



Transcriptional Profiles in the Chicken Ductus Arteriosus During Hatching

40

Satoko Shinjo, Toru Akaike, Eriko Ohmori, Ichige Kajimura, Nobuhito Goda, and Susumu Minamisawa

Keywords

Ductus arteriosus · Microarray analysis · Chicken

The ductus arteriosus (DA) of oviparous animals differs from mammals' in the embryonic gas exchange system and its anatomy. We performed transcriptional analysis of chicken DA, attempting to elucidate the similarity and diversity in the mechanism of DA closure among species.

Chicken DA proximal to the pulmonary arteries (proximal DA) closes after hatching while the DA proximal to the aorta (distal DA) remains open even after hatching [1]. Histological analysis revealed that the ductal wall of the proximal DA became thicker with fragmented elastic fibers from embryonic day (ED) 19. Therefore, we performed microarray analysis with proximal DA, distal DA, and aorta from chicken embryo at ED19. Clustering analysis found that the expression pattern of distal DA was similar to that of proximal DA than that of aorta although

S. Shinjo · T. Akaike · I. Kajimura
Department of Cell Physiology, The Jikei University School of Medicine,
Minato, Tokyo, Japan

E. Ohmori · S. Minamisawa (✉)
Department of Cell Physiology, The Jikei University School of Medicine,
Minato, Tokyo, Japan

Department of Life Science and Medical Bioscience, School of Advanced Science and
Engineering, Waseda University, Shinjuku, Tokyo, Japan
e-mail: sminamis@jikei.ac.jp

N. Goda
Department of Life Science and Medical Bioscience, School of Advanced Science and
Engineering, Waseda University, Shinjuku, Tokyo, Japan

distal DA has a similar structure to aorta. Subsequent pathway analysis with DAVID [2] revealed that proximal DA had enhanced expression of the melanogenetic genes compared with distal DA and aorta (Table 40.1). This result appears reasonable because proximal DA shares its developmental origin, neural crest, with melanocytes. Although several known genes such as transcription factor AP-2 beta (tfap2b) [3] were highly expressed in proximal and distal DA, we newly found proximal-DA-dominant genes. Further investigation would be required to understand the role of these genes in DA closure not only for the chick but also for mammals.

Table 40.1 KEGG pathway analysis by DAVID from chicken DA microarray

Gene symbol	Gene name	Proximal DA/aorta	Distal DA/aorta	Note
Proximal DA dominant pathways				
<i>Melanogenesis/tyrosine metabolism</i>				
DCT	Dopachrome tautomerase (dopachrome delta-isomerase, tyrosine-related protein 2)	1.44*	1.06	
EDNRB2	Endothelin receptor B subtype 2	1.31*	1.07	(1)
TYR	Tyrosinase (oculocutaneous albinism IA)	1.54*	0.94	(1)
TYRP1	Tyrosinase-related protein 1	1.48*	1.03	
WNT11	Wingless-type MMTV integration site family, member 11	1.33*	1.14	
<i>Arachidonic acid metabolism</i>				
CYP2C45	Cytochrome P-450 2C45	1.44*	1.02	
HPGDS	Hematopoietic prostaglandin D synthase	1.56*	1.02	
PTGS2	Prostaglandin-endoperoxide synthase 2 (prostaglandin G/H synthase and cyclooxygenase)	1.31*	1.21	
DA dominant pathways				
<i>Focal adhesion/ECM-receptor interaction</i>				
FIGF	c-fos-Induced growth factor (vascular endothelial growth factor D)	1.50*	1.29*	(2)
KDR	Kinase insert domain receptor (a type III receptor tyrosine kinase)	1.34*	1.32*	(2)
LAMA4	Laminin subunit alpha 4	1.33*	1.24	
LAMB1	Laminin, beta 1	1.36*	1.27*	
TNC	Tenascin C	2.15*	1.82*	
VWF	von Willebrand factor	1.39*	1.31*	
CHAD	Chondroadherin	1.24	1.22*	
ITGA1	Integrin, alpha 1	1.28	1.21*	
<i>VEGF signaling pathway</i>				
HSPB1	Heat shock 27kDa protein 1	1.34*	1.21*	
KDR	Kinase insert domain receptor (a type III receptor tyrosine kinase)	1.34*	1.32*	
PTGS2	Prostaglandin-endoperoxide synthase 2 (prostaglandin G/H synthase and cyclooxygenase)	1.31*	1.21*	
PLCG2	Phospholipase C, gamma 2 (phosphatidylinositol-specific)	1.17	1.20*	

The asterisk indicates that the component genes appeared in the result of DAVID analysis

(1) Only appeared in “melanogenesis,”

(2) Only appeared in “focal adhesion”

Acknowledgment This work was supported by grants from the Ministry of Education, Culture, Sports, Science and Technology of Japan (T.A., S.M.), MEXT-Supported Program for the Strategic Research Foundation at Private Universities (S.M.), the Vehicle Racing Commemorative Foundation (S.M.), The Jikei University Graduate Research Fund (S.M.), and the Miyata Cardiology Research Promotion Foundation (S.M.).

References

1. Belanger C, Copeland J, et al. Morphological changes in the chicken ductus arteriosus during closure at hatching. *Anat Rec (Hoboken)*. 2008;291:1007–15. <https://doi.org/10.1002/ar.20720>.
2. Huang da W, Sherman BT, et al. Systematic and integrative analysis of large gene lists using DAVID bioinformatics resources. *Nat Protoc*. 2009;4:44–57. <https://doi.org/10.1038/nprot.2008.211>.
3. Zhao F, Bosserhoff AK, et al. A heart-hand syndrome gene: *Tfap2b* plays a critical role in the development and remodeling of mouse ductus arteriosus and limb patterning. *PLoS One*. 2011;6:e22908. <https://doi.org/10.1371/journal.pone.0022908>.

Open Access This chapter is licensed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license and indicate if changes were made.

The images or other third party material in this chapter are included in the chapter's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the chapter's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder.

