

**animal**

<http://journals.cambridge.org/ANM>

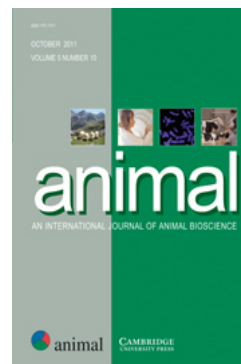
Additional services for **animal**:

Email alerts: [Click here](#)

Subscriptions: [Click here](#)

Commercial reprints: [Click here](#)

Terms of use : [Click here](#)



---

## Effect of dietary supplementation with different sources of selenium on growth response, selenium blood levels and meat quality of intensively finished Charolais young bulls

G. Cozzi, P. Prevedello, A. L. Stefani, A. Piron, B. Contiero, A. Lante, F. Gottardo and E. Chevaux

animal / Volume 5 / Issue 10 / August 2011, pp 1531 - 1538

DOI: 10.1017/S1751731111000711, Published online: 10 May 2011

**Link to this article:** [http://journals.cambridge.org/abstract\\_S1751731111000711](http://journals.cambridge.org/abstract_S1751731111000711)

### How to cite this article:

G. Cozzi, P. Prevedello, A. L. Stefani, A. Piron, B. Contiero, A. Lante, F. Gottardo and E. Chevaux (2011). Effect of dietary supplementation with different sources of selenium on growth response, selenium blood levels and meat quality of intensively finished Charolais young bulls. *animal*, 5, pp 1531-1538 doi:10.1017/S1751731111000711

**Request Permissions :** [Click here](#)

# Effect of dietary supplementation with different sources of selenium on growth response, selenium blood levels and meat quality of intensively finished Charolais young bulls

G. Cozzi<sup>1†</sup>, P. Prevedello<sup>1</sup>, A. L. Stefani<sup>2</sup>, A. Piron<sup>3</sup>, B. Contiero<sup>1</sup>, A. Lante<sup>4</sup>, F. Gottardo<sup>1</sup> and E. Chevaux<sup>3</sup>

<sup>1</sup>Dipartimento di Scienze Animali, Università degli Studi di Padova, Viale dell'Università 16, 35020 Legnaro (PD), Italy; <sup>2</sup>Istituto Zooprofilattico Sperimentale delle Venezie, Viale dell'Università 16, 35020 Legnaro (PD), Italy; <sup>3</sup>Lallemand SAS, 19 rue des Briquetiers, 31702 Blagnac, France; <sup>4</sup>Dipartimento di Biotecnologie Agrarie, Viale dell'Università 10, 35020 Legnaro (PD), Italy

(Received 21 October 2010; Accepted 8 April 2011; First published online 10 May 2011)

*The study aimed at comparing three strategies of supplementing selenium (Se) during the finishing period of Charolais young bulls: (1) administration of sodium selenite throughout the finishing (NaSe); (2) administration of an Se-enriched yeast strain (Saccharomyces cerevisiae NCYC R397) throughout the finishing (Se-Y); (3) administration of sodium selenite for 140 days replaced by Se-enriched yeast during the last 70 days of finishing (Switch). Eighty-four young bulls (mean initial BW = 434.2 ± 31.9 kg; mean age = 382 ± 52 days) were stratified by live weight and equally assigned to one of three Se treatments. Experimental groups were fed the same diets and the inclusion rate of the different treatments was targeted to achieve 0.3 mg of Se/kg of dry matter (DM) in the complete feed. The average daily gain of bulls was 1.36 kg/d and no differences due to Se treatment were recorded. Dry matter intake and feed conversion ratio were not affected by Se treatment resulting in, on average, 10.3 kg/d and 7.65, respectively. Repeated blood samples were taken at days 0, 120, 180 and 210 of finishing to assess the Se status of the animals. As compared to NaSe, both organic Se treatments (Se-Y and Switch) increased plasma Se in the last two sampling sessions according to a significant treatment × time interaction (P < 0.001). A similar trend was observed for serum total antioxidant status of the young bulls, whereas there was only a significant time effect (P < 0.001) on glutathione peroxidase activity that was raised by all Se treatments. The finishing period lasted 210 days and at the abattoir there were no differences across Se treatments in carcass weight and dressing percentage. A higher Se content in the Longissimus thoracis (LT) muscle was instead observed in Se-Y samples as compared with NaSe (0.85 v. 0.47 mg/kg DM; P < 0.05). Meat quality evaluation was carried out on LT samples after 6 and 11 days of ageing under a vacuum package. Regardless of ageing time, meat from young bulls supplemented with Se yeast had higher colour lightness (L\*) values than those receiving NaSe (38.1 v. 36.6; P < 0.01) and showed a significant decrease in shear force (3.69 v. 4.22 kg/cm<sup>2</sup>; P < 0.01). The outcomes of the study suggest that the provision of Se yeast throughout the finishing period is a strategy to increase the benefits of the replacement of sodium selenite with organic selenium in beef cattle.*

**Keywords:** selenium source, beef cattle, fattening, meat quality

## Implications

The administration of a selenium-enriched yeast strain has been shown to be an interesting alternative to inorganic selenium in the supplementation of the trace element to finishing beef cattle. As compared to sodium selenite, the supplementation with Se yeast throughout the finishing did not modify cattle growth performance, but it was a way to enrich the Se content in muscle tissue. This result along with

further positive effects induced by this treatment on meat quality, such as the increased colour lightness and the decreased drip loss and shear force, should have a favourable impact on both retailer and consumer choices.

## Introduction

Selenium is an essential trace element for humans and animals because, in the form of selenocysteine, it is a core structural component of several specific enzymes (Arteel and Sies, 2001). Antioxidant enzymes glutathione peroxidases

† E-mail: giulio.cozzi@unipd.it

(GSH-Px) containing selenium catalyse the reduction of lipid and hydrogen peroxides to less harmful products providing a defence against oxidative stress (Hardy and Hardy, 2004). Therefore, an adequate Se provision is needed to decrease the risk of immunodeficiency, myopathy, cardiovascular diseases and other Se deficiency syndromes (Hartikainen, 2005).

Selenium content in feed crops is variable and it depends to a large extent on their growing site. In Europe, the Se content in soil is quite low (Oldfield, 2002) and dietary Se supplementation of farm animals has become a common practice. So far, the demand for Se in cattle rations has been generally fulfilled by supplementing with inorganic sources such as sodium selenite or selenate. However, organic sources of Se from enriched yeast strains have been recently approved in animal nutrition by the European Union (Commission Regulation: 2006/1750/EC). Organic Se is less toxic than selenite (Brown *et al.*, 2000), which may act as a pro-oxidant at increased dietary concentrations (Seko *et al.*, 1989).

Comparisons between Se yeast and inorganic sources of Se in fattening cattle were mainly focused on the retention of the element in muscle tissue and the consequent effect on the oxidative stability of beef meat (O'Grady *et al.*, 2001; Juniper *et al.*, 2008b) as well as on the tolerance of the animals to the administration of supranutritional doses as a strategy to produce an Se-enriched beef product (Juniper *et al.*, 2008a).

According to Swecker *et al.* (1989), Se supplementation may improve animal performance indirectly by strengthening its immunocompetence and enabling it to better withstand the stressful condition of the intensive fattening systems. To verify this hypothesis, the present study compared different strategies of dietary Se supplementation during the finishing period of intensively fattened Charolais young bulls focusing on cattle growth response, Se blood levels and slaughter performance. Moreover, since the antioxidant functions of Se have been shown to persist *post mortem* delaying the onset of oxidation reactions in muscle tissue (DeVore *et al.*, 1983), the experimental protocol addressed the effects of Se treatments on a large set of meat quality traits. The experimental treatments considered the single or combined administration of two sources of the trace element (sodium selenite and a selenium-enriched yeast). To broaden the existing knowledge on the effect of Se supply in beef cattle, the adopted target concentration of trace element in the diet was chosen to be in agreement with previous studies.

## Material and methods

### *Animals and management*

The study was carried out in a commercial intensive beef fattening unit located in the town of Correzzola (PD), Italy using a single batch of 84 Charolais young bulls imported from France. This cattle breed is the main European beef breed accounting for >25% of the total population for suckling cows (Charolais, 2011). The animals had a mean initial age of  $382 \pm 52$  days and a mean initial BW of  $434.2 \pm 31.9$  kg and they were stratified by live weight and

equally assigned to one of three Se treatments. Each treatment consisted of 28 bulls housed in four pens of seven bulls each. All the pens were contiguous and located in the same barn and they had a fully slatted floor with a space allowance of 3 m<sup>2</sup>/head and a manger space of 0.6 m/head. Each pen was equipped with two waterers to allow free access to drinking water.

The three treatment groups of cattle received the same basal diets prior to and during the experimental period which started on 4 April 2009 and which was concluded on 2 November 2009, after 210 days. An initial basal diet (Table 1) was fed for 15 days preceding the onset of the experimental period to 1st August and it was then replaced by a final basal diet in which high moisture ear maize was substituted with maize meal and dried sugar beet pulp to maintain the same energy and protein contents (Table 1). The daily diet was prepared as total mixed ration (TMR) by weighing and mixing all the feed ingredients for 20 min with an AGM mixing wagon (Unifast S.p.A., Bagnoli di Sopra, Italy). The TMR was offered at 0930 h in a single administration. An *ad libitum* intake was assured by adjusting the amount of food delivered to each experimental pen in order to obtain an approximate recovery of a 3% food residue (as-fed basis) 24 h after delivery.

### *Experimental treatments*

From the start of the experimental period, young bulls were fed a given basal diet supplemented with three different sources of selenium:

- NaSe: administration of sodium selenite during the entire finishing period;
- Se-Y: administration during the entire finishing period of a selenised yeast [Alkosel<sup>®</sup> R397 produced by a specific strain of *Saccharomyces cerevisiae* (NCYC R397), manufactured and supplied by Lallemand SAS (Blagnac, France)];
- Switch: administration of sodium selenite for 140 days replaced by Alkosel<sup>®</sup> R397 during the last 70 days of the finishing period.

The last treatment aimed at evaluating the effects of the provision of selenium yeast as an alternative to inorganic Se for a short time period, as the last part of the finishing process. The inclusion rate of the Se sources (sodium selenite or Alkosel<sup>®</sup> R397) in all treatments was targeted to achieve 0.3 mg of Se/kg of dry matter (DM) in the complete feed. This concentration was chosen to be consistent with previous studies on dietary supplementation with the trace element in beef cattle (Nicholson *et al.*, 1991; O'Grady *et al.*, 2001; Juniper *et al.*, 2008b),

In order to promote a balanced intake of Se among all the pen-mates, the two Se sources were separately pre-mixed with dried grape seeds (Table 1). An amount of 0.3 kg/head of a given Se pre-mixture was top-dressed and then thoroughly mixed manually with the basal TMR delivered into each pen manger of the corresponding Se treatment at the time of feed distribution.

**Table 1** Ingredient and chemical composition of the basal total mixed rations fed during the finishing period and chemical composition of dried grape seeds used for selenium administration

	Total mixed ration		Dried grape seeds
	Initial	Final	
Ingredient (g/kg fresh weight basis)			
Maize silage	472	453	
Maize meal	74	200	
High moisture ear maize	215	–	
Dried sugar beet pulp	–	112	
Rice middlings	59	57	
Maize gluten feed	59	57	
Soybean meal	32	34	
Wheat straw	47	45	
Protein–mineral–vitamin supplement <sup>†</sup>	29	29	
Hydrolysed fat	12	13	
Chemical composition			
DM (g/kg)	534	575	895
CP (g/kg DM)	131	130	132
Ether extract (g/kg DM)	56	60	50
Ash (g/kg DM)	64	62	77
Starch (g/kg DM)	322	328	33
Neutral-detergent fibre (g/kg DM)	324	317	747
Selenium (mg/kg DM)	0.04	0.05	0.06
<i>Unité Fouragère Viande</i> (/kg DM)	0.93	0.94	0.14

DM = dry matter.

<sup>†</sup>Contained per kg of supplement: CP 260 g; ether extract 35 g; ash 380 g; neutral-detergent fibre 123 g; Ca 80 g; Na 33 g; P 45 g; Mg 18 g; urea 40 g; Zn 1200 mg; Mn 960 mg; Fe 360 mg; Cu 200 mg; I 16 mg; Co 10 mg; vitamin A 80 350 UI; vitamin D3 10 500 UI;  $\alpha$ -tocopherol 180 mg.

### Measurements

The individual weight of bulls was recorded in five weighting sessions during the experimental period: at the start of the finishing period (day 0), at days 60, 120 and 180 and at the end of the finishing period (day 210). Average daily gain (ADG) was measured by dividing the difference between final and initial live weight by the number of days of the period. In order to better estimate the effect of the substitution of NaSe with Se-Y in the Switch treatment, the daily gain of bulls was analysed for the whole fattening period and separately for the two sub-periods from 0 to 120 days and from 121 to 210 days. Pen dry matter intake (DMI) was recorded three times a week during the experimental fattening period by weighing the feed distributed and the residue in the manger 24 h later. Pen feed conversion ratio was calculated by dividing DMI by the mean ADG of the pen-mates. Individual blood samples were collected at days 0, 120, 180 and 210 of the fattening period from the same four bulls/pen randomly chosen among the pen-mates. In this way, data from 16 bulls/treatment were available at each sampling session. Samples were taken from the jugular vein and they consisted of two 5-ml Vacutainer<sup>®</sup> tubes (Becton Dickinson Italia, Buccinasco, Italy) filled per animal: one tube with lithium heparin for Se and GSH-Px analysis and one tube without anticoagulant for serum total antioxidant status determination (TAS). Glutathione peroxidase is an Se-containing enzyme that protects body tissues from oxidative

damage by removing peroxides resulting from free radicals, whereas TAS is a simple assay to estimate the total anti-oxidative capacity in serum. After 210 days of finishing, all animals were slaughtered at the same EU-approved abattoir which had routine veterinary inspection. Carcasses were weighed and their dressing proportion was calculated as (hot carcass weight/slaughter live weight)  $\times$  100. Subsequently, the carcasses were chilled and stored at 2 to 4°C and two sample joints of the muscle *Longissimus thoracis* (LT) were excised from the fifth to the seventh ribs of each right half carcass from a random sample of four bulls/pen 24 h *post mortem*. These samples were weighed, packaged in a vacuum atmosphere and then stored for 5 and 10 days, respectively, at 4°C in a chilling room for a total ageing period of 6 and 11 days *post mortem*. Meat samples were blot dried and weighed again after the two ageing periods, and drip losses were calculated as the difference between the weight prior to vacuum storage and that at the end of the ageing period. Subsequently, meat pH was measured with a portable pH-meter (HANNA Instruments<sup>®</sup>, Inc., Woonsocket, RI, USA) equipped with a glass electrode (3 mm  $\varnothing$  conic tip) suitable for meat penetration. Instrumental colour determination was carried out on a fresh, unoxidised surface of the samples after exposure for 1 h to air at 2°C (Boccard *et al.*, 1981). A CR 100 Chromameter (Minolta Camera, Osaka, Japan) equipped with C illuminant was used and colour data were expressed according to the Hunter-Lab system.

Weight cooking losses were determined on 2.5 cm-thick steaks heated in a water bath at 75°C for 50 min and cooled in running tap water for at least 40 min (Boccard *et al.*, 1981). Ten meat cylindrical cores, 1.25 cm in diameter, were then excised from the cooked steak for the instrumental measurement of meat tenderness carried out using a Warner-Bratzler shear force meter (Instron, High Wycombe, UK; Joseph, 1979). A further sub-sample of LT was collected from the same four bulls/pen 24 h *post mortem* to be freeze-dried and stored for meat composition analysis.

#### Chemical analysis

Samples of the experimental diet were collected every month and chemically analysed for DM, CP, ether extract, ash and starch according to the methods of the Association of Official Analytical Chemists (AOAC, 1990). Neutral-detergent fibre analysis of the same samples was conducted according to Van Soest *et al.* (1991). The net energy content of the diet measured in *Unité Fouragère Viande* units was calculated using for all feed ingredients the reference values reported by the Institut National de la Recherche Agronomique (INRA, 1988). The meat protein, ash and intramuscular fat contents were measured according to the Association of Official Analytical Chemists (1990) after grinding the freeze-dried samples. Total Se in TMR, plasma and meat samples was determined after sample mineralisation by using inductivity coupled plasma/mass spectrometry (Agilent Technologies, Palo Alto, CA, USA) with an instrumental detection limit of 50 ng/l. Serum TAS and blood GSH-Px were determined by commercial kits (Randox Laboratories, Crumlin, UK) applied to a BM Hitachi 911 analyzer (ROCHE, Basel, Switzerland). Meat sub-samples from the joints stored for the two ageing periods were submitted to a rapid aqueous acid extraction thiobarbituric acid method for measuring malondialdehyde (MDA) as a marker of lipid peroxidation in muscle tissue (Botsoglou *et al.*, 1994).

#### Statistical analysis

Pen was the experimental unit for live weight of bulls, ADG, feed intake and feed conversion ratio and these data were statistically analysed adopting a linear model which considered the effects of treatment (source of Se) and pen within treatment (random effect). The same linear model was used for the processing of carcass measurements and meat chemical composition data. A repeated mixed model which considered the effects of treatment, time (sampling session), pen within treatment and a random effect of animal within the pen was used for processing blood variables recorded at different sampling sessions as well as meat quality data recorded after the two ageing periods. In all the statistical analyses, the 2 degrees of freedom of the treatment factor were used to perform the following set of orthogonal contrasts:

1. NaSe *v.* Others [(Se-Y + Switch)/2];
2. Se-Y *v.* Switch.

Differences were considered significant at  $P < 0.05$ . Overall and partial correlation coefficients between plasma Se and GSH-Px as well as between TAS and meat Se content were

calculated. All the statistical analyses were carried out using the SAS package (SAS, 2001).

## Results

Despite the changes in the feed ingredients, the chemical composition of the two diets was constant throughout the fattening period (Table 1). Monthly samples collected during the progress of the experiment showed an Se content of  $0.29 \pm 0.04$  mg/kg of DM (mean  $\pm$  s.d.) in the complete feed for NaSe treatment, whereas the mean value for the diet supplemented with Se yeast was  $0.28 \pm 0.07$  mg/kg of DM.

#### Growth performance

No differences due to Se treatment were recorded for ADG and final live weight of bulls (Table 2). Regardless of Se treatment, the animals tended to reduce their growth rate in the second part of the fattening period. Consistent with the daily gain, DMI and feed conversion ratio of bulls were not affected by the different Se sources (Table 2).

#### Blood selenium status

Mean values recorded for plasma Se in the three treatments at different sampling times are shown in Figure 1a. The statistical analysis of this variable showed a significant treatment  $\times$  time interaction ( $P < 0.001$ ). The supplementation with the trace element promoted a general increase in plasma Se concentration during the first 120 days. Afterwards, plasma Se showed a steady state in NaSe bulls, whereas a further increase was observed in both groups of animals receiving organic Se (Se-Y and Switch).

According to a significant time effect ( $P < 0.001$ ), the concentration of GSH-Px was sensitive to Se supplementation, but no treatment effect was observed for this variable (Figure 1b). Selenium supplementation promoted a significant increase in GSH-Px activity during the first 120 days of finishing. The enzyme concentration slowly decreased in the final sampling session, but it always remained significantly higher than the initial baseline (day 0). There was a good overall correlation between plasma Se and GSH-Px concentration ( $r = 0.63$ ;  $P < 0.001$ ), and the calculation of partial correlation coefficients showed similar  $r$  values for the different Se treatments (NaSe:  $r = 0.72$ ,  $P < 0.001$ ; Se-Y:  $r = 0.67$ ,  $P < 0.001$  and Switch:  $r = 0.60$ ,  $P < 0.001$ ).

Dietary Se supplementation also increased serum TAS, but the response observed at the following sampling sessions was not consistent across treatments according to a significant treatment  $\times$  time interaction ( $P < 0.001$ ) (Figure 1c). As compared to the day 0 values, only bulls that received organic Se during the whole finishing process (Se-Y) or in its final part (Switch) showed a significant increase in serum TAS at all the following sampling sessions (Figure 1c). Overall correlation between plasma Se and TAS ( $r = 0.41$ ;  $P < 0.001$ ) was significant but lower than that with GSH-Px. Correlation coefficients between plasma Se and TAS for the three Se treatments showed that the significance was substantially due to provision

**Table 2** Effect of different strategies of dietary selenium supplementation during the finishing period on growth performance and feed consumption of Charolais young bulls

	Selenium treatment			Orthogonal contrast		s.e.
	NaSe	Se-Y	Switch	1.	2.	
<b>Growth performance</b>						
Observations (n)	28	28	28			
Live weight (kg)						
Day 0	432.6	435.6	436.5	ns	ns	13.0
Day 120	612.1	605.6	598.6	ns	ns	15.9
Day 210	718.8	721.4	719.9	ns	ns	18.1
Average daily gain (kg/d)						
0 to 120 days	1.50	1.41	1.35	ns	ns	0.07
121 to 210 days	1.19	1.28	1.34	ns	ns	0.07
0 to 210 days	1.36	1.37	1.35	ns	ns	0.06
<b>Feed consumption</b>						
Observations (n)	4	4	4			
Dry matter intake (kg/d)	10.22	10.56	10.65	ns	ns	0.37
Feed conversion ratio	7.53	7.98	8.07	ns	ns	0.44

NaSe = sodium selenite during the entire finishing; Se-Y = Alkoseal<sup>®</sup> R397 during the entire finishing period; Switch = sodium selenite for 140 days replaced by Alkoseal<sup>®</sup> R397 during the last 70 days of finishing.

1. Contrast NaSe v. Others [(Se-Y + Switch)/2].

2. Contrast Se-Y v. Switch.

ns  $P > 0.05$ .

of the Se yeast (NaSe:  $r = 0.20$ ,  $P > 0.05$ ; Se-Y:  $r = 0.49$ ,  $P < 0.001$  and Switch:  $r = 0.29$ ,  $P < 0.05$ ).

#### Slaughter performance and meat quality

Consistent with the growth performance, at the abattoir, there were no differences due to the Se treatment in carcass weight and dressing percentage (Table 3). The protein, fat and ash content of the LT muscle did not show any significant effect due to Se treatment. However, a significant difference ( $P < 0.01$ ) due to the source of the trace element was instead observed for Se content in muscle tissue (Table 3). Compared with NaSe, higher Se concentrations were recorded in meat samples taken from Se-Y bulls, whereas Switch treatment resulted in intermediate concentrations. There was a significant overall correlation between plasma Se and Se content in muscle tissue ( $r = 0.41$ ;  $P < 0.001$ ). However, as for TAS, the observed significance was substantially due to the two Se treatments using the organic source of the trace element (NaSe:  $r = 0.20$ ,  $P > 0.05$ ; Se-Y:  $r = 0.48$ ,  $P < 0.001$  and Switch:  $r = 0.30$ ,  $P < 0.05$ ).

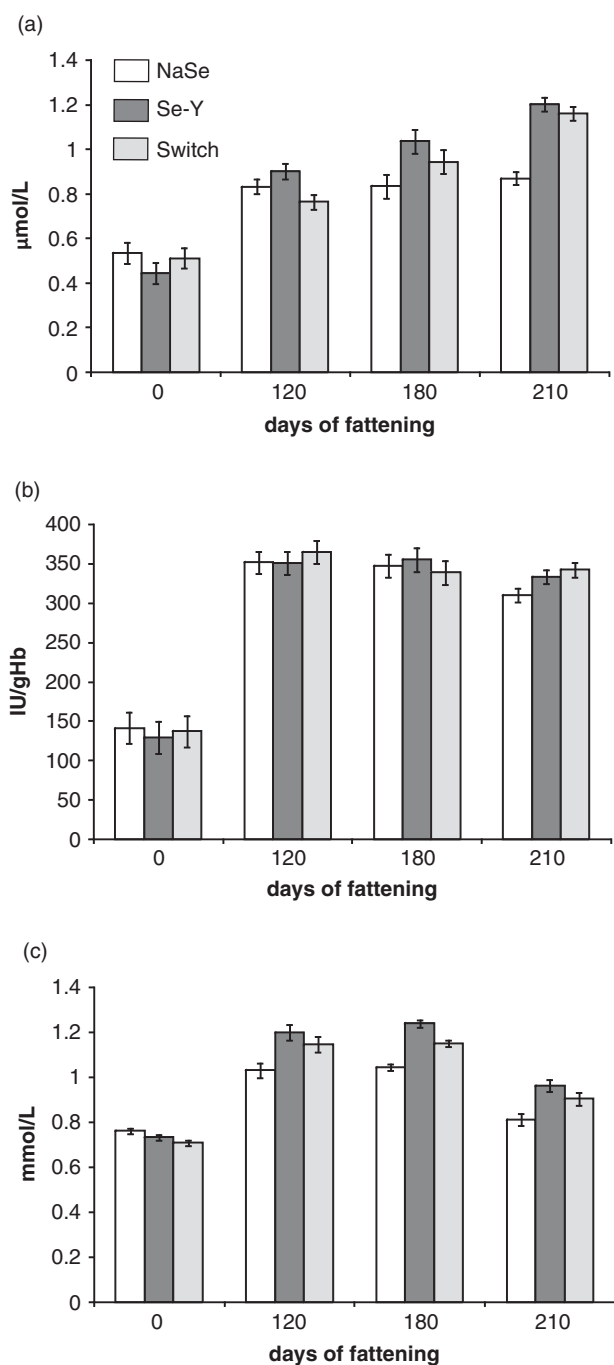
The outcomes of the quality evaluation carried out on meat samples after 6 and 11 days of ageing under vacuum package are reported in Table 4. A significant treatment  $\times$  ageing interaction was observed for drip loss and pH. Meat drip loss increased according to the time of ageing, but this trend was less marked in Se-Y samples. Meat pH was always within the range of normality. The instrumental colour determination showed a significant treatment effect on meat lightness ( $P < 0.01$ ). Regardless of the ageing days, meat from bulls fed Se yeast had higher  $L^*$  values than that from NaSe bulls. The levels of MDA measured after 6 days of

ageing were always below the detection threshold of the spectrophotometer (0.001 mg of MDA/kg), whereas mean values recorded after 11 days of ageing were very low and similar across Se treatments (NaSe = 0.007; Se-Y = 0.007 and Switch = 0.011 mg of MDA/kg). Meat cooking losses were not affected by Se treatment while they increased according to the ageing days ( $P < 0.05$ ). As expected, prolonged ageing had a positive effect on meat tenderness as shown by the reduced shear force ( $P < 0.001$ ). On this parameter, there was also an Se treatment effect ( $P < 0.01$ ) because meat samples from bulls supplemented with Se yeast required a reduced shear force in comparison to NaSe ones.

#### Discussion

The study compared different strategies of supplementing Se during the finishing period of intensively finished Charolais young bulls by using two sources of the trace element (sodium selenite and Se yeast). The growth performance and slaughter traits of bulls were not affected by the different sources of Se supplemented during the finishing period, confirming previous results recorded in growing and finishing cattle (Nicholson *et al.*, 1991; Lawler *et al.*, 2004).

As compared with the values measured at day 0, all Se treatments increased plasma Se concentration. However, as in previous studies (Awadeh *et al.*, 1998; Gunter *et al.*, 2003), the provision of Se yeast further increased the bio-availability of the trace element resulting in higher plasma Se concentrations than NaSe at the end of the fattening period. While supplementing fattening steers with a similar target



**Figure 1** Effect of selenium supplementation with NaSe, Se-Y or Switch during the finishing period on (a) plasma selenium concentration; (b) glutathione peroxidase activity and (c) total antioxidant status of Charolais young bulls (mean  $\pm$  s.e.).

level of Se adopted in the present study, Juniper *et al.* (2008b) found that plasma Se concentration would achieve a steady state after approximately 150 days of treatment, but the asymptotic value obtained with the provision of an organic source was significantly higher than that of sodium selenite.

Consistent with Van Ryssen *et al.* (1989) and Gunter *et al.* (2003), no significant differences in GSH-Px activity were observed in animals that had been offered diets supplemented with comparable doses of inorganic Se and Se yeast. However,

a better status of animals receiving Se yeast was indicated by the higher values of TAS observed for Se-Y and Switch treatments in the final part of the finishing process. This integrated parameter, which considers the cumulative action of all the antioxidants present in plasma, is more sensitive and reliable than the measure of a single antioxidant and it may be a useful indicator to assess animal welfare (Celi, 2010).

The bimodal trend observed for GSH-Px and TAS in all the Se treatments might be the result of a temporary pro-oxidant behaviour caused by a prolonged supplementation with the trace element. However, as reported by Halliwell (2008) and Mézes and Balogh (2009), the pro-oxidant activity of Se may not be detrimental because it provides a useful cytoprotection effect against infective agents.

As for farm animals, selenium is an essential trace mineral for humans and Se deficiency is still a global problem in many countries requiring specific strategies to produce Se-rich foods (Fisinin *et al.*, 2009). Beef is a major source of dietary selenium for humans (Zhang *et al.*, 2010) and the result of the present study indicated that the prolonged supplementation with organic Se (Se-Y) is more effective to increase Se concentration in muscle tissue than inorganic Se. The efficacy of Se yeast to enrich the trace mineral content in meat has been reported not only in beef (Juniper *et al.*, 2008b) but also in veal (Skřivanová *et al.*, 2007), lamb (Vignola *et al.*, 2009), chicken (Skřivan *et al.*, 2008) and pork (Mahan *et al.*, 1999). According to Juniper *et al.* (2008b), the greater concentration of Se in the muscle tissue of bulls offered organic Se when compared with NaSe might arise from an enhanced availability and tissue retention of the trace element. Both Se sources can be incorporated into GSH-Px, but Se-methionine from the organic one is also non-specifically incorporated into body proteins as a replacement for methionine (Vignola *et al.*, 2009). In this regard, Juniper *et al.* (2008b) reported a higher concentration of Se in almost all tissues of beef cattle fed Se yeast than selenite as the result of a greater proportion of total Se comprised in tissue protein as Se-methionine. Besides the enrichment of beef and beef products, the increased Se muscular content promoted by the organic source of trace element could also be beneficial for the animal. Muscle tissue acts as an important endogenous reserve of the trace element being utilised in case of periods of Se deficiency (Vignola *et al.*, 2009).

The provision of Se yeast during the whole finishing process led to lower drip losses in meat aged under vacuum package for 11 days according to a treatment  $\times$  ageing interaction. This positive effect of organic Se on beef drip loss is consistent with that observed in chicken meat from broilers supplemented with organic Se rather than selenite (Edens, 1996), whereas in pork meat a similar trend was close to the threshold of statistical significance (Mahan *et al.*, 1999). The effect of Se on muscle drip loss could arise from a decreasing cell membrane oxidation (Dunshea *et al.*, 2005).

The ageing of the meat samples under vacuum package did not allow us to verify whether the different source of selenium was capable of reducing lipid peroxidation. However, O'Grady *et al.* (2001) suggested that dietary Se has limited

**Table 3** Effect of different strategies of dietary selenium supplementation during the finishing period on carcass traits and chemical composition of Longissimus thoracis muscle of Charolais young bulls

	Selenium treatment			Orthogonal contrast		s.e.
	NaSe	Se-Y	Switch	1.	2.	
<b>Carcass traits</b>						
Observations (n)	28	28	28			
Weight (kg)	430.7	440.5	428.2	ns	ns	14.5
Dressing proportion (%)	59.9	60.9	59.4	ns	ns	0.60
<b>Meat chemical composition</b>						
Observations (n)	16	16	16			
CP (g/kg DM)	861	862	861	ns	ns	10
Intramuscular fat (g/kg DM)	96	95	98	ns	ns	10
Ash (g/kg DM)	43	43	41	ns	ns	6
Selenium (mg/kg DM)	0.47 <sup>b</sup>	0.85 <sup>a</sup>	0.58 <sup>ab</sup>	**	ns	0.07

NaSe = sodium selenite during the entire finishing; Se-Y = Alkosel<sup>®</sup> R397 during the entire finishing period; Switch = sodium selenite for 140 days replaced by Alkosel<sup>®</sup> R397 during the last 70 days of finishing; DM = dry matter.

1. Contrast NaSe v. Others [(Se-Y + Switch)/2].

2. Contrast Se-Y v. Switch.

<sup>a,b</sup>Mean values within a row with different superscript letters are significantly different at  $P < 0.05$ .

ns  $P > 0.05$ ; \*\* $P < 0.01$ .

**Table 4** Effect of different strategies of dietary selenium supplementation during the finishing period on quality traits of Longissimus thoracis muscle of Charolais young bulls after 6 and 11 days of ageing post mortem

	Ageing days (A)	Selenium treatment (S)			Significance			s.e.
		NaSe	Se-Y	Switch	A	S	A × S	
Observations (n)		16	16	16				
Drip loss (%)	6	0.88 <sup>c</sup>	0.87 <sup>c</sup>	0.90 <sup>c</sup>	***	**	***	0.04
	11	1.63 <sup>a</sup>	1.22 <sup>b</sup>	1.38 <sup>ab</sup>				
pH	6	5.51 <sup>b</sup>	5.60 <sup>a</sup>	5.51 <sup>b</sup>	ns	***	**	0.01
	11	5.53 <sup>b</sup>	5.57 <sup>ab</sup>	5.56 <sup>ab</sup>				
<b>Colour</b>								
Lightness ( $L^*$ )	6	36.0	37.7	38.3	ns	***†	ns	0.39
	11	37.1	38.0	38.5				
Redness ( $a^*$ )	6	14.9	15.4	15.0	ns	ns	ns	0.37
	11	15.7	15.4	14.9				
Yellowness ( $b^*$ )	6	14.9	14.6	14.9	ns	ns	ns	0.28
	11	15.5	14.5	15.6				
Cooking loss (%)	6	33.5	34.6	34.9	*	ns	ns	0.49
	11	35.4	35.9	35.0				
Shear force (kg/cm <sup>2</sup> )	6	4.43	3.88	4.03	***	***†	ns	0.14
	11	4.00	3.32	3.52				

NaSe = sodium selenite during the entire finishing; Se-Y = Alkosel<sup>®</sup> R397 during the entire finishing period; Switch = sodium selenite for 140 days replaced by Alkosel<sup>®</sup> R397 during the last 70 days of finishing.

<sup>a,b,c</sup>Mean values within a variable having different superscript letters differ at  $P < 0.05$ .

ns  $P > 0.05$ ; \* $P < 0.05$ ; \*\* $P < 0.01$ ; \*\*\* $P < 0.001$ .

†Contrast NaSe v. Others [(Se-Y + Switch)/2] significant at  $P < 0.05$ .

potential for increasing the oxidative stability of beef. Beef lightness is unrelated to myoglobin chemical status changes (McKenna *et al.*, 2005), but it is closely related to muscle and protein structures (MacDougall, 1982). Therefore, the increased meat lightness and the decreased shear force observed in samples from bulls receiving organic Se could arise from changes in muscle tissue most likely promoted by the increased accumulation of selenoamino acids. Regardless of the Se treatment, the prolonged ageing time of meat

samples in a vacuum atmosphere led to a decreased shear force confirming the results of Lindahl *et al.* (2010). Consistent with the same authors, the prolonged ageing period also increased beef cooking losses.

## Conclusions

In comparison with an inorganic source of Se, the supplementation of the finishing diet with Se yeast targeted to



achieve 0.3 mg of Se/kg of DM in the complete feed did not affect the growth and slaughter performance of bulls. However, the provision of organic Se throughout the finishing period improved the Se status of the young bulls by increasing the Se content and TAS of blood. In addition, Se-Y treatment was a way to increase the Se content in muscle tissue and to reduce meat drip loss and shear force. The replacement of sodium selenite with organic selenium in the last part of the finishing process led to similar advantages, except for the enrichment of the trace mineral content in meat.

## Acknowledgements

The authors are grateful to the personnel of the Guzzo farm for the logistic and practical support to the research. A very special thanks to the farm vet Dr. Pierluigi Zuanon and to Dr. Daniele Dalle Palle, Mr. Roberto Chimento and Mr. Roberto Rampazzo for their skilled technical assistance.

## References

- Arteel GE and Sies H 2001. The biochemistry of selenium and the glutathione system. *Environmental Toxicology and Pharmacology* 10, 153–158.
- AOAC (Association of Official Analytical Chemists) 1990. Official methods of analysis, 15th edition. AOAC, Washington, DC, USA.
- Awadeh FT, Kincaid RL and Johnson KA 1998. Effect of level and source of dietary selenium on concentrations of thyroid hormones and immunoglobulins in beef cows and calves. *Journal of Animal Science* 76, 1204–1215.
- Boccard R, Buchter L, Casteels E, Cosentino E, Dransfield E, Hood DE, Joseph RL, McDougall DB, Rhodes DN, Schön I, Timbergen BJ and Touraille C 1981. Procedures for measuring meat quality characteristics in beef production experiments. Report of a Working Group in the Commission of the European Communities (CEC) beef production research program. *Livestock Production Science* 8, 385–397.
- Botsoglou NA, Fletouris DJ, Papageorgiou GE, Vassilopoulos VN, Mantis AJ and Trakatellis AG 1994. Rapid, sensitive and specific thiobarbituric acid method for measuring lipid peroxidation in animal tissue, food and feedstuff samples. *Journal of Agricultural and Food Chemistry* 42, 1931–1937.
- Brown KM, Pickard K, Nicol F, Beckett GJ, Duthie GG and Arthur JR 2000. Effects of organic and inorganic selenium supplementation on selenoenzyme activity in blood lymphocytes, granulocytes, platelets and erythrocytes. *Clinical Science* 98, 593–599.
- Celi P 2010. The role of oxidative stress in small ruminants' health and production. *Revista Brasileira de Zootecnia* 39, 348–363.
- Charolais 2011. Le Charolaise. Chiffres clés. Retrieved March 31, 2011, from <http://www.charolaise.fr/>
- DeVore VR, Colnago GL, Jensen LS and Breene BE 1983. Thiobarbituric acid values and glutathione peroxidase activity in meat from chickens fed a Se supplemented diet. *Journal of Food Science* 48, 300–301.
- Dunshea FR, D'Souza DN, Pethick DW, Harper GS and Warner RD 2005. Effects of dietary factors and other metabolic modifiers on quality and nutritional value of meat. *Meat Science* 71, 8–38.
- Edens FW 1996. Organic selenium: from feathers to muscle integrity to drip loss. Five years onwards: no more selenite!. In *Biotechnology in the feed industry, Proceedings of the 12th Annual Symposium* (ed. TP Lyons and KA Jacques), pp. 165–185. Nottingham University Press, Nottingham, UK.
- Fisinin V, Tigran T and Surai PF 2009. Producing selenium-enriched eggs and meat to improve the selenium status of the general population. *Critical Reviews in Biotechnology* 29, 18–28.
- Gunter SA, Beck PA and Phillips JM 2003. Effects of supplementary selenium source on the performance of blood measurements in beef cows and their calves. *Journal of Animal Science* 81, 856–864.
- Halliwel B 2008. Are polyphenols antioxidants or pro-oxidants? What do we learn from cell culture and *in vivo* studies? *Archives of Biochemistry and Biophysics* 476, 107–112.
- Hardy G and Hardy I 2004. Selenium: the Se-XY nutraceutical. *Nutrition* 20, 590–593.
- Hartikainen H 2005. Biochemistry of selenium and its impact on food chain quality and human health. *Journal of Trace Elements in Medicine and Biology* 18, 309–318.
- INRA (Institut National de la Recherche Agronomique) 1988. Alimentation des bovins, ovins et caprins. INRA, Paris, France.
- Joseph RL 1979. Recommended method for assessment of tenderness. In *The future of beef production in the European community* (ed. JC Bowman and P Susmel), pp. 596–606. Martinus Nijhoff, The Hague, The Netherlands.
- Juniper DT, Phipps RH, Givens DJ, Jones AK, Green C and Bertin G 2008a. Tolerance of ruminant animals to high dose in-feed administration of a selenium-enriched yeast. *Journal of Animal Science* 86, 197–204.
- Juniper DT, Phipps RH, Ramos-Morales E and Bertin G 2008b. Effect of dietary supplementation with selenium-enriched yeast or sodium selenite on selenium tissue distribution and meat quality in beef cattle. *Journal of Animal Science* 86, 3100–3109.
- Lawler TL, Taylor JB, Finley J and Wand Caton JS 2004. Effect of supranutritional and organically bound selenium on performance, carcass characteristics, and selenium distribution in finishing steers. *Journal of Animal Science* 82, 1488–1493.
- Lindahl G, Lagerstedt Å, Ertbjerg P, Sampels S and Lundström K 2010. Ageing of large cuts of beef loin in vacuum or high oxygen modified atmosphere – effect on shear force, calpain activity, desmin degradation and protein oxidation. *Meat Science* 85, 160–166.
- MacDougall DB 1982. Changes in the color and opacity of meat. *Food Chemistry* 9, 75–88.
- Mahan DC, Cline TR and Richert B 1999. Effects of dietary levels of selenium-enriched yeast and sodium selenite as selenium sources fed to growing-finishing pigs on performance, tissue selenium, serum glutathione peroxidase activity, carcass characteristics, and loin quality. *Journal of Animal Science* 77, 2172–2179.
- McKenna DR, Mies PD, Baird BE, Pfeiffer KD, Ellebracht JW and Savell JW 2005. Biochemical and physical factors affecting discoloration characteristics of 19 bovine muscles. *Meat Science* 70, 665–682.
- Mézes M and Balogh K 2009. Prooxidant mechanisms of selenium toxicity – a review. *Acta Biologica Szegediensis* 53 (suppl. 1), 15–18.
- Nicholson JWG, McQueen RE and Bush RS 1991. Response of growing cattle to supplementation with organically bound or inorganic sources of selenium or yeast cultures. *Canadian Journal of Animal Science* 71, 803–811.
- O'Grady MN, Monohan FJ, Fallon RJ and Allen P 2001. Effects of dietary supplementation with vitamin E and organic selenium on the oxidative stability of beef. *Journal of Animal Science* 79, 2827–2834.
- Oldfield JE 2002. Selenium world atlas (2002 updated edition). Selenium-Tellurium Development Association, Grimbergen, Belgium.
- SAS (Statistical Analysis Systems Institute) 2001. User's guide: statistics, version 6. SAS Institute Inc., Cary, NC, USA.
- Seko Y, Saito Y, Kitahara J and Imura N 1989. Active oxygen generation by the reaction of selenite with reduced glutathione *in vitro*. In *Selenium in biology and medicine* (ed. A Wendel), pp. 33–70. Springer-Verlag, Berlin, Deutschland.
- Skrivan M, Marounek M, Dlouha G and Sevcikova S 2008. Dietary selenium increases vitamin E contents of egg yolk and chicken meat. *British Poultry Science* 49, 482–486.
- Skřivanová E, Marounek M, De Smet S and Raes K 2007. Influence of dietary selenium and vitamin E on quality of veal. *Meat Science* 76, 495–500.
- Swecker WS Jr, Eversole DE, Thatcher CD, Blodgett DJ, Schrig GG and Meldrum JB 1989. Influence of supplemental selenium on humoral immune response in weaned beef calves. *American Journal of Veterinary Research* 50, 1760–1763.
- Van Ryssen JBJ, Deagen JT, Beilstein MA and Whanger PD 1989. Comparative metabolism of organic and inorganic selenium by sheep. *Journal of Agricultural and Food Chemistry* 37, 1358–1363.
- Van Soest PJ, Robertson JB and Lewis BA 1991. Method for dietary fiber, neutral detergent fiber and nonstarch polysaccharides in relation to animal nutrition. *Journal of Dairy Science* 74, 3583–3597.
- Vignola G, Lambertini L, Mazzone G, Giammarco M, Tassinari M, Martelli G and Bertin G 2009. Effects of selenium source and level of supplementation on the performance and meat quality of lambs. *Meat Science* 81, 678–685.
- Zhang W, Xiao S, Samaraweera H, Lee EJ and Ahn DU 2010. Improving functional value of meat products. *Meat Science* 86, 15–31.