## iScience



### Article

# Learning and visual discrimination in newly hatched zebrafish



Maria Santacà, Marco Dadda, Luisa Dalla Valle, Camilla Fontana, Gabriela Gjinaj, Angelo Bisazza

santacamaria@gmail.com

#### Highlights

Newly hatched zebrafish can associate a visual stimulus to a food reward

Learning occurs in just two training days and can be used to study perception

Hatchlings compare to adults in color and shape discrimination

Poorer performance was observed in discriminating rotated shapes and mirror images

Santacà et al., iScience 25, 104283 May 20, 2022 © 2022 The Author(s). https://doi.org/10.1016/ j.isci.2022.104283

## **iScience**

#### Article

## Learning and visual discrimination in newly hatched zebrafish

Maria Santacà,<sup>1,4,\*</sup> Marco Dadda,<sup>2</sup> Luisa Dalla Valle,<sup>1</sup> Camilla Fontana,<sup>1</sup> Gabriela Gjinaj,<sup>2</sup> and Angelo Bisazza<sup>2,3</sup>

#### SUMMARY

With the exception of humans, early cognitive development has been thoroughly investigated only in precocial species, well developed at birth and with a broad behavioral and cognitive repertoire. We investigated another highly altricial species, the zebrafish, *Danio rerio*, whose embryonic development is very rapid (< 72 h). The hatchlings' nervous system is poorly developed, and their cognitive capacities are largely unknown. Larvae trained at 8 days post fertilization rapidly learned to associate a visual pattern with a food reward, showing significant performance at 10 days post fertilization. We exploited this ability to study hatchlings' discrimination learning capacities. Larvae rapidly and accurately learned color and shape discriminations. They also discriminated a figure from its mirror image and from its 90°-rotated version, although with lower performance. Our study revealed impressive similarities in learning and visual discrimination capacities between newborn and adult zebrafish, despite their enormous differences in brain size and degree of development.

#### INTRODUCTION

The study of brain maturation and early cognitive development has played a central role for the understanding of how the nervous system works. Unlike other topics of cognitive sciences and neurobiology that have been studied mainly on animal models, almost all knowledge about the cognitive development of vertebrates comes from studies in humans. Humans are characterized by an extreme form of altriciality, which has recently evolved in response to mechanical and energetic constraints imposed by cranial capacity expansion and bipedal locomotion (Haeusler et al., 2021). Newborn infants are helpless and highly dependent on their parents. The sensory and motor systems are poorly developed, partly because the degree of myelination is extremely low at birth (Gibson, 1991). At birth, learning, memory, and other cognitive abilities are already present, but in a rudimentary form. They gradually improve during infancy, childhood, and adolescence in parallel with the nervous system's general maturation (Bjorklund and Causey, 2017). This is exemplified by development of numerical discrimination abilities. In the first week of life, infants discriminate a 0.33 ratio between the smaller and larger quantities; at six months, they discriminate a 0.50 ratio, and, at 10 months, they discriminate a 0.67 ratio but not a 0.80 ratio (reviewed in Cantrell and Smith, 2013). Numerical acuity continues to increase throughout infancy and adolescence: six-year-old children can discriminate a 0.83 numerical ratio and adults a 0.90 ratio (Halberta and Feigenson, 2008).

The cognitive development is much less studied in the other vertebrates. Altriciality is by far the most common condition in mammals and birds, whose young ones are usually born in a state of extreme immaturity of the nervous system. Nevertheless, the ontogeny of perception, learning, and other cognitive functions has been examined in detail in only a few precocial species. The most studied species is undoubtedly the domestic chicken (*Gallus gallus*). Chicks are largely independent at birth, and their sensory and cognitive motor skills are fully developed. Research on newborn chicks revealed that they are born with a set of sophisticated cognitive abilities and that these capacities are in many cases not dissimilar to those exhibited by adult hens (Daisley et al., 2011; Vallortigara et al., 2010).

Cognitive development of cold-blooded vertebrates has been studied mainly in the guppy (*Poecilia retic-ulata*), an extremely precocial teleost species. After a four-week pregnancy, guppies give birth to young ones that are miniatures of the adults. Newborn guppies swim in a coordinated fashion, perform predator inspection, discriminate social groups based on their numerosity, distinguish familiar from unfamiliar objects, and can rapidly learn to associate a specific object or a given number of items with a food reward

<sup>1</sup>Department of Biology, University of Padova, Viale Giuseppe Colombo 3 - Via Ugo Bassi 58/B, 35131 Padova, Italy

<sup>2</sup>Department of General Psychology, University of Padova, 35131 Padova, Italy

<sup>3</sup>Padua Neuroscience Center, University of Padova, 35131 Padova, Italy

<sup>4</sup>Lead contact

\*Correspondence: santacamaria@gmail.com https://doi.org/10.1016/j.isci. 2022.104283













Figure 1. Experimental set up and stimuli of Experiment 1 (A) Apparatus used for individual training of zebrafish larvae. (B) The two patterns used for the learning task of Experiment 1.

### (Bisazza et al., 2010; Evans and Magurran, 2000; Magurran and Seghers, 1990; Miletto Petrazzini et al., 2012; Piffer et al., 2013).

It is clear from the above description that although an extreme immaturity at birth is the typical condition of mammals, birds, and teleosts, there is very little information on species with this developmental mode. This prevents us from understanding whether the developmental patterns observed in humans are typical of all vertebrates or rather they reflect the unique features that characterize the nervous system of our species. In zebrafish (*Danio rerio*), development time from fertilization to hatching is among the shortest of all vertebrates, lasting between 48 and 72 h. After hatching, larvae spend most of the time inactive, lying on their sides. Zebrafish start free swimming from 120 hpf (hours post fertilization). However, it is only after 140 hpf that development of the digestive tract is completed, and they begin feeding. This period is defined as a non-feeding eleutheroembryo stage, and it is considered a post-hatching extension of embryonic development (Balon, 1975; Strähle et al., 2012). In the period following the start of autonomous feeding, the zebrafish does not exhibit complex behaviors other than swimming and feeding. Attraction to conspecifics and complex social behavior appears much later, when zebrafish are approximately three weeks old (Dreosti et al., 2015).

Simple forms of learning can be demonstrated in larval zebrafish. For example, 7–10 days post fertilization (dpf) larvae can be conditioned to avoid the darker side of their tank by delivering a mild electric shock (Yang et al., 2019). It was shown that the visual system of zebrafish matured precociously, at the time of hatching. Larvae showed a startle response to a rapid change in light intensity at approximately 72 hpf, and tracking eye movements developed at approximately 80 hpf (Easter and Nicola, 1996). Little is known about the capacity to discriminate objects in the first week after hatching. Indirect evidence suggests that beginning from 14 dpf, larval zebrafish recognized a familiar stimulus based on its color or shape (Bruzzone et al., 2020). In addition, at 14 dpf, larvae have marked preferences for certain visual patterns, suggesting that at this age they can discriminate their features (Gatto et al., 2021).

The aim of this study was 2-fold. The objective of the first experiment was to determine whether one-weekold zebrafish could associate a food reward to a specific visual pattern and to validate an appetitive learning procedure for larval zebrafish. In four subsequent experiments, we used this procedure to determine the visual discrimination capacities of newborn zebrafish in tasks requiring discrimination between stimuli that differed in color, shape, or spatial orientation.

#### RESULTS

#### Experiment 1. Development and validation of an operant conditioning procedure

Twelve zebrafish larvae were trained in a two-compartment apparatus for five consecutive days (from 8 to 12 dpf), and food was delivered near one of the two visual patterns placed at the two far ends (Figure 1). To minimize social isolation, for the first two days, training was done in a group. Subjects were then individually

#### iScience Article





#### Figure 2. Learning performance in the five days of training (from 8 to 12 dpf) of Experiment 1

Performance was calculated as preference for the reinforced stimulus in the 90-min period preceding the delivery of the food reward. For the first two days of training (group training), preference is expressed as the proportion of larvae in the compartment with the reinforced stimulus. For the remaining three days (individual training), preference is expressed as the proportion of the time that a subject spent in the compartment with the reinforced stimulus. The figure shows distinct learning curves for subjects trained using vertical bars or a dot array as positive. Data points represent mean  $\pm$  SEM The black dotted line represents chance performance.

trained for the remaining three days. Performance was measured as time spent in the compartment with the positive stimulus during the 90-min period preceding food delivery. During each 90-min period, larvae moved between compartments on average  $18.24 \pm 9.78$  times. Subjects significantly preferred the reinforced stimulus (proportion of time  $0.700 \pm 0.086$ ; one sample t-test, t(11) = 8.085, p < 0.001, achieved power = 1.000). Performance did not significantly vary across days (LMM:  $F_{2,60} = 1.662$ , p = 0.198; Figure 2), and there was no difference between larvae reinforced on vertical bars or on a dot array as positive ( $F_{1,60} = 0.002$ , p = 0.969; Figure 2). Timing of stimuli inversion had no effect ( $F_{1,60} = 0.236$ , p = 0.629), and no interaction was statistically significant (all p values > 0.351). Preference for the reinforced stimulus was significant, even considering only the first day of individual training (0.625  $\pm$  0.151; one sample t-test, t(11) = 2.864, p < 0.05).

As larvae showed a significant performance already on the first day of individual training, we reconstructed the initial stages of learning by re-analyzing the videos that had been recorded in the group training phase for the purpose of checking if larvae had learned to move between the compartments. The proportion of subjects swimming in the compartment with the reinforced stimulus was not significantly different from chance on the first day of group training (proportion in the reinforced compartment 0.514  $\pm$  0.102, one sample t-test, t(5) = 0.328, p = 0.756), but it was significant on the second day of group training (0.575  $\pm$  0.040, t(5) = 4.555, p < 0.01; Figure 2). When individual and group training were analyzed together, we found a significant correlation between the day and the degree of preference (Kendall non-parametric correlation:  $\tau$  = 0.236; p < 0.01) revealing a significant improvement in subjects' accuracy over training sessions.

#### **Experiment 2. Color discrimination**

With the same procedure as the previous experiment, twelve zebrafish larvae were tested on a red-green discrimination (Figure 3A). The mean number of passages between compartments was 11.93  $\pm$  4.94 (Mean  $\pm$  SD). Subjects significantly preferred the reinforced stimulus (0.729  $\pm$  0.110; one sample t-test, t(11) = 7.219, p < 0.001, achieved power = 1.000). Performance did not vary across days (LMM:  $F_{2,60}$  = 0.819, p = 0.446; Figure 3A). Larvae reinforced on the green or red did not differ in performance ( $F_{1,60}$  = 0.330, p = 0.566); timing of stimuli inversion had no effect ( $F_{1,60}$  = 1.265, p = 0.265); and no interaction was statistically significant (all p values > 0.121).

#### **Experiment 3. Shape discrimination**

Twelve zebrafish larvae were tested on a triangle-circle discrimination (Figure 3B). The mean number of passages between compartments was 15.35  $\pm$  6.81 (Mean  $\pm$  SD). Subjects significantly preferred the reinforced stimulus (0.631  $\pm$  0.139; one sample t-test, t(11) = 3.251, p = 0.008, achieved power = 0.844). Performance did not vary across days (LMM:  $F_{2,60}$  = 0.571, p = 0.568; Figure 3B). Larvae reinforced on the







#### Figure 3. Results of Experiment 2-5

Learning curves (proportion of the time spent in the compartment with the reinforced stimulus) of zebrafish larvae in the three days of individual training of Experiment 2-5 (A: color discrimination; B: shape discrimination; C: mirror-image discrimination; D: horizontal-vertical discrimination). Data points represent mean  $\pm$  SEM The dotted lines represent chance performance.

triangle or circle did not differ in performance ( $F_{1,60} = 4.852$ , p = 0.062); timing of stimuli inversion had no effect ( $F_{1,60} = 0.979$ , p = 0.354); and no interaction was statistically significant (all p values > 0.328).

#### **Experiment 4. Mirror-image discrimination**

Twelve zebrafish larvae were tested in a discrimination of a figure from its mirror image (Figure 3C). The mean number of passages was 15.85  $\pm$  4.09 (Mean  $\pm$  SD). Subjects significantly preferred the reinforced stimulus (0.543  $\pm$  0.051; one sample t-test, t(11) = 2.932, p = 0.014, achieved power = 0.758). Performance did not vary across days (LMM:  $F_{2,60} = 0.381$ , p = 0.685; Figure 3C). Larvae reinforced on E or its mirror reverse did not differ in performance ( $F_{1,60} = 0.015$ , p = 0.903); timing of stimuli inversion had no effect ( $F_{1,60} = 0.506$ , p = 0.480); and no interaction was statistically significant (all p values > 0.711).

#### **Experiment 5. Horizontal-vertical discrimination**

Twelve zebrafish larvae were tested in a discrimination of a figure from the same figure rotated by 90° (Figure 2D). The mean number of passages was 12.42  $\pm$  3.74 (Mean  $\pm$  SD). Subjects significantly preferred the reinforced stimulus (0.572  $\pm$  0.099; one sample t-test, t(11) = 2.517, p = 0.029, achieved power = 0.631). Performance did not vary across days (LMM:  $F_{2,60} = 0.030$ , p = 0.822; Figure 3D). Larvae trained with horizontal or vertical bars had similar performances ( $F_{1,60} = 0.499$ , p = 0.483); timing of stimuli inversion had no effect ( $F_{1,60} = 0.246$ , p = 0.622); and no interaction was statistically significant (all p values > 0.418).







#### Figure 4. Comparison of the results of the four discrimination tasks

The boxplots report median, lower, and upper quartiles; whiskers represent values within 1.5 times the interquartile range. The asterisks (\*) denote a significant departure from chance level (one sample t-test, p < 0.05).

#### **Comparison of the discrimination tasks**

The overall analysis on the proportion of the time spent in two choice compartments revealed that there was a significant effect of the type of discrimination test on larvae performance (LMM:  $F_{3,60} = 5.161$ , p = 0.003; Figure 4) but no effect of the day ( $F_{2,262} = 1.144$ , p = 0.320) nor of the interaction ( $F_{6,262} = 0.233$ , p = 0.965). Tukey post hoc test revealed that performance was significantly better in the color discrimination than in the mirror-image and horizontal-vertical discriminations (both p-values < 0.05). No other comparison was statistically significant (all p-values > 0.339).

#### DISCUSSION

Previous studies showed that newborn zebrafish can learn to respond to simple stimuli. For example, they can be conditioned to turn their tails to obtain relief from heat exposure or learn to stay away from the darker part of the tank to avoid an electric shock (Yang et al., 2019). Using a new appetitive learning paradigm, we demonstrated that, one week after hatching, larval zebrafish rapidly associate a visual pattern with a food reward, and they can accurately discriminate two objects based on their color or shape. Moreover, we found that young larvae can perform complex visual discrimination typical of higher vertebrates, such as discriminating a figure from its mirror image or from its 90°- rotated version.

#### Appetitive learning in larval zebrafish

The first experiment aimed to develop an operant conditioning procedure for young larvae. From 8 dpf to 12 dpf, we exposed larvae twice a day to two visual patterns, delivering food near one of the two stimuli. As consequence of this pairing, larvae increased time spent in association with the reinforced pattern. Moreover, we found no difference between subjects trained on one or the other stimulus, excluding the possibility that our results were explained by innate preferences for some patterns. To reduce the length of social isolation, the first two days of training were conducted in a group, and therefore, we have data on individual performance only for the last three days of training. Quite unexpectedly, we observed learning to occur very early, with a significant performance observed in the first day of individual training, when larvae were 10 dpf. Although we are unable to determine exactly when each subject started to learn, we re-analyzed the videos that were recorded during group training to check movements between compartments. Measuring the proportion of subjects present in the two compartments in the 90-min period preceding the delivery of food reward, we found a significant preference for the reinforced stimulus from the second day of training, at 9 dpf.

This is an especially interesting finding because, for the first time, it shows the existence of appetitive learning in zebrafish in the first week after hatching. It has been suggested that the ability of a larva to learn to avoid a dark place where it previously received an electric shock is adaptive because, in nature, this capacity allows them to avoid dangerous parts of their habitats, such as those containing predators (Yang et al., 2019). Similarly, it is likely beneficial for recently hatched zebrafish to learn and remember a particular location or the characteristics of the microhabitat where they had previously consumed food.





The two patterns used in the first experiment differed for multiple cues (e.g., shape, size, and number of the single items composing the patterns), and larvae may have considered one of these cues or even a more basic feature of the image [e.g., the spatial frequency (Murphy et al., 2015; Orger et al., 2000)] to discriminate between them. Indirect evidence suggests that, after 14 dpf, larval zebrafish might be able to discriminate the color and the shape of an object (Bruzzone et al., 2020; Gatto et al., 2021). The visual system of zebrafish starts to mature earlier, one to two days after hatching, and at 7–10 dpf, larvae have been shown to perform simple discriminations, such as recognizing the dark side of their tank (Easter and Nicola, 1996). Given the results of the first experiment, we performed a series of discrimination learning experiments in the study's second part to investigate perceptual abilities of zebrafish at this developmental stage.

#### Color and shape discrimination

In the five days of training, larvae learned to discriminate two figures based on either color or shape. In both cases, the performance was very good, and there was no difference between the subjects trained on one or the other stimulus. In both experiments, we did not observe an improvement in performance during the experiment, but this may depend on the fact that we did not have information on the first two training days.

Hatching much earlier than the other teleosts, zebrafish larvae have completed only part of their development, and for the first two weeks, they exhibit a limited behavioral repertoire. However, the results of these experiments seem to indicate that some advanced cognitive functions are already present a few hours after hatching. These include the capacity to discriminate objects and memorize their characteristics and the possibility to associate specific reactions (e.g., approach or avoidance) to these objects as consequence of previous experience with them.

From this point of view, newly hatched zebrafish are not much different from newborn guppies, which are delivered after a more than 4-week gestation, and whose neural development is at a much more advanced stage (Miletto Petrazzini et al., 2012). The parallel with the condition of human infants is also interesting. In a context of general underdevelopment of the nervous system at birth and a complete helpless condition, some functions are more developed in human infants than would be typical for an altricial species. The senses of human infants—hearing, olfaction, and vision—are already functional at birth (and to some extent already in the fetus). Immediately after birth, human babies begin visual searching and exhibit preferences for looking at some patterns, orienting toward human faces in particular (reviewed in Rosenberg, 2021). Newborn babies show coordination of auditory and visual functions, being able to localize an auditory stimulus and turn their eyes in its direction, can discriminate shape, color and numerosity and show simple forms of learning (Cantrell and Smith, 2013; Slater et al., 1983; Wertheimer, 1961). Conversely, capabilities such as motor control of hand, locomotion, or vocal communication are extremely immature in the neonate relative to the adult individual (Bjorklund and Casey, 2017; Rosenberg, 2021). It was suggested that this developmental heterochrony has been shaped by natural selection and that early developing functions are those best suited for their time in life, for example those that allow an efficient interaction with their social group and a rapid acquisition of key information about their environment (Bjorklund and Casey, 2017; Rosenberg, 2021). Similarly, it might be adaptive for newly hatched zebrafish larvae developing neural structures that underlie functions that are essential to survive in the post-hatching environment (for example, those allowing to associate a visual pattern with food or hazards), while delaying maturation of other functions (for example, those necessary for social interactions) that do not confer significant benefits at birth.

It is interesting to note that, in our study, only five stimulus-reward pairings were necessary to establish a significant association, and that larvae reached 70%–80% accuracy in color and shape discrimination after only 10 reinforced trials. By comparison, in a recent study that used a similar appetitive paradigm and identical stimuli, adult zebrafish required five to six days, 12 trials a day, to reach similar levels of performance (Santacà et al., 2021). There may be various explanations for this apparent superiority in discrimination learning by larvae. From an ecological perspective, it is possible that, in the habitat in which larvae live, the association between food availability and particular environmental feature is stable over time, and it pays to memorize rapidly the landscape features that are associated with the likelihood of finding food. Investigations of learning and memory in insects have shown that learning speed is not necessarily related to computational capacity. Notwithstanding their extremely small brains, species such as bees or moths can learn an association between stimulus and reward after just one or a few stimulus-reward pairings (Balkenius, 2010; Menzel et al., 1993). Rapid learning, for example, may have evolved in bees





because, once they discover a plant that produces nectar, it pays to continue to forage on the same plant species during the following hours or days (Muth et al., 2016). Other explanations cannot be ruled out. One possible reason for faster learning in larvae is a difference in food motivation. Larval zebrafish eat a few times a day, and missing a meal may more severely affect a developing larva than an adult, which can usually starve for days without too serious consequences.

#### **Discrimination of stimulus orientation**

In Experiment 4-5, we examined larval zebrafish in two discrimination tasks that are generally considered complex, the discrimination of a figure from the same figure rotated by 90° and the discrimination of a figure from its mirror image. Some species seem to have problems in discriminating rotated figures, although many others easily learn this type of task (Messenger and Sanders, 1971; Srinivasan et al., 1994; Sutherland and Carr, 1964). In adult zebrafish, the learning rate for 90°-rotated figures (the same stimuli used in the present study) matched the learning rate for color and shape discrimination (Santacà et al., 2021). Larvae were able to discriminate between the two rotated figures, but their performances were lower than that shown in previous experiments. Therefore, the comparison with the performance of adult zebrafish seems to indicate that the ability to discriminate figures that differ in orientation is less developed at birth, and this capacity improves later in life with the maturation of the nervous system or with experience.

In the discrimination of a figure from its mirror image, the accuracy of larval zebrafish was just above chance, reaching approximately 55% preference for the reinforced stimulus by the experiment's end, the lowest performance of the whole study. This result is not surprising, as the discrimination of mirror images was found to be an extremely difficult task for a variety of other organisms, such as octopuses, fish, rats, monkeys, and humans (Bornstein et al., 1978; Gierszewski et al., 2013; Gregory and McCloskey, 2010; Lashley, 1938; Sanford and Ward, 1986; Sutherland, 1957). Though very low, the performance of newborn zebrafish does not appear much different from the performances observed in the adults of the investigated teleost species (Gierszewski et al., 2013; Lucon-Xiccato and Bisazza, 2016; Santacà et al., 2021). The difficulty of discriminating mirror images is commonly explained with the fundamentally bilateral symmetry of the perceiving organism's nervous system (Corballis and Beale, 2020; Noble, 1968). However, other authors point out that there are rare situations in nature in which it would be beneficial for an organism to discriminate mirror images, and probably, there has been little or no selective pressure for evolving this capacity (Bornstein et al., 1978).

#### Conclusions

Rather unexpectedly for an extremely altricial species, we found that newly hatched zebrafish possess cognitive capacities that, in some domains, compare with those of the adults. A mechanism for appetitive learning adds to the other forms of associative learning previously described. Furthermore, larvae appear capable of rapidly memorizing the characteristics of an object based on multiple cues and operating subtle discriminations among similar objects. Our results suggest that, as observed in human infants (reviewed in Rosenberg, 2021; Trevathan and Rosenberg, 2016), in a highly altricial species, some advanced cognitive functions may emerge early in brain development if these functions have a high survival value at a specific developmental stage.

Zebrafish have recently emerged as an important model for understanding the mechanisms of brain functioning and for investigating human neuropathologies. Most research in this field is done on embryos and larvae before 10 dpf due to their amenability to genetic manipulation, high-resolution brain imaging, and to the possibility of high-throughput *in vivo* screening. For example, mutant zebrafish lines have been obtained showing alterations of TAU protein functioning that express early in development and resemble the key pathological features of human TAU-related neurodegeneration (Paquet et al., 2009). Zebrafish models have been generated to investigate the neurodevelopmental basis of psychiatric disorders, for developing new antipsychotic drugs (e.g., Norton, 2013; Parker et al., 2014), and for studying re-myelination process in pathologies such as multiple sclerosis or brain injuries (Buckley et al., 2008). Until now, research has been hampered by the difficulty to evidence complex behaviors and advanced cognitive functions in early developing larvae. The discovery that zebrafish can be rapidly conditioned and possess remarkable discrimination abilities within days of hatching opens the way for future developments in research, for example studying the neural organization of complex cognitive functions or using new measures of the effectiveness of therapeutic intervention on brain diseases.





#### Limitations of the study

We describe the occurrence of appetitive learning in recently hatched zebrafish larvae. The present study, however, has several limitations. First, to reduce the effects of social isolation, in this study, we trained larval zebrafish in a group for the first part of the experiment. The study revealed that associative learning occurred in larvae faster than we expected, and we therefore missed important information on the early stages of individual learning. Future studies should refine the procedure to overcome this problem. Another limitation of our study is that we tested larval zebrafish from 8 dpf, and we did not attempt to assess the precise onset of associative learning capacities in hatchlings. This might limit our procedure's application in neurobiological research because most brain imaging and brain mapping studies are conducted on younger larvae (usually 6 dpf), and many genetic tools have been developed for even earlier developmental stages. Future experiments would be necessary to assess our method's validity for brain studies. Lastly, our appetitive conditioning procedure lasted five days and at least some tasks show a gradual improvement in performance. At this age, the nervous system of a larval zebrafish is rapidly developing and our conditioning paradigm did not allow evaluating the relative contribution of learning and maturation to the performance curve. To unravel this point, future studies should attempt to employ faster protocols for assessment of cognitive functions, for example tests based on habituation or aversive conditioning that can usually be performed in hours or minutes (Yang et al., 2019).

#### **STAR**\***METHODS**

Detailed methods are provided in the online version of this paper and include the following:

- KEY RESOURCES TABLE
- **RESOURCE AVAILABILITY** 
  - O Lead contact
  - Materials availability
  - Data and code availability
- EXPERIMENTAL MODEL AND SUBJECT DETAILS
  - O Animals
- METHOD DETAILS
  - O Apparatus
  - O Stimuli
  - O Procedure
  - O Familiarization phase
  - Group training
  - O Individual training
- QUANTIFICATION AND STATISTICAL ANALYSIS

#### SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.isci.2022.104283.

#### ACKNOWLEDGMENTS

We would like to thank Stefano Massaccesi for his help in setting up the experimental apparatuses and all the students that helped with testing the animals and with the video analyses. The present work was carried out within the scope of the research program "Dipartimenti di Eccellenza" (art.1, commi 314–337 legge 232/2016, MIUR grant to the Department of General Psychology, University of Padova (Padova, Italy).

#### **AUTHOR CONTRIBUTIONS**

A.B. and L.D.V. conceived the research question. A.B., M.S., and M.D. designed the experiment. M.S. and G.G. conducted the experiments. C.F. contributed to the early development of the procedure and was in charge of breeding and fish maintenance. M.S. analyzed the data and all authors discussed the results. A.B. and M.D. obtained the fundings. A.B. and M.S. wrote the manuscript and finalized it with inputs from all authors. All authors approved the manuscript for publication.

#### **DECLARATION OF INTERESTS**

The authors declare no competing interests.



Received: December 26, 2021 Revised: March 14, 2022 Accepted: April 19, 2022 Published: May 20, 2022

#### REFERENCES

Balkenius, A., and Balkenius, C. (2010). Behaviour towards an unpreferred colour: can green flowers attract foraging hawkmoths? J. Exp. Biol. 213, 3257–3262. https://doi.org/10.1242/jeb.045161.

Balon, E.K. (1975). Terminology of intervals in fish development. J. Fish. Res. Board Can. 32, 1663–1670. https://doi.org/10.1139/f75-196.

Bilotta, J., and Saszik, S. (2001). The zebrafish as a model visual system. Int. J. Dev. Neurosci. *19*, 621–629. https://doi.org/10.1016/s0736-5748(01) 00050-8.

Bisazza, A., Piffer, L., Serena, G., and Agrillo, C. (2010). Ontogeny of numerical abilities in fish. PLoS One 5, e15516. https://doi.org/10.1371/ journal.pone.0015516.

Bjorklund, D.F., and Causey, K.B. (2017). Children's Thinking: Cognitive Development and Individual Differences (Sage Publications).

Bornstein, M.H., Gross, C.G., and Wolf, J.Z. (1978). Perceptual similarity of mirror images in infancy. Cognition 6, 89–116. https://doi.org/10. 1016/0010-0277(78)90017-3.

Bruzzone, M., Gatto, E., Lucon-Xiccato, T., Dalla Valle, L., Fontana, C.M., Meneghetti, G., and Bisazza, A. (2020). Measuring recognition memory in zebrafish larvae: issues and limitations. PeerJ *8*, e8890. https://doi.org/10.7717/peerj. 8890.

Buckley, C.E., Goldsmith, P., and Franklin, R.J.M. (2008). Zebrafish myelination: a transparent model for remyelination? Dis. Model. Mech. 1, 221–228. https://doi.org/10.1242/dmm.001248.

Cantrell, L., and Smith, L.B. (2013). Open questions and a proposal: a critical review of the evidence on infant numerical abilities. Cognition 128, 331–352. https://doi.org/10.1016/j. cognition.2013.04.008.

Corballis, M.C., and Beale, I.L. (2020). The Psychology of Left and Right (Routledge).

Daisley, J.N., Rosa Salva, O., Regolin, L., and Vallortigara, G. (2011). Social cognition and learning mechanisms: experimental evidence in domestic chicks. Interact. Stud. 12, 208–232. https://doi.org/10.1075/is.12.2.02dai.

Dreosti, E., Lopes, G., Kampff, A.R., and Wilson, S.W. (2015). Development of social behavior in young zebrafish. Front. Neural Circuits *9*, 39. https://doi.org/10.3389/fncir.2015.00039.

Easter Jr, S.S., and Nicola, G.N. (1996). The development of vision in the zebrafish (*Danio rerio*). Dev. Biol. *180*, 646–663. https://doi.org/10. 1006/dbio.1996.0335.

Erdfelder, E., Faul, F., and Buchner, A. (1996). GPOWER: A general power analysis program. Behav. Res. Meth. Instrum. Comput. *28*, 1–11. https://doi.org/10.3758/bf03203630. Evans, J.P., and Magurran, A.E. (2000). Multiple benefits of multiple mating in guppies. Proc. Natl. Acad. Sci. U S A 97, 10074–10076. https://doi.org/ 10.1073/pnas.180207297.

Gatto, E., Bruzzone, M., and Lucon-Xiccato, T. (2021). Innate visual discrimination abilities of zebrafish larvae. Behav. Process. 193, 104534. https://doi.org/10.1016/j.beproc.2021.104534.

Gerlach, G., Hodgins-Davis, A., Avolio, C., and Schunter, C. (2008). Kin recognition in zebrafish: a 24-hour window for olfactory imprinting. Proc. Biol. Sci. 275, 2165–2170. https://doi.org/10. 1098/rspb.2008.0647.

Gibson, K.R. (1991). Myelination and behavioral development: a comparative perspective on questions of neoteny, altriciality and intelligence. In Brain Maturation and Cognitive Development: Comparative and Cross-Cultural Perspectives, K.R. Gibson and A.C. Peterson, eds. (New York: Routledge), pp. 29–63.

Gierszewski, S., Bleckmann, H., and Schluessel, V. (2013). Cognitive abilities in Malawi cichlids (*Pseudotropheus* sp.): matching-to-sample and image/mirror-image discriminations. PLoS One 8, e57363. https://doi.org/10.1371/journal.pone. 0057363.

Gregory, E., and McCloskey, M. (2010). Mirrorimage confusions: implications for representation and processing of object orientation. Cognition 116, 110–129. https://doi.org/10.1016/j. cognition.2010.04.005.

Haeusler, M., Grunstra, N.D., Martin, R.D., Krenn, V.A., Fornai, C., and Webb, N.M. (2021). The obstetrical dilemma hypothesis: there's life in the old dog yet. Biol. Rev. 96, 2031–2057. https://doi. org/10.1111/brv.12744.

Halberda, J., and Feigenson, L. (2008). Developmental change in the acuity of the "number sense": the approximate number system in 3-4-5-and 6-year-olds and adults. Dev. Psychol. 44, 1457–1465. https://doi.org/10.1037/ a0012682.

Lashley, K.S. (1938). The mechanism of vision: XV. Preliminary studies of the rat's capacity for detail vision. J. Gen. Psychol. 18, 123–193. https://doi. org/10.1080/00221309.1938.9709894.

Lucon-Xiccato, T., and Bisazza, A. (2016). Male and female guppies differ in speed but not in accuracy in visual discrimination learning. Anim. Cogn. 19, 733–744. https://doi.org/10.1007/ s10071-016-0969-2.

Maack, G., and Segner, H. (2003). Morphological development of the gonads in zebrafish. J. Fish Biol. 62, 895–906. https://doi.org/10.1046/j.1095-8649.2003.00074.x.

Magurran, A.E., and Seghers, B.H. (1990). Population differences in the schooling behaviour of newborn guppies, *Poecilia reticulata*. Ethology 84, 334–342. https://doi.org/10.1111/j.1439-0310. 1990.tb00807.x.

Menzel, R., Greggers, U., and Hammer, M. (1993). Functional organization of appetitive learning and memory in a generalist pollinator, the honey bee. In Insect learning, D.R. Papaj and A.C. Lewis, eds. (Springer), pp. 79–125.

Messenger, J.B., and Sanders, G.D. (1971). The inability of Octopus vulgaris to discriminate monocularly between oblique rectangles. Int. J. Neurosci. 1, 171–173. https://doi.org/10.3109/ 00207457109146968.

Miletto Petrazzini, M.E., Agrillo, C., Piffer, L., Dadda, M., and Bisazza, A. (2012). Development and application of a new method to investigate cognition in newborn guppies. Behav. Brain Res. 233, 443–449. https://doi.org/10.1016/j.bbr.2012. 05.044.

Murphy, M.S., Brooks, D.I., and Cook, R.G. (2015). Pigeons use high spatial frequencies when memorizing pictures. J. Exp. Psychol. Anim. Learn. Cogn. 41, 277–285. https://doi.org/10. 1037/xan0000055.

Muth, F., Papaj, D.R., and Leonard, A.S. (2016). Bees remember flowers for more than one reason: pollen mediates associative learning. Anim. Behav. 111, 93–100. https://doi.org/10. 1016/j.anbehav.2015.09.029.

Noble, J. (1968). Paradoxical interocular transfer of mirror-image discriminations in the optic chiasm sectioned monkey. Brain Res. *10*, 127–151. https://doi.org/10.1016/0006-8993(68)90118-2.

Norton, W.H.J. (2013). Toward developmental models of psychiatric disorders in zebrafish. Front. Neural Circuits 7, 79. https://doi.org/10. 3389/fncir.2013.00079.

Orger, M.B., Smear, M.C., Anstis, S.M., and Baier, H. (2000). Perception of Fourier and non-Fourier motion by larval zebrafish. Nat. Neurosci. *3*, 1128– 1133. https://doi.org/10.1038/80649.

Paquet, D., Bhat, R., Sydow, A., Mandelkow, E.M., Berg, S., Hellberg, S., Fälting, J., Distel, M., Köster, R.W., Schmid, B., and Haass, C. (2009). A zebrafish model of tauopathy allows in vivo imaging of neuronal cell death and drug evaluation. J. Clin. Invest. *119*, 1382–1395. https:// doi.org/10.1172/jci37537.

Parker, M.O., Brock, A.J., Sudwarts, A., and Brennan, C.H. (2014). Atomoxetine reduces anticipatory responding in a 5-choice serial reaction time task for adult zebrafish. Psychopharmacology 231, 2671–2679. https:// doi.org/10.1007/s00213-014-3439-z.

Piffer, L., Miletto Petrazzini, M.E., and Agrillo, C. (2013). Large number discrimination in newborn fish. PLoS One 8, e62466. https://doi.org/10. 1371/journal.pone.0062466.





Rosenberg, K.R. (2021). The evolution of human infancy: why it helps to Be helpless. Annu. Rev. Anthropol. 50, 423–440. https://doi.org/10.1146/ annurev-anthro-111819-105454.

Sanford, C.G., and Ward, J.P. (1986). Mirror image discrimination and hand preference in the bushbaby (*Galago senegalensis*). Psychol. Rec. 36, 439–449. https://doi.org/10.1007/bf03394964.

Santacà, M., Agrillo, C., Miletto Petrazzini, M.E., and Bisazza, A. (2020). The ontogeny of continuous quantity discrimination in zebrafish larvae (Danio rerio). Anim. Cogn. 23, 731–739. https://doi.org/10.1007/s10071-020-01384-1.

Santacà, M., Dadda, M., Miletto Petrazzini, M.E., and Bisazza, A. (2021). Stimulus characteristics, learning bias and visual discrimination in zebrafish (Danio rerio). Behav. Process. 192, 104499. https://doi.org/10.1016/j.beproc.2021. 104499.

Slater, A., Morison, V., and Rose, D. (1983). Perception of shape by the new-born baby. Br. J. Dev. Psychol. 1, 135–142. https://doi.org/10. 1111/j.2044-835x.1983.tb00551.x. Srinivasan, M.V., Zhang, S.W., and Witney, K. (1994). Visual discrimination of pattern orientation by honeybees: performance and implications for 'cortical' processing. Philos. T. Roy. Soc. B 343, 199–210.

Strähle, U., Scholz, S., Geisler, R., Greiner, P., Hollert, H., Rastegar, S., Schumacher, A., Selderslaghs, I., Weiss, C., Witters, H., and Braunbeck, T. (2012). Zebrafish embryos as an alternative to animal experiments—a commentary on the definition of the onset of protected life stages in animal welfare regulations. Reprod. Toxicol. 33, 128–132. https://doi. org/10.1016/j.reprotox.2011.06.121.

Sutherland, N.S., and Carr, A.E. (1964). Shape discrimination by rats: squares and rectangles. Br. J. Psychol. 55, 39–48. https://doi.org/10.1111/j.2044-8295.1964. tb00896.x.

Sutherland, N.S. (1957). Visual discrimination of orientation by octopus. Br. J. Psychol. 48, 55–71. https://doi.org/10.1111/j.2044-8295.1957. tb00599.x.



**iScience** 

Article

Vallortigara, G., Regolin, L., Chiandetti, C., and Rugani, R. (2010). Rudiments of mind: insights through the chick model on number and space cognition in animals. Comp. Cogn. Behav. Rev. 5, 78–99. https://doi.org/10.3819/ccbr.2010.50004.

Wertheimer, M. (1961). Psychomotor coordination of auditory and visual space at birth. Science 134, 1692. https://doi.org/10.1126/ science.134.3491.1692.

Yang, W., Meng, Y., Li, D., and Wen, Q. (2019). Visual contrast modulates operant learning responses in larval Zebrafish. Front. Behav. Neurosci. 13, 4. https://doi.org/10.3389/fnbeh. 2019.00004.

Zellner, D., Padnos, B., Hunter, D.L., MacPhail, R.C., and Padilla, S. (2011). Rearing conditions differentially affect the locomotor behavior of larval zebrafish, but not their response to valproate-induced developmental neurotoxicity. Neurotoxicol. Teratol. 33, 674–679. https://doi. org/10.1016/j.ntt.2011.06.007.





#### **STAR\*METHODS**

#### **KEY RESOURCES TABLE**

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Experimental models: Organisms/strains		
Danio rerio	Housed at the Department of General	n/a
	Psychology, University of Padova, Italy	
Software and algorithms		
R Studio	R Development Core Team, 2020; R Foundation for statistical computing	https://www.r-project.org/

#### **RESOURCE AVAILABILITY**

#### Lead contact

Information and requests about this study should be directed to and will be fulfilled by the lead contact, Maria Santacà (santacamaria@gmail.com).

#### **Materials** availability

No new materials were generated in this study.

#### Data and code availability

- All data and code can be requested from the lead contact.
- This paper does not report original code.
- Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

#### **EXPERIMENTAL MODEL AND SUBJECT DETAILS**

#### Animals

The subjects were wild-type zebrafish larvae that originated from an outbred stock maintained in the zebrafish facility of the Department of Biology, University of Padova. The larvae were housed in several petri dishes (10 cm Ø, h:1.5 cm) in a solution of Fish Water 1× (0.5 mM NaH<sub>2</sub>PO<sub>4</sub>\*H<sub>2</sub>O, 0.5 mM Na<sub>2</sub>HPO<sub>4</sub>\*H<sub>2</sub>O, 1.5 g Instant Ocean, 1 L deionised H<sub>2</sub>O) and Methylene blue (0.0016 g/l). Before the experiment, they were housed in the same room at a density of approximately 30 larvae per petri dish, which were maintained at a temperature of 28.5 ± 1°C and illuminated according to a 14:10 h light:dark cycle. Zebrafish larvae were fed twice a day with dry food (an admixture of GEMMA Micro 75 and TetraMin flakes, particle size: 0.75 mm) from the age of 6 dpf.

We tested 12 zebrafish in each experiment, for a total of 60 subjects. Sample size was determined in a pilot experiment (N = 4) with a procedure essentially similar to that of Experiment 1. In this test the preference for the reinforced stimulus was  $0.630 \pm 0.110$ . We estimated a minimum sample size of 10 to achieve a statistical significance with 90% power and a two-tailed p-value of 0.05 (G\*Power 3.1.9.2 software; Erdfelder et al., 1996). Throughout this manuscript, we used the standard age classification for zebrafish studies, which starts with the fertilisation day and is expressed in days post fertilisation or dpf (e.g., Bilotta and Saszik, 2001). At the beginning of the individual test (see details below), larvae were 10 dpf, and their sex was undetermined because sexual differentiation occurs much later at approximately week 11–12 post-fertilisation (Maack and Segner, 2003).

#### **METHOD DETAILS**

#### Apparatus

Two different experimental apparatuses were used for this study. The familiarisation phase and the initial group training were conducted in an hourglass-shaped apparatus (12 × 4.8 cm and 4 cm high; Figure S1;





Santacà et al., 2020), 3D printed with white PLA material, and filled with 3.5 cm of Fish Water  $1 \times$ . A central corridor (4.3 cm in length) connected two identical compartments. In the middle of the corridor, a 3D printed, grey plastic panel (3 × 3.2 cm) with a central hole (1 cm Ø) allowed larvae to move between compartments, at the same time preventing the diffusion of food and minimizing spread of food scent from one compartment to the other.

The individual visual discrimination tests were conducted in a similar but smaller apparatus (7  $\times$  4 cm and 4 cm high; Figure 1A) filled with 3.5 cm of Fish Water 1 $\times$ . Four group training apparatuses were placed in a white plastic box (60  $\times$  40 cm and 27 cm high) and lit by two 0.72 W LED lamps placed symmetrically along the major axis (Figure S2). Internally, each wall of the box was covered with white plastic sheets. An identical box was used for the individual training phase and could contain up to twelve apparatuses at the same time. Video cameras were placed above apparatuses to record larvae behaviour.

#### Stimuli

Stimuli were created using Microsoft PowerPoint and printed on 4.8  $\times$  3.8 cm white, laminated cards. Experiment 1 aimed to develop and validate an operant conditioning procedure for larval zebrafish. To minimise the role of stimulus discrimination, we used two different visual patterns. We avoid using two hues, such as black and white, because larvae have strong preferences for the luminance of their environment. Instead, we used two patterns that differed for multiple cues (e.g., orientation and spatial frequency of the pattern, shape, size and number of the single items composing it), although they had the same percentage of black and white surfaces. One pattern consisted of three identical and parallel vertical bars (3.65 x 0.30 cm), the other consisted of an array of 63 equally spaced (0.2 cm) dots (Figure 1B).

In Experiments 2–5, we used the same stimuli used in a recent study with adult zebrafish (Santacà et al., 2021). In Experiment 2, we tested zebrafish in a colour discrimination. The colour stimuli were one red and one green square of the same brightness (red: RGB: 255, 0, 0; green: RGB: 0, 255, 0;  $2 \times 2$  cm; Figure S3). In Experiment 3, we tested subjects in a shape discrimination. The stimuli were one black outlined circle and one black outlined triangle (circle:  $\emptyset = 2.26$  cm, area = 3.55 cm<sup>2</sup>, perimeter = 7.10 cm; triangle: base = 2.80 cm, height = 2.26 cm, area = 3.16 cm<sup>2</sup>, perimeter = 8.12 cm; Figure S3). In Experiment 4, we tested each subject in mirror-image discrimination. The stimuli consisted of the capital letter E from the Latin alphabet (height = 2.50 cm, width = 1.60 cm) and its left–right mirror-reversed image (Figure S3). In Experiment 5, we tested each subject in an orientation discrimination. The stimuli consisted of four parallel black bars (height = 2.88 cm, width = 0.4 cm) arranged vertically in one stimulus and horizontally in the other (Figure S3).

#### Procedure

The procedure was developed and validated in the first experiment and remained unchanged in the following four experiments. Previous studies suggested that social behaviour develops in larval zebrafish at approximately three weeks after fertilisation (Dreosti et al., 2015); however, it is possible that they need conspecific visual or olfactory cues for proper maturation and that may suffer from social isolation (Gerlach et al., 2008). In addition, keeping newly-hatched larvae isolated from conspecifics have been shown to reduce their activity (Zellner et al., 2011), an effect that could potentially reduce the number of passages between compartments and ultimately impair stimulus-reward association. Indeed, a pilot experiment confirmed that at 8 and 9 dpf the number of passages between compartments was extremely reduced in isolated larvae (number of passages in 90 min:  $0.22 \pm 0.26$ , N = 8) compared to larvae kept in group (number of passages per larva in 90 min: 5.01  $\pm$  2.63, N = 6 groups). This suggests that larval zebrafish have difficulty to learn to move between compartments if placed individually in our apparatus from the beginning. To minimise the duration of social isolation, larvae were initially trained for two days in a group and then individually for three days. The experiment consists of three phases, an initial two-day familiarisation phase (6-7 dpf), a two-day group training phase (8-9 dpf), and a three-day individual training phase (10-12 dpf). The first two phases were conducted in the larger version of the apparatus, whereas the individual training phase was conducted in the smaller individual apparatuses. For the entire duration of the experiment, larvae were fed twice a day (h 10:30 and h 15:00).

#### **Familiarization phase**

Twenty 6 dpf larvae were moved using a Pasteur pipette in the familiarisation apparatus, in which no grey panel was inserted. The morning of the subsequent day, the panel with the hole was inserted to habituate larvae to pass through the hole and move between the two compartments.





#### **Group training**

Group training began when larvae were 8 dpf and lasted two consecutive days. Half subjects were trained on one stimulus (dots array), and the other half were trained on the other (vertical bars). Each day, we administered two reinforced trials, one in the morning and one in the afternoon (at h 10:30 and h 15:00). This was performed by leaning the two stimuli against the apparatus's two short walls and delivering the food in the compartment with the reinforced stimulus. Stimuli remained in place during feeding and were removed 90 min after the delivery of the food reward. To avoid side biases, stimuli were swapped between compartments every other trial. For half of the groups, this was done before the morning trial, whereas for the other half, it was done before the afternoon trial. In Experiment 1, we also video-recorded the group training phase. The originally scope of these recordings was to check whether the new procedure was functioning i.e. if larvae were sufficiently active and if they had learned to move freely between the two compartments. After discovering that performance was already significant in the first day of individual training, these videos were also used to measure whether larvae had started to associate with the reinforced stimulus prior to the individual training phase.

#### Individual training

At the end of the second day of the group training, larvae were moved from the group apparatus to the individual test apparatuses. The test phase lasted three consecutive days, during which we administered two reinforced trials per day with the same schedule of the group training phase and video-recorded the subjects' behaviour for 90 min prior to food administration. To avoid any cueing effect due to food residuals from the previous trial, each subject was gently pipetted and moved to an identical clean apparatus at the end of each trial. The schedule for stimuli swapping was the same as the group learning phase. Overall, each subject performed 10 reinforced trials, four during the group training and six during the individual training.

#### QUANTIFICATION AND STATISTICAL ANALYSIS

From the video recordings of the individual test phase, we initially recorded the number of passages between the two compartments. Subject that moved less than ten times in the overall individual phase were considered inactive and their performance was not analysed. We then scored for each trial the time each larva spent in the compartment with the reinforced stimulus and in the compartment with the non-reinforced stimulus. One-third of the videos from all experiments were analyzed by two different experimenters. Interrater reliability was calculated with the Spearman rank correlation and was found to be high ( $\rho = 0.968$ , p < 0.001).

In Experiment 1, we additionally measured the number of larvae in the two compartments during the group training phase. Scoring was done by pausing the videos at one-minute intervals and counting the number of larvae in each compartment.

Analyses were performed in R version 3.5.2 (The R Foundation for Statistical Computing, Vienna, Austria, https:// www.r-project.org). We analysed the proportion of time spent in the compartment with the reinforced stimulus using one-sample t-tests to evaluate whether it differed compared to chance level (0.50) for each experiment. We computed achieved power using G\*Power 3.1.9.2 software (Erdfelder et al., 1996). Additionally, we performed a linear mixed-effects model (LMM, 'Imer' function of the 'Ime4' R package) for each experiment to assess whether the performance differed between days, between the type of reinforced stimulus, and between the type of inversion of the reinforced stimulus (between daily sessions or between days). Thus, the LMM was fitted with the serial number of the day, the type of reinforced stimulus, and the inversion as fixed factors and the individual ID as random factors. Lastly, we performed a LMM to investigate whether the larvae performance differed between the four different discrimination tests (i.e., colour, shape, mirror-image, and horizontal-vertical). This model was fitted with the discrimination and the serial number of the day as fixed effects, and the individual ID nested within the discrimination test as random effect. In case of a factor's significant effect, pairwise comparisons were performed with Tukey post hoc tests.

To analyse learning performance in the group training phase of Experiment 1, we checked whether the proportion of larvae in the compartments with the reinforced stimulus differed from the chance level (0.50) using one-sample *t*-tests. For this experiment, we used non-parametric Kendall's correlation test to analyse whether subjects' accuracy improved over training session when considering both the group and the individual training days.