

Differential dynamics of Epstein-Barr virus in individuals infected with human immunodeficiency virus-1 receiving intermittent interleukin-2 and antiretroviral therapy

Nicoletta Burighel Silvia Ghezzi Silvia Nozza Paola Del Bianco Adriano Lazzarin Giuseppe Tambussi Guido Poli Anita De Rossi	ghel Interleukin-2 (IL-2) increases circulating CD4 ⁺ lymphocytes in patients infected with human immunodeficiency virus-1. We studied Epstein-Barr virus (EBV) dynamics in 4 patients treated with antiretroviral therapy (ART) plus different IL-2 regimens. EBV-DN tended to increase in both peripheral blood cells and plasma after continous infusion for lowed by intermittent subcutaneous <i>high-dose</i> IL-2, while EBV-DNA decreased in cel (<i>p</i> =0.0078) and disappeared in plasma after intermittent subcutaneous <i>low-dose</i> IL-2 i (<i>p</i> =0.0184) and plasma (<i>p</i> =0.0114). Thus, as a function of dose, IL-2 therapy may sign icantly affect the dynamics of EBV infection.					
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The effects of IL-2 therapy on the dynamics of EBV infection are unknown. Several IL-2-induced cytokines, such as tumor necrosis factor- α and β and IL-6, may stimulate the proliferation of B cells, thus leading to expansion of EBV-positive B cells.7-9 Furthermore, IL-2 can directly stimulate the proliferation of B cells from HIV-1-infected individuals.¹⁰ In addition, B-cell stimulation and EBV-DNA load were shown to increase in patients with a significant gain in CD4⁺ lymphocytes but incomplete suppression of HIV-1 plasma viremia during ART." Because of the high incidence of HIV-1- and EBVcoinfected patients, we investigated the impact of different IL-2 therapeutic regimens on EBV-DNA load in peripheral blood

mples were ized for 12 nent groups: IL-2 (continof 12 million /day for 2 followed by twice daily oup B: ART twice daily oup C: ART ice daily for up D: ART ens and clincteristics of elsewhere.2 Fatients enrolled in group C showed approximately half the adverse effects associated with IL-2 toxicity than did patients enrolled in groups A or B². PBMC and plasma samples, obtained at study entry (baseline) and at study completion after 12 months (posttherapy), were cryopreserved at -80°C.

Quantification of EBV-DNA. The EBV in cells and plasma was quantified by a realtime quantitative polymerase chain reaction, exactly as detailed elsewhere." EBV load was expressed as EBV-DNA copies/10⁵ cells. A conversion factor of 25x was used to estimate the number of EBV-DNA copies/mL of plasma.11

Statistical analysis. Baseline CD4⁺ cell counts, HIV-1 RNA plasma viremia and EBV-DNA loads in cells and plasma were compared pairwise between groups by the Mann-Whitney test. Changes within groups were estimated using the Wilcoxon's signed-rank test, and between groups using the Mann-Whitney test. Both within and between group comparisons for EBV in cells and plasma were also stratified according to the immunological response of ART-treated patients. An immunological response was defined as an increase >30% from baseline in the CD4⁺ cell count, with an absolute value >100 cells/ μ L.¹¹ Arbitrary values were attributed to plasma samples with undetectable EBV levels to include them in the statistical analyses: similar results were obtained using either 0 or 25 copies/mL as the arbitrary value. All p values were based on two-sided testing, and statistical analyses were carried out with SAS statistical software (Release 8.02; SAS Institute, Cary, NC, USA, USA).

Results and Discussion

At baseline, the number of $CD4^+$ cells/µL and values of HIV-1 plasma viremia were not significantly different among the four arms of the trial. All patients were positive for EBV-DNA in PBMC, with a mean viral load of 311 (range, 2-2,294) copies/10⁵ cells. Eight patients also had EBV-DNA detectable in plasma (range, 59-620 copies/mL). Neither cell nor plasma EBV values differed significantly among the groups (Table 1). All patients were on stable ART based on two non-nucleoside reverse transcriptase inhibitors at study entry. A protease inhibitor was added to the pre-existing regimens at the beginning of the study.² Over the treatment period, HIV-1 plasma viremia decayed in most patients, but this decrease was statistically significant (p=0.0244)only in those treated with high-dose IL-2 (Figure 1A, panel B). In spite of persistent HIV-1 plasma viremia, the number of CD4⁺ lymphocytes increased significantly in all IL-2-treated patients. Changes (± standard error) of CD4⁺ cells from baseline were +681 (\pm 153) cells/ μ L for the *civ/high-dose* IL-2 arm (p=0.0039), +819 (±146) cells/ μ L for the *high-dose* IL-2 arm (*p*= 0.0010), and +795 (± 160) cells/µL for the low-dose IL-2 arm (p=0.0078) (Figure 1A, panels A, B, and C). These increases were significantly higher than those observed in patients receiving ART alone (Figure 1B), in whom CD4⁺ lymphocytes only increased from 353 (±29) to 446 (±43) cells/ μ L (+93 ± 35 cells/ μ L) (Figure 1A, panel D). In particular, only six of 12 ART-treated patients showed a gain in CD4⁺ lymphocytes (i.e., immunological responders); in the others, CD4⁺ cell counts remained fairly stable or decreased (Figure 1A, panels D1 and D2). In agreement with a previous study," EBV-DNA levels increased from baseline (+815±361 copies/10⁵ cells; p=0.0306) in the subset of ART-treated immunological responders, while they remained fairly stable in the remaining patients (+28±76 copies/10⁵ cells; p=0.687) (Figure 1A, panels D1 and D2). EBV-DNA tended to increase also in patients treated with *civ/high-dose* IL-2. although this increase was not statistically significant $(+439\pm295 \text{ copies}/10^5 \text{ cells}; p=0.109)$ (Figure 1A, panels A and B). An opposite trend was observed in all patients treated with low-dose IL-2 whose EBV-DNA levels after

Table 1	Baseline	characteristics	of	HIV-1-infected	natients
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	All patients	ART+IL-2 civ/high dose	Treatment ART+IL-2 high dose	ART + IL-2 low dose	ART
No. of individuals	40	9	11	8	12
CD4⁺ T cells/µL mean (range)	348 (189-610)	337 (221-459)	332 (189-610)	377 (275-506)	353 (230-500)
HIV-1 RNA plasma copies/mL mean (range)	23,399 (20- 181,184)	39,064 (53- 181,184)	16,071 (188- 52,348)	31,601 (603- 134,688)	12,901 (20- 50,000)
EBV-DNA copies /10 ⁵ cells mean (range)	311 (2-2,294)	291 (2-1,669)	458 (47-2,294)	242 (2-1,353)	237 (2-1,160)
No. of individuals EBV-DNA+ (plasma) EBV-DNA copies/mL (range)	8 (59-620)	1 (63)	2 (70-93)	2 (96-139)	3 (59-620)

12 months were significantly lower than at baseline (- 202 ± 137 copies/10⁵ cells; p=0.0078) (Figure 1A, panel C). This change in EBV load differed significantly from those observed in patients treated with *civ/high-dose* IL-2 (p=0.0184) and in the subset of ART-treated immuno-logical responders (p=0.0097) (Figure 1B).

Only eight patients had detectable EBV-DNA in plasma at baseline, while 13 had detectable levels after 12 months. A weak correlation was found between cellassociated and plasma EBV-DNA values (Figure 2A). Consistent with the trend observed in PBMC, plasma EBV-DNA load tended to increase in patients treated with *civ/high-dose* IL-2 (p=0.0625). In contrast, all patients treated with *low-dose* IL-2, including two who were positive at baseline, tested negative for EBV-DNA in plasma after 12 months of therapy (Figure 2B). The change in plasma EBV-DNA load observed in the *civ/high-dose* IL-2 arm differed significantly from the changes observed in the *low-dose* IL-2 arm (p=0.0114) and in the subset of ART-treated non-immunological responders (p=0.0124).

None of the patients had a history of symptomatic EBV infection prior to or during the study. One subject who had received high-dose IL-2 developed and died of Castelman's disease 2 years after study completion, and a second subject who had received *civ/high-dose* IL-2 developed non-Hodgkin's lymphomas 4 months after termination of the study. In this study, only 50% of patients treated with ART alone showed a moderate increase in CD4⁺ cell counts and, consistent with previous observations," these patients also showed a concomitant increase in EBV-DNA load. A similar trend was observed in patients treated with *civ/high-dose* IL-2. In contrast, EBV-DNA levels decreased significantly in individuals who received *low-dose* IL-2. This opposite effect may be due to several factors. In ART-treated patients, increases in EBV load have been associated with increased immunoglobulin levels, a surrogate marker for B-cell stimulation." Although the impact of



Figure 1, A. Distribution, mean and standard error (SE) of plasma HIV-1 RNA , CD4⁺ T-cell count and cell-associated EBV-DNA load at baseline and post-therapy in individuals treated with civ/high dose IL-2 (A), high dose IL-2 (B), low dose IL-2 (C), or ART alone (D). Patients treated with ART alone were further divided into subgroups according to increased (D1) or not increased (D2) CD4+ cell count. B. Change (mean and SE) from baseline to post-therapy values of CD4⁺ cell count and cellassociated EBV-DNA load in the different groups of patients.

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IL-2 on EBV expression is unknown, studies have shown that IL-2 induces a dose-dependent proliferation of B cells in vitro.12 Moreover, B cells from HIV-1-infected patients express higher levels of IL-2 receptor than do B cells from normal donors, resulting in an increased IL-2 responsiveness.¹⁰ In addition, among the IL-2-induced cytokines, tumor necrosis factor- α and β promote several B-cell functions, including cell proliferation and immunoglobulin production.^{7,13} Of interest, levels of tumor necrosis factor- α were shown to increase in patients treated with high-dose IL-2, but to decrease in patients treated with low-dose IL-2.14 Thus, dissimilar B-cell stimulation in patients receiving either low or high doses of IL-2 may account for the different EBV dynamics described here. Furthermore, while high concentrations of IL-2 may enhance production of several pro-inflammatory cytokines, via binding to low-affinity receptors expressed on NK cells,^{3,15} low concentrations

Figure 2 (left), A. Relationship between plasma and cell-associated EBV-DNA load at baseline (open circles) and after 12 months (closed circles) of therapy in patients treated with civ/high dose IL- (\bigcirc, \bullet) , high dose IL-2 $(\triangle, \blacktriangle)$, low dose IL-2 (\Box, \blacksquare) , or ART alone (\diamondsuit, \bullet) . B) Baseline and post-therapy values of EBV in plasma in subjects treated with *civ/high* dose IL-2 (\bullet) and *low dose* IL-2 (\bigcirc) .

of IL-2 may promote expansion of cytotoxic T lymphocytes, thus restoring protective immunity against EBV, by binding to high-affinity IL-2 receptors expressed on T lymphocytes. In this regard, previous studies demonstrated that low dose IL-2 prevented the development of EBV-associated lymphoproliferative disease in mice reconstituted with PBMC from EBV-seropositive subjects, a protective effect mainly mediated by CD8⁺ lymphocytes.¹⁶

Although specific studies are required to investigate the impact of IL-2 and tumor necrosis factor- α/β on Bcell stimulation and EBV expression, the present findings suggest that intermittent therapy with *low-dose* IL-2 regimens should be considered in EBV- and HIV-1coinfected patients, particularly in those at risk of developing EBV-induced malignancies.

References

- Kovacs JA, Vogel S, Albert JM, Falloon J, Davey RT, Walker RE, et al. Controlled trial of interleukin-2 infusions in patients infected with the human immunodeficiency virus. N Engl J Med 1996;335:1350-6.
 Tambussi G, Gezzi S, Nozza S, Vallanti
- Tambussi G, Gezzi S, Nozza S, Vallanti G, Magenta L, Guffanti M, et al. Efficacy of low dose intermittent subcutaneous interleukin (IL)-2 in antiviral drug-experienced human immunodeficiency virus-infected person with detectable virus load: a controlled study of three IL-2 regimens with antiviral drug therapy. J Infect Dis 2001; 183:1476-84.
- Jacobson LE, Pilaro F, Smith KA. Rational interleukin 2 therapy for HIV positive individuals: daily low doses enhance immune function without toxicity. Proc Natl Acad Sci USA 1996; 93:10405-10.
- Malnati M, Broccolo F, Nozza S, Samati L, Ghezzi S, Locatelli G, et al. Retrospective analysis of HHV-8 viremia and cellular viral load in HIVseropositive patients receiving interleukin 2 in combination with antiretroviral therapy. Blood 2002:100:1575-8
- viral therapy. Blood 2002;100:1575-8. 5. Ometto L, Menin C, Masiero S, Bonaldi L, Del Mistro A, Cattelan AM, et

al. Molecular profile of Epstein-Barr virus in immunodeficiency virus type 1-related lymphadenopathies and lymphomas. Blood 1997;90:313-22.

- Gaidano G, Carbone A, Dalla Favera R. Pathogenesis of AIDS-related lymphomas: molecular and histogenetic heterogeneity. Am J Pathol 1998; 152: 623-30.
- Kerl JH, Miller A, Fauci AS. Effect of tumor necrosis factor α on mitogenactivated human B cells. J Exp Med 1987;166:786-91.
- Macchia D, Almerigogna F, Parronchi P, Ravina A, Maggi E, Romagnani S. Membrane tumor necrosis factor α is involved in the polyclonal B-cell activation induced by HIV-infected human T cells. Nature 1993;363:464-6.
- Mauray S, Fuzzati-Armentero MT, Trouillet P, Ruegg M, Nicoloso G, Hart M, et al. Epstein-Barr virus-dependent lymphoproliferative disease: critical role of IL-6. Eur J Immunol 2000; 30: 2065-73.
- David D, Bani L, Moreau JL, Treilhou MO, Nakara T, Joussemet M, et al. Regulatory dysfunction of the interleukin-2 receptor during HIV infection and the impact of triple combination therapy. Proc Natl Acad Sci USA 1998; 95:11348-53.
- Righetti E, Ballon G, Ometto L, Menin C, Zanchetta M, Chieco-Bianchi L, et al. Dynamics of Epstein Barr virus in

ADR, GP, AL, GT conceived the study, and contributed to the discussion of results; NB performed the experiments under the direction of ADR; SG and SN collected samples, and virological and immunological data of the patients; PDB performed the statistical analyses. ADR and GP wrote the manuscript with the contribution from other authors. The authors have no conflicts of interest

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HIV-1 infected subjects on highly active antiretroviral therapy. AIDS 2002;16:63-73.

- Nakanishi K, Hirose S, Yoshimoto T, Ishizashi H, Hiroishi K, Tanaka T, et al. Role and regulation of interleukin (IL)-2 receptor α and β chains in IL-2 driven B-cell proliferation. Proc Natl Acad Sci USA 1992;89:3551-5.
- Patke CL, Shearer WT. gp120 and TNFα-induced modulation of human B cell function: proliferation, cyclic AMP generation, Ig production, and B-cell receptor expression. J Allergy Clin Immunol 2000;105:975-82.
- Fortis C, Soldini L, Ghezzi S, Colombo S, Tambussi G, Vicenzi E, et al. Tumor necrosis factor alfa, interleukin 2, and soluble interleukin 2 receptor levels in human immunodeficiency virus type 1-infected individuals receiving intermittent cycles of interleukin 2. AIDS Res Hum Retrov 2002;18:491-9.
- Becknell B, Caligiuri MA. Interleukin-2, interleukin 15, and their role in human natural killer cells. Adv Immunol 2005; 86:209-39.
- 16. Baiocchi RA, Caligiuri MA. Low-dose interleukin 2 prevents the development of Epstein-Barr virus (EBV)-associated lymphoproliferative disease in scid/scid mice reconstituted i.p. with EBV-seropositive human peripheral blood lymphocytes. Proc Natl Acad Sci USA 1994;91:5577-81.