002; Wells and Mercenier, 2003; Hanniffy et al., 2004).

On the other hand, transfected bacteria have been used to deliver cytokines, but this technique was recently used to investigate other biological properties. Steidler chose to construct recombinant *Lactobacillus lactis* strains secreting murine IL-10 (Steidler et al., 2000). These authors demonstrated that these recombinant strains were able to prevent and treat inflammation in two murine models of colitis. Significantly, these same effects were obtained with much lower doses of IL-10 than those

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required when IL-10 itself was used as a drug. The same authors further constructed a safe (no antibiotic resistance markers and a chromosomally integrated transgenic) strain of *Lactobacillus lactis* secreting IL-10 of human origin (Steidler et al., 2003). Authorization by a local ethical committee to carry out a phase I clinical study on voluntary patients has been obtained in the Netherlands (Braat et al., 2006). In this study, Crohn's disease patients were treated with recombinant *Lactobacillus lactis* (LL-THY 12) in which the thymidylate synthase gene was replaced with a synthetic sequence encoding mature human IL-10. The oral administration of this strain was safe and a decrease in disease activity was observed. The authors concluded that the use of genetically modified bacteria for mucosal delivery of therapeutic proteins is a feasible strategy in human beings. This strategy avoids systemic side effects and appears suitable as maintenance treatment for chronic intestinal diseases. Novel therapeutic strategies for acute and chronic colitis based on recombinant probiotics were also assessed by the generation and in vivo evaluation of *Lactobacillus lactis* strains secreting bioactive murine trefoil factors (TFF) (Van den Broucke et al., 2004). The authors demonstrated that intra gastric administration of this bacterial strain, but not of purified TFF, led to prevention and healing in the acute dextran sodium sulfate (DSS)-induced murine model of colitis, and were similarly effective in reducing established chronic DSS colitis. It has also to be remembered that production and mucosal delivery of different bioactive molecules such as allergens, digestive enzymes and single-chain Fw antibodies have been achieved using lactic acid bacteria (3). Targeted diseases included vaginal candidiosis (Beninati et al., 2000), dental caries (Ktuger et al., 2002), allergies (Kruisselbrink et al., 2001; Chatel et al., 2001; Repa et al., 2003; Daniel et al., 2006), autoimmune diseases (Maassen et al., 1999; Madsen et al., 1999), HPV-induced tumors (Bermudez-Humaran et al., 2005) and pancreatic insufficiency (Drouault et al., 2002). More recently, Rosberg-Cody (Rosberg-Cody et al., 2010) investigated whether a recombinant strain of *Lactobacillus paracasei*, previously isolated from the human gastrointestinal tract, expressing conjugated linoleic acid (CLA) isomerase from *Propionibacterium acnes*, could influence the fatty-acid composition of different tissues in the mouse. Ingestion of the *Lactobacillus paracasei* strain expressing CLA isomerase was associated with a 4-fold increase ($p > 0.001$) in t10c12 CLA in adipose tissues of the mice when compared with animals that received the non CLA producing isogenics strain. These data show that a single gene encoding CLA isomerase expressed by an intestinal bacterium can influence the fatty-acid composition of the host adipose tissue. This t10c12 CLA isomer is also associated with decreased body fat and increased lean body mass in various animal species (Park et al., 1997; Cherian et al., 2005; Yamasaki et al., 2003; Navarro et al., 2006; Ostrowska et al., 1999) and, to some extents, human beings (Blankson et al., 2000; Mougios et al., 2001; Riserus et al., 2001; Smedman and Vessby, 2001; Thom et al., 2001). It is also well known that t10c12 CLA isomer is the most potent isomer in terms of potential to prevent cell proliferation and induce apoptosis in cancer cells

(Cho et al., 2005; Cho et al., 2006; Kim et al., 2002a; Lee et al., 2006a; Ochoa et al., 2004) notably, when the microbial derived t10c12 CLA was incubated with SW480 colon cancer cells for 5 days, cell viability was decreased by 92% (Rosberg-Cody et al., 2007), and it is possible that a CLA-producing probiotic will be able to keep colon cancer cells in check. Although commensal *Bifidobacterium* and *Lactobacillus* species from the gastro-intestinal tract have been shown to produce CLA in vitro (Barrett et al., 2007; Coakley et al., 2003; Rosberg-Cody et al., 2004), the majority of these studies have demonstrated the production of c9t11 CLA from linoleic acid, while only a few bacteria such as *Propionibacterium acnes* (Verhulst et al., 1987), the rumen bacterium *Megasphaera elsodenii* (Kim et al., 2002b), and the human derived *Lactobacillus rhamnosus* PL60 and *Lactobacillus plantarum* PL 62 (Lee et al., 2006b; Lee et al., 2007) have been reported to produce t10c12 CLA. Modulation of fatty acid production by bacteria may represent a very important probiotic activity and recombinant probiotics may become useful for this in the near future.

Recombinant probiotics may be linked not only to the addition of one or more genes but also to the deletion of one or more genes. In fact, to study the molecular mechanisms involved in the induction and repression of intestinal inflammation, Mohamadzadeh (Mohamadzadeh et al., 2011) has recently deleted the phosphoglycerol transferase gene that plays a key role in lipoteichoic acid (LTA) biosynthesis in *Lactobacillus acidophilus* NCK 56.

The results of these authors show that the *Lactobacillus acidophilus* LTA⁻ not only down regulated IL 12 and TNF alfa, which are known pro-inflammatory cytokines, but also significantly enhanced IL-10 production by dendritic cells (DC) and controlled the regulation of co-stimulatory DC functions, resulting in their inability to induce CD 4⁺ T cell activation. The treatment of mice with *L. acidophilus* LTA⁻, compared with *L. acidophilus* LTA⁺, significantly counteracted DSS-induced colitis. These authors concluded that directed alteration of cell-surface components of *L. acidophilus* represents a potential new strategy for the treatment of inflammatory intestinal disorders.

Moreover, efforts have been devoted to improve the efficacy of probiotic bacteria as delivery systems; in this context cell wall mutants of *Lactobacillus plantarum* and *Lactobacillus lactis* defective in alanine racemase (*alr* gene) were constructed (Palumbo et al., 2004; Steen et al., 2005): each of these mutants behaved as a substantially improved antigen delivery system compared with its wild-type counterpart (Grangette et al., 2004). The potency of the *Lactobacillus plantarum* Alr⁻ mutant was further confirmed using a weak immunogenic, such as *Helicobacter pylori* urease B, as a protective antigen; a significant reduction of the *Helicobacter pylori* load in the mouse stomach was achieved after immunization with the recombinant mutant *Lactobacillus plantarum* strain in contrast to results obtained with its wild-type counterpart (Blaise-Corthesy et al., 2005).

Any gene coding for an active molecule, potentially useful for human health, may be used to generate recombinant probiotic bacteria; in this context, an impressive number of

options are available to be investigated in *in vitro* and *in vivo* studies. It is worthy of note, however, that several gene products need glycosylation, phosphorylation or other more complex chemical changes; these may require the enzymatic machinery of eukaryotic cells. Thus, although current available genomic information should greatly facilitate the generation of useful recombinant probiotics, several technical issues and biologically limiting factors have to be taken attentively in consideration. In any case, the use of rapidly evolving genomic technology will surely help to evolve this intriguing and fascinating field and we can expect that from the present pioneering status we will soon progress to the generation of innovative therapeutic tools.

REFERENCE

- Barrett, E., Ross, R. P., Fitzgerald, G. F., Stanton, C. (2007) Rapid screening method for analyzing the conjugated linoleic acid production capabilities of bacterial cultures. *Applied and Environmental Microbiology* **73**, 2333–2337.
- Bermudez-Humaran, L.G., Cortes-Perez, N.G., Lefevre, F., Guimaraes, V., Rabot, S., Alcocer-Gonzalez, J.M., Gratadoux, J.J., Rodriguez-Padilla, C., Tamez-Guerra, R.S., Corthier, G., Gruss, A., Langella, P. (2005) A novel mucosal vaccine based on live lactococci expressing E7 antigen and IL-12 induces systemic and mucosal immune responses and protects mice against human papillomavirus type 16-induced tumors. *Journal of Immunology* **175**, 297–7302.
- Beninati, C., Oggioni, M.R., Boccanera, M., Spinoza, M.R., Maggi, T., Conti, S., Magliani, W., De Bernardis, F., Teti, G., Cassone, A., Pozzi, G., Polonelli, L. (2000) Therapy of mucosal candidiasis by expression of an anti-idiotypic in human commensal bacteria. *Nature Biotechnology* **18**, 1060–1064.
- Blankson, H., Stakkestad, J. A., Fagertun, H., Thom, E., Wadstein, J., Gudmundsen, O. (2000) Conjugated linoleic acid reduces body fat mass in overweight and obese humans. *Journal of Nutrition* **130**, 2943–2948.
- Braat, H., Rottier, P., Hommes, D.W., Huyghebaert, N., Remaut, E., Remon, J.P., van Deventer, S.J., Neiryck, S., Peppelenbosch, M.P., Steidler, L. (2006) A phase I trial with transgenic bacteria expressing interleukin-10 in Crohn's disease. *Clinical Gastroenterology and Hepatology* **4**, 754–759.
- Caselli, M., Vaira, G., Calò, G., Holton, J., Vaira, D. (2011) Structural bacterial molecules as potential candidates for an evolution of the classical concept of probiotics. *Advances in Nutrition* **2**, 372–376
- Chatel, J.M., Langella, P., Adel-Patient, K., Commissaire, J., Wal, J.M., Corthier, G. (2001) Induction of mucosal immune response after intranasal or oral inoculation of mice with *Lactococcus lactis* producing bovine beta-lacto globulin. *Clinical and Diagnostic Laboratory Immunology* **8**, 545–851.
- Cherian, G., Wu, A., Goeger, P. M. (2005) Maternal dietary conjugated linoleic acid alters hepatic triacylglycerol and tissue fatty acids in hatched chicks. *Lipids* **40**, 131–136.
- Cho, H. J., Kim, W. K., Jung, J. I., Kim, E. J., Lim, S. S., Kwon, D. Y., Park, J. H. (2005) Trans-10,cis-12, not cis-9,trans-11, conjugated linoleic acid decreases ErbB3 expression in HT-29 human colon cancer cells. *World Journal of Gastroenterology* **11**, 5142–5150.
- Cho, H. J., Kim, E. J., Lim, S. S., Kim, M. K., Sung, M. K., Kim, J. S., Park, J. H. (2006) Trans-10,cis-12, not cis-9,trans-11, conjugated linoleic acid inhibits G1-S progression in HT-29 human colon cancer cells. *Journal of Nutrition* **136**, 893–898.
- Coakley, M., Ross, R. P., Nordgren, M., Fitzgerald, G., Devery, R., Stanton, C. (2003) Conjugated linoleic acid biosynthesis by human-derived *Bifidobacterium* species. *Journal of Applied Microbiology* **94**, 138–145.
- Corthèsy, B., Gaskins, H., Mercenier, A. (2007) Cross-Talk between Probiotic Bacteria and the Host Immune System. *Journal of Nutrition* **37**, 7815–7905.
- Corthesy, B., Boris, S., Isler, P., Grangette, C., Mercenier, A. (2005) Oral immunization of mice with lactic acid bacteria producing *Helicobacter pylori* urease B subunit partially protects against challenge with *Helicobacter felis*. *Journal of Infectious Diseases* **192**, 1441–1449.
- Daniel, C., Repa, A., Wild, C., Pollack, A., Pot, B., Breiteneder, H., Wiedermann, U., Mercenier, A. (2006) Modulation of allergic immune responses by mucosal application of recombinant lactic acid bacteria producing the major birch pollen allergen Bet v1. *Allergy* **61**, 812–819.
- Drouault, S., Juste, C., Marteau, P., Renault, P., Corthier, G. (2002) Oral treatment with *Lactococcus lactis* expressing *Staphylococcus hyicus* lipase enhances lipid digestion in pigs with induced pancreatic insufficiency. *Applied and Environmental Microbiology* **68**, 3166–3168.
- Grangette, C., Muller-Alouf, H., Geoffroy, M., Goudercourt, D., Turneer, M., Mercenier, A. (2002) Protection against tetanus toxin after intragastric administration of two recombinant lactic acid bacteria: impact of strain viability and in vivo persistence. *Vaccine* **20**, 3304–3309.
- Grangette, C., Muller-Alouf, H., Hols, P., Goudercourt, D., Delcour, J., Turneer, M., Mercenier, A. (2004) Enhanced mucosal delivery of antigen with cell wall mutants of lactic acid bacteria. *Infection and Immunity* **72**, 2731–2737.
- Hanniffy, S., Wiedermann, U., Repa, A., Mercenier, A., Daniel,

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- C., Fioramonti, J., Tlaskolova, H., Kozakova, H., Israelsen, H., Madsen, S., Vrang, A., Hols, P., Delcour, J., Bron, P., Kleerebezem, W., Wells, J. (2004) Potential and opportunities for use of recombinant lactic acid bacteria in human health. *Advances in Applied Microbiology* **56**, 1-64.
- Kim, E. J., Holthuisen, P. E., Park, H. S., Ha, Y. L., Jung, K. C. & Park, J. H. (2002a) Trans-10, cis-12-conjugated linoleic acid inhibits Caco-2 colon cancer cell growth. *American Journal of Physiology Gastrointestinal and Liver Physiology* **283**, G357-G367.
- Kim, Y. J., Liu, R. H., Rychlik, J. L., Russell, J. B. (2002b) The enrichment of a ruminal bacterium (*Megasphaera elsdenii* YJ-4) that produces the trans-10, cis-12 isomer of conjugated linoleic acid. *Journal of Applied Microbiology* **92**, 976-982.
- Kruger, C., Hu, Y., Pan, Q., Marcotte, H., Hultberg, A., Delwar, D., van Dalen, P.J., Pouwels, P.H., Leer, R.J., Kelly, C.G., van Dollenweerd, C., Ma, J.K., Hammarstrom, L. (2002) In situ delivery of passive immunity by lactobacilli producing single-chain antibodies. *Nature Biotechnology* **20**, 702-706.
- Kruisselbrink, A., den Bak-Glashouwer, M.J., Havenith, C.E., Thole, J.E., Janssen R. (2001) Recombinant *Lactobacillus plantarum* inhibits house dust mite-specific T-cell responses. *Clinical and Experimental Immunology* **126**, 2-8.
- Lee, S. H., Yamaguschi, K., Kim, J. S., Eling, T. E., Safe, S., Park, Y., Baek, S. J. (2006a) Conjugated linoleic acid stimulates an anti-tumorigenic protein NAG-1 in an isomer specific manner. *Carcinogenesis*. **27**, 972-981.
- Lee, H. Y., Park, J. H., Seok, S. H., Baek, M. W., Kim, D. J., Lee, K. E., Paek, K. S., Lee, Y., Park, J. H. (2006b) Human originated bacteria, *Lactobacillus rhamnosus* PL60, produce conjugated linoleic acid and show anti-obesity effects in diet-induced obese mice. *Biochimica et Biophysica Acta*. **1761**, 736-744.
- Lee, K., Paek, K., Lee, H. Y., Park, J. H. & Lee, Y. (2007) Anti-obesity effect of trans-10, cis-12-conjugated linoleic acid-producing *Lactobacillus plantarum* PL62 on diet-induced obese mice. *Journal of Applied Microbiology* **103**, 1140-1146.
- Maassen, C.B., Laman, J.D., den Bak-Glashouwer, M.J., Tielen, F.J., van Holten-Neelen, J.C., Hoogteijling, L., Antonissen, C., Leer, R.J., Pouwels, P.H., Boesma, W.J., Shaw, D.M. (1999) Instruments for oral disease intervention strategies: recombinant *Lactobacillus casei* expressing tetanus toxin fragment C for vaccination or myelin proteins for oral tolerance induction in multiple sclerosis. *Vaccine*. **17**, 2117-2128.
- Madsen, K.L., Doyle, J.S., Jewel, L.D., Tavemini, M.M., Fedorak, R.N. (1999) *Lactobacillus* species prevents colitis in interleukin-10 gene-deficient mice. *Gastroenterology*. **116**, 1107-1114.
- Mohamadzadeh, M., Pfeiler, E.A., Brown, J.B., Zadeh, M., gramarossa, M., Managlia, E., Bere, P., Sarrai, B., Khan, M.W., Pakanati, K.C., Ansari, M.J., O'Flaherty, S., Barrett, T., Klaenhammer, T.R. (2011) Regulation of induced colonic inflammation by *Lactobacillus acidophilus* deficient in lipoteichoic acid. *Proceedings of the National Academy of Sciences USA* **108** (Suppl. 1), 4623-4630
- Mougiou, V., Matsakas, A., Petridou, A., Ring, S., Sagredos, A., Melissopoulou, A., Tsigilis, N., Nikolaidis, M. (2001) Effect of supplementation with conjugated linoleic acid on human serum lipids and body fat. *Journal of Nutritional Biochemistry* **12**, 585-594.
- Navarro, V., Miranda, J., Churrua, I., Fernández-Quintela, A., Rodríguez, V. M., Portillo, M. P. (2006) Effects of trans-10, cis-12 conjugated linoleic acid on body fat and serum lipids in young and adult hamsters. *Journal of Physiology and Biochemistry*. **62**, 81-87.
- Ochoa, J. J., Farquharson, A. J., Grant, I., Moffat, L. E., Heys, S. D., Wahle, K. W. (2004) Conjugated linoleic acids (CLAs) decrease prostate cancer cell proliferation: different molecular mechanisms for cis-9, trans-11 and trans-12, cis-12 isomers. *Carcinogenesis*. **25**, 1185-1191.
- Ostrowska, E., Muralitharan, M., Cross, R. F., Bauman, D. E., Dunshea, F. R. (1999) Dietary conjugated linoleic acids increase lean tissue and decrease fat deposition in growing pigs. *Journal of Nutrition* **129**, 2037-2042.
- Palumbo, E., Favier, C.F., Deghorain, M., Cocconcelli, P.S., Grangette, C., Mercenier, A., Vaughan, E.E., Hols, P. (2004) Knockout of the alanine racemase gene in *Lactobacillus plantarum* results in septation defects and cell wall perforation. *FEMS Microbiology Letters* **233**, 131-138.
- Park, Y., Albright, K. L., Liu, W., Storkson, J. M., Cook, M. E., Pariza, M. W. (1997) Effect of conjugated linoleic acid on body composition in mice. *Lipids*. **32**, 853-858.
- Repa, A., Grangette, C., Daniel, C., Hochreiter, R., Hoffmann-Sommergruber, K., Thalhamer, J., Kraft, D., Breiteneder, H., Mercenier, A., Wiedermann, U. (2003) Mucosal co-application of lactic acid bacteria and allergen induces counter-regulatory immune responses in a murine model of birch pollen allergy. *Vaccine*. **22**, 87-95.
- Risérus, U., Berglund, L., Vessby, B. (2001) Conjugated linoleic acid (CLA) reduced abdominal adipose tissue in obese middle-aged men with signs of the metabolic syndrome: a randomized controlled trial. *International Journal of Obesity and Related*

Metabolic Disorders **25**, 1129–1135.

Rosberg-Cody, E., Stanton, C., O'Mahony, L., Wall, R., Shanahan, F., Quigley, E.M., Fitzgerald, G.F., Ross, R.P. (2011) Recombinant lactobacilli expressing linoleic acid isomerase can modulate the fatty acid composition of host adipose tissue in mice. *Microbiology* **157**, 609–615.

Rosberg-Cody, E., Johnson, M. C., Fitzgerald, G. F., Ross, R. P. & Stanton C.(2007) Heterologous expression of linoleic acid isomerase from *Propionibacterium acnes* and ant proliferative activity of recombinant trans-10, cis-12 conjugated linoleic acid. *Microbiology*. **153**, 2483–2490.

Rosberg-Cody, E., Ross, R. P., Hussey, S., Ryan, C. A., Murphy, B. P., Fitzgerald, G. F., Devery, R., Stanton, C. (2004) Mining the microbiota of the neonatal gastrointestinal tract for CLA-producing bifidobacteria. *Applied and Environmental Microbiology* **70**, 4635–4641.

Seegers JF. (2002) Lactobacilli as live vaccine delivery vectors: progress and prospects. *Trends in Biotechnology* **20**, 508–515.

Smedman, A., Vessby, B. (2001) Conjugated linoleic acid supplementation in humans—metabolic effects. *Lipids*. **36**, 773–781.

Steen, A., Palumbo, E., Deghorain, M., Cocconcelli, P.S., Delcour, J., Kuipers, O., Kok, J., Buist, G., Hols, P. (2005) Autolysis of *Lactococcus lactis* is increased upon D-alanine depletion of peptidoglycan and lipoteichoic acids. *Journal of Bacteriology* **187**, 114–124.

Thom, E., Wadstein, J., Gudmundsen, O. (2001) Conjugated linoleic acid reduces body fat in healthy exercising humans. *Journal of International Medical Research* **29**, 392–396.

Steidler, L., Hans, W., Schotte, L., Neiryck, S., Obermeier, F., Falk, W., Fiers, W., Remaut, E. (2000) Treatment of murine colitis by *Lactococcus lactis* secreting interleukin-10. *Science*. **289**, 1352–1355.

Steidler, L., Neiryck, S., Huyghebaert, N., Snoeck, V., Vermeire, A., Goddeeris, B., Cox, E., Remon, J.P., Remaut, E. (2003) Biological containment of genetically modified *Lactococcus lactis* for intestinal delivery of human interleukin 10. *Nature Biotechnology* **21**, 785–789.

Vandenbroucke, K., Hans, W., Van Huysse, J., Neiryck, S., Demetter, P., Remaut, E., Rottiers, P., Steidler, L. (2004) Active delivery of trefoil factors by genetically modified *Lactococcus lactis* prevents and heals acute colitis in mice. *Gastroenterology*. **127**, 502–513.

Verhulst, A., Janssen, G., Parmentier, G., Eyssen, H. (1987) Isomerization of polyunsaturated long chain fatty acids by

Propionibacterium. *Systematic and Applied Microbiology* **9**, 12–15.

Wells, J.M., Mercenier, A. (2003) Lactic acid bacteria as mucosal delivery system. In: Wood, B.J.J., Warner, P.J. (Eds). Genetics of lactic acid bacteria. New York: Kluwer Academic Publishers; pp261–290.

Yamasaki, M., Ikeda, A., Oji, M., Tanaka, Y., Hirao, A., Kasai, M., Iwata, T., Tachibana, H., Yamada, K. (2003) Modulation of body fat and serum leptin levels by dietary conjugated linoleic acid in Sprague-Dawley rats fed various fat level diets. *Nutrition*. **19**, 30–35.