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Abstract	<p>Constructed wetlands represent an increasingly expanding technology for treatment and reuse of poor quality waters and for the development of marginal areas. The exploitation of herbaceous biomass for biogas production may add further appeal to its adoption. Codigestion of lignocellulosic plant materials with pig slurry could meet the need for biomass hydration and possibly improve biogas yields. The objectives of this study were: (1) to evaluate the biomethanation potential of biomass from several species which are of interest for use in constructed wetlands, and its relationship with plant composition; (2) to evaluate the influence of codigestion of selected wetland species with pig slurry on methane production rate and yield. Biogas production was preliminarily measured in laboratory conditions using as substrates biomass samples belonging to 23 plant species coming from different environments. Eight of them were then tested for biogas production, alone or in codigestion with pig slurry (volatile solid ratio: 1/1). In monodigestion, CH₄ yields were on average 213 mL CH₄ g⁻¹ volatile solids. Biogas production was positively related with N content and negatively with acid detergent fiber concentration and C to N ratio. The time for the joining of the maximum methane production was 25 % shorter and the amount of methane was 30 % higher for wetland biomass in codigestion with pig slurry than in monodigestion. The use of pig slurry as hydration medium for anaerobic digestion can improve the biomethanation potential of wetland biomass.</p>	
Keywords (separated by '-')	Biomethanation - Constructed wetlands - Lignocellulosic biomass - Pig slurry	
Footnote Information		

2 **Biomethanation Potential of Wetland Biomass in Codigestion**
3 **with Pig Slurry**

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8 expanding technology for treatment and reuse of poor
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Keywords Biomethanation · Constructed wetlands · 36
Lignocellulosic biomass · Pig slurry 37

Introduction 38

A wetland is a land area that is saturated with water, either 39
permanently or seasonally, so that it takes on the charac- 40
teristics of a distinct ecosystem. A wetland differs from 41
other land environments or water bodies because its veg- 42
etation is adapted to unique soil conditions. Constructed 43
wetlands (CWs) are a technology developed in recent years 44
for treatment and reuse of poor quality waters and for the 45
development of marginal areas. They are systems of 46
purification of municipal, agricultural and industrial 47
wastewater, which reproduce the principle of self-purifi- 48
cation typical of aquatic environments and wetlands. Plant 49
species more frequently utilized are water macrophytes. 50
The most commonly exploited species in Europe are 51
Phragmites australis, and species belonging to the genera 52
Carex, *Scirpus*, *Typha* [36], emergent macrophytes well 53
tolerating high nutrient and pollution levels. 54

The exploitation of herbaceous biomass from wetlands 55
for energy production (heat, electricity and fuels) may add 56
further appeal to the adoption of this practice [15, 19, 22]. 57
In fact, wetland plant species are well adapted to growing 58
in wastewater and are often vigorous, high-productive 59
plants. In recent years wetland biomass utilization for 60
biogas production has received growing attention [1, 2, 25, 61
35]. Earlier studies on conversion of plant biomass into 62
methane [30] revealed particular suitability of water hya- 63
cinth (*Eichornia crassipes* Mart) and napier grass 64

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65 (*Pennisetum purpureum* L.) for biogas production. Dipu
66 et al. [12] evaluated 6 macrophyte species belonging to
67 genera *Typha*, *Pistia*, *Eichornia*, *Salvinia*, *Azolla*, and
68 *Lemna*, using cow dung as inoculum, and found higher
69 biogas production in codigestion slurries than in cow dung
70 digested alone. Cohen et al. [9] have proposed an inte-
71 grated treatment system, including CWs for water polish-
72 ing and anaerobic digestion (AD) of wetland-derived
73 phytomass, for enhancing the economic feasibility of
74 wastewater treatment processes.

75 Limited amounts of lignocellulosic biomass are com-
76 monly used in co-digestion with manure for biogas pro-
77 duction in order to enrich manure with volatile solids
78 without excessively enlarging the digester size. However,
79 the frequency of AD using vegetal biomass without manure
80 has recently increased, due to the incentive policies for
81 renewable energies. Government incentives have also
82 raised the interest of the agroindustry (such as olive oil
83 mills, cheese factories, breweries) toward the exploitation
84 of agro-industrial waste for biogas production with no
85 connection with livestock.

86 Fresh lignocellulosic biomass has usually a high (i.e.,
87 >35 %) dry matter content, especially when ensilage or
88 drying is applied to prolong its storage life. Dry fer-
89 mentation is the most suitable system for biogas pro-
90 duction from materials with low moisture content.
91 However, the most spread AD systems nowadays are of
92 the Continuously Stirred Tank Reactor (CSTR) type.
93 Biogas production in CSTR systems requires a dry matter
94 content lower than 10 % [38]. Consequently, lignocellu-
95 losic biomass, when used for biogas production in CSTR
96 systems, needs to be diluted. The use of water for bio-
97 mass dilution is arguable, because water occupies volume
98 without producing biogas. Liquid animal manure
99 (“slurry”) seems the most suitable dilution material
100 because, on the one hand, it hydrates the biomass while
101 supplying it with nutrients; on the other hand, the use of
102 animal manure contributes to the solution of the wide-
103 spread problem of a proper manure management. The
104 hypothesis at the basis of this research is that the use of
105 animal manure in codigestion with wetland biomass may
106 contribute to biogas yield improvement while fulfilling to
107 the general environmental need of a proper manure
108 management.

109 The aims of this study were: (1) to evaluate the
110 biomethanation potential (BMP) of wetland biomass,
111 coming either from natural environments or from CWs.
112 Our interest focused on the overall effect of wetland bio-
113 mass as substrate for AD, regardless of the species; (2) to
114 verify the effect of codigestion of wetland biomass with pig
115 slurry on methane production rate and yield.

Materials and Methods 116

Materials 117

118 Samples of wetland biomass were collected in autumn, at
119 the end of the growing season, in their natural environment
120 (Italy, Po Valley, Veneto region, 45°38'N, 11°40'E, 10 m
121 a.s.l.) or in CWs experimental plants, located in the same
122 area and managed by the DAFNAE Department of the
123 Padua University. Samples belonging to 23 plant species
124 were obtained (Table 1). The environment of these species
125 is characterized by high levels of soil moisture. For this
126 reason they have been assessed in experimental tests for
127 their potential use in constructed wetlands, for removing
128 high levels of N and organic load from animal slurry or
129 digestate [26]. Some of them are typical macrophytes,
130 others live in riparian environments or uncultivated lands,
131 some others grow in humid areas close to the sea, in saline
132 environments. Representative subsamples were dried at
133 65 °C at constant weight, and then milled at 1 mm (Cutting
134 Mill SM 100 Comfort, Retsch, Germany). Each sample
135 was a composite of aboveground biomass from 5 plant
136 individuals collected in the same site. As each plant species
137 was represented by only one sample collected at a single
138 site, no statistical inference was drawn on the species effect
139 on AD, which is beyond the scope of this work.

140 Fresh pig slurry to be used in co-digestion with wetland
141 biomasses was drawn after biomass mixing with a pumping
142 system from the CREA farm storage tank collecting faeces,
143 urine, tap water used for cleaning pens from a fattening
144 piggery, and rainwater. Its average composition was: total
145 solids (TS), 1.39 % fresh weight (FW; SD, 0.045 %);
146 volatile solids (VS), 0.98 % FW (SD, 0.040 %); ashes,
147 0.41 % FW (SD, 0.0006 %); organic C, 396 g kg⁻¹ TS
148 (SD, 2.40 g kg⁻¹); total N, 56.1 g kg⁻¹ TS (SD,
149 1.83 g kg⁻¹); ammonium N, 26.3 g kg⁻¹ TS (SD,
150 0.75 g kg⁻¹); pH in water, 7.14 (SD, 0.08); total P,
151 22.4 g kg⁻¹ TS (SD, 0.72 g kg⁻¹); lignin, 5.7 % TS (SD,
152 0.09 %); hemicellulose, 10.6 % TS (SD, 0.05 %); cellu-
153 lose, 6.1 % TS (SD, 0.08 %). These composition values are
154 consistent with historical data from our laboratory regard-
155 ing pig slurry produced in our experimental farm.

Experimental Set-Up 156

157 A preliminary experiment was carried out to test the
158 average biomethanation potential (BMP) that can be
159 expected when using wetland biomass as AD substrate,
160 using 23 wetland biomass samples. This experiment was
161 also used to examine the relationship between plant com-
162 position and AD performances.

Table 1 Wetland plant species used as a source of biomass for AD

Species	Common name	Natural environment of growth
<i>Arctium lappa</i> L.	Greater burdock	DA, NRS
<i>Artemisia caerulescens</i> L.	Sea mugwort	Salty soils
<i>Arundo donax</i> L.	Giant reed	Riparian
<i>Aster tripolium</i> L.	Sea aster	Moist salty soils
<i>Calamagrostis epigejos</i> (L.) Roth	Wood small reed	Moist salty soils
<i>Carex acutiformis</i> Ehrh.	Lesser pond sedge	Wetland
<i>Carex riparia</i> L.	Great pond sedge	Wetland
<i>Cynodon dactylon</i> (L.) Pers.	Bermudagrass	DA, NRS, moist sites along rivers
<i>Elytrigia atherica</i> (Link) Kerguélen	Wheatgrass	Moist salty soils
<i>Glyceria maxima</i> (Hartm.) Holmb	Reed mannagrass	Wetland
<i>Halimione portulacoides</i> (L.) Aellen	Sea purslane	Moist salty soils
<i>Helianthus tuberosus</i> L.	Jerusalem artichoke	Riparian
<i>Inula crithmoides</i> L.	Golden samphire	Moist salty soils
<i>Iris pseudacorus</i> L.	Yellow flag	Wetland
<i>Juncus maritimus</i> Lam.	Sea rush	Wetland
<i>Limonium narbonense</i> Mill.	Sea lavender	Moist salty soils
<i>Miscanthus x giganteus</i> Greef et Deu.	Giant miscanthus	Moist meadows
<i>Phalaris arundinacea</i> L.	Reed canarygrass	Wetland
<i>Pucciniella palustris</i> (Seen.) Hayek	Alkaligrass	Moist salty soils
<i>Sarcocornia fruticosa</i> (L.) A.J.Scott	Glasswort	Moist salty soils
<i>Scirpus sylvaticus</i> L.	Woodland bulrush	Wetland
<i>Symphytus x uplandicum</i> Nyman	Comfrey	DA, UL, NRS
<i>Typha latifolia</i> L.	Broadleaf cattail	Wetland

DA disturbed areas, UL uncultivated land, NRS nitrogen rich soils

163 In the second experiment, the rate and yield of methane
164 production were compared for wetland biomass samples
165 (*Plant material*), in monodigestion (PS−) or in codigestion
166 (PS+) with pig slurry (*Treatment*), in a completely ran-
167 domized block design with 3 replications. The plant
168 materials which had given the best or the worst results in
169 the first experiment were selected for this comparison:
170 *Arundo donax*, *Carex riparia*, *Cynodon dactylon*, *Elytrigia*
171 *atherica*, *Halimione portulacoides*, *Inula crithmoides*,
172 *Scirpus sylvaticus*, *Phragmites australis*, which is one of
173 the dominant wetland species in Europe [36], was also
174 included.

175 Anaerobic Digestion and BMP Determination

176 Digestate from pig slurry was used as inoculum source. It
177 was prepared as follows: 200 mL of a definite synthetic
178 medium for methanogens (phosphate buffered basal med-
179 ium, PBBM; [14]) without energy sources was mixed in
180 500-mL serum bottles with 200 mL fresh liquid fraction of
181 pig slurry collected from the farm storage tank after sep-
182 aration of the solid phase, in a N₂-CO₂ (80:20) atmosphere.
183 This mixture was left to incubate in strictly anaerobic
184 conditions and the head space composition was analyzed

185 for CH₄ accumulation. The inoculum was considered as
186 ready for use when CH₄ production had stopped, indicating
187 exhaustion of endogenous energy sources.

188 Anaerobic digestion was carried out using dried and
189 milled wetland biomass samples as substrates. In the first
190 experiment, the reaction mixture included 1.25-g dried
191 sample (2.5 %; “substrate”), 50 mL of PBBM (“hydration
192 medium”) without energy sources, and 5 mL inoculum, in
193 100-mL reactors (118.5 mL effective volume), in triplicate
194 (69 reactors, in total). The pH of the reaction mixtures
195 varied between 6.0 and 7.7. In the second experiment, each
196 reactor contained 1 g VS. Precisely, in each PS− reactor,
197 1 g VS of plant biomass was added to 50 mL PBBM; in
198 each PS+ reactor, 0.51 g VS of plant biomass were added
199 to 50 mL pig slurry, containing 0.49 g VS, for a total of 1 g
200 VS. The total VS concentration in all the reactors was 2 %.
201 Pig slurry alone was inoculated as control. Fifty-one
202 reactors were prepared in total (8 plant materials × 2
203 substrate levels + pig slurry alone, ×3 replicates). Five-
204 mL inoculum was added to all 100-mL reactors (118.5 mL
205 effective volume). The average pH of wetland biomass
206 after mixing with PBBM was 6.72 (SD, 0.52), while in the
207 presence of pig slurry it was 6.80 (SD, 0.50). The head-
208 space of the reactors was gassed with N₂-CO₂ (80:20)

throughout the preparation steps before the start of the experiment. Reactors were plugged with butyl rubber stoppers and aluminum seals and incubated at 35 °C for 90 days.

The biogas production (volume and composition) was measured according to Owen et al. [24] 2 days after the start of the incubation and then weekly for 3 months. Biogas was collected by means of 100-mL glass syringes. The incubation period was completed when there was no more biogas production in any of the reactors. No methane production was detected in the control reactors (inoculum in PBBM without energy source).

Biomethanation potential (mL CH₄ g⁻¹ VS) was expressed as the maximum amount of CH₄ cumulated over time that can be produced by a given substrate per g of volatile solids, including the amounts of CH₄ released in the syringe at each measurement date as well as the CH₄ volume remaining within the reactor.

In the second experiment, the parameters of the cumulative CH₄ production curves were evaluated by means of a modified 3-parameter Gompertz equation [17]:

$$M(t) = M_{max} \exp \left\{ -\exp \left[\left(\frac{e R_{max}}{M_{max}} \right) (\lambda - t) + 1 \right] \right\}$$

where M(t) (mL) is the total amount of CH₄ produced at the culture time t (d); e is exp(1); M_{max} (mL) is the maximum cumulative CH₄ production; R (mL d⁻¹) is the daily rate of CH₄ accumulation in the linear phase of CH₄ accumulation; and λ is the lag time duration (d), that is the time of microbial adaptation before exponential CH₄ production. This function is often utilized for interpolating growth curves, in general, and microbial growth curves, in particular [42].

Since in this experiment each reactor contained 1 g of VS, the M_{max} value (mL CH₄) coincided with the BMP value (mL CH₄ g⁻¹ VS).

Analytical Methods

The following parameters describing plant composition were determined: pH of the reaction mixture, TS, VS, total N, total P, organic C, C to N ratio (C/N), neutral detergent fiber (NDF), acid detergent fiber (ADF), lignin (ADL), hemicellulose, cellulose, total polyphenols (TP), soluble carbohydrates (SC), starch (Sta), total carbohydrates (TC = SC + Sta).

Total solids were determined gravimetrically by thermal treatment at 105 °C at constant weight. Analyses of the plant materials were conducted on samples dried at 65 °C at constant weight and milled at 1 mm. Organic C was determined by dichromate oxidation with external heating and reflux condenser. Total N was determined with the Kjeldahl apparatus. Total P was determined on ashes by colorimetry with ammonium molybdate, after solubilization by means of HCl

1 N. The pH was determined after suspension, 2-h stirring and sedimentation of 1.5 g dry matter in 50 mL distilled water. Fiber fractions (NDF, ADF, ADL) were determined according to Van Soest et al. [33]. The hemicellulose content was estimated as the difference between NDF and ADF; the cellulose content as the difference between ADF and ADL. For SC and Sta determination, plant tissues (20 mg) were washed with pure acetone to remove the interfering pigments and then centrifuged [21]. Soluble carbohydrates were extracted twice with 2.5 mL ethanol 80 % and determined on the centrifuged supernatant by the anthrone method [23]. Five mL HCl 1.1 % were added to the centrifuged residual pellet, and diluted to 10 mL with distilled water after heating in a water bath at 100 °C for 10 min. Soluble carbohydrates after hydrolysis were determined with the anthrone method. Soluble carbohydrates, starch and TC are expressed as mg glucose g⁻¹ of dry matter. Total polyphenols were determined according to the Folin–Ciocalteu colorimetric assay [31] and expressed as mg tannic acid per g of dry matter.

Pig slurry and digestates were analyzed according to APHA [3]. In the first experiment, digestate analysis was performed on a composite sample obtained by mixing the digestate of the 3 treatment replicates. In the second experiment, the single replicates were used for analysis.

Methane concentration in the biogas was determined by means of a MicroGC Agilent 3000 gas chromatograph, equipped with 2 columns, Molsieve and Plot U; detector: TCD. Carrier gas: argon.

Statistical Analysis

The correlation matrix between AD and BMP was obtained by means of the PROC CORR of the SAS package [28]. ANOVA was applied to compare the effect of wetland digestion with or without pig slurry in the second experiment. Comparisons of the means were based on the Tukey test at α = 0.01.

Model fitting for the description of CH₄ accumulation curves was performed using the PROC NLIN of the SAS package. The parameter values were estimated according to the Gauss–Newton method. The time (d) necessary to reach M_{max} was estimated by calculating the ratio M_{max}/R. Data from 3 replicates was merged for the parameter value estimation.

Results and Discussion

Biomethanation Potential of Wetland Biomasses

The BMP of the plant materials was on average 213 mL CH₄ g⁻¹ VS (n = 23, CV = 18.6 %). Nearly 75 % of the plant materials (17 out of 23 plant species; Fig. 1) showed

305 a BMP > 200 mL CH₄ g⁻¹ VS, with 5 among them pro-
 306 ducing more than 250 mL CH₄ g⁻¹ VS. These amounts are
 307 lower than those reported for energy crops and other
 308 agricultural by products, which may produce even more
 309 than 400 mL CH₄ g⁻¹ VS [5, 6]. However, they were of the
 310 same level or even higher than those that can be obtained
 311 from agro-industrial waste [11] or wheat straw [34].

312 Residual VS in wetland biomass digestates were on
 313 average 51.7 % of the initial VS content (Table 2). The
 314 mean VS decrease was then 48.3 %, lower than that
 315 reported by Bouallagui et al. [6] for AD of fruit and veg-
 316 etable waste (58–65 %). Klimiuk et al. [16], for silages of
 317 four crop species, found large differences in residual VS
 318 content at the end of AD, depending on the species. The
 319 decrease in organic C content and C to N ratio caused by
 320 CH₄ and CO₂ release during AD was accompanied by an
 321 increase (nearly doubling) of N and P concentrations in the
 322 digestate, compared to those measured at the start of the
 323 process (Tab. 3) in agreement with the results of Tambone
 324 et al. [32].

325 Relationship Between Anaerobic Digestion 326 and Plant Composition

327 Among plant composition parameters, most varying
 328 (CV > 60 %) among species were: C to N ratio, soluble
 329 carbohydrates and starch (and, consequently, total carbo-
 330 hydrates; Table 3). Possible reasons for differences among
 331 plant materials in suitability to AD were evaluated by
 332 means of correlations between plant composition param-
 333 eters and BMP. Biomethanation potential was positively
 334 correlated with plant N content ($r = 0.59$, $P < 0.01$) and
 335 negatively correlated with C to N ratio ($R = -0.63$,
 336 $P < 0.01$), ADF (i.e., lignin + cellulose) content ($R =$
 337 -0.71 , $P < 0.001$), and cellulose content ($R = -0.53$,
 338 $P < 0.01$).

Table 2 Simple statistics of selected composition parameters, for digestate coming from AD of wetland biomass (n = 23)

Parameter	Mean	Minimum	Maximum	CV (%)
TS (%)	1.36	1.10	1.78	14.8
VS (% initial VS)	51.7	36.0	73.3	19.2
N (g kg ⁻¹ TS)	47.4	4.8	96.6	59.4
P (g kg ⁻¹ TS)	5.4	0.9	11.3	59.1
Organic C (g kg ⁻¹ TS)	310	223	367	10
C/N	13.1	3.2	75.8	128

The N content and the C to N ratio are important factors 339
 for the improvement of biogas production, even though 340
 contrasting effects on BMP were reported, probably 341
 depending on the range of explored values [29, 37, 41]. 342
 These results suggest the opportunity to increase biogas 343
 yields from wetland biomass by appropriate modulation of 344
 the C to N ratio. 345

It is well known that lignin, among VS components, is 346
 especially recalcitrant to AD [8]. Alvinge [2] tested for 347
 biogas production two macrophyte species, *Typha latifolia* 348
 and *Phalaris arundinacea*, with or without treatment of 349
 demolition of the lignocellulosic tissues (mechanical mil- 350
 ling, alkaline treatment with lime and fungal degradation), 351
 and he was able to obtain increased CH₄ production by 352
 16–27 %, depending on the kind of applied pretreatment. 353

Polyphenols, common tissue components of several 354
 plant species, had been included in the analysis because 355
 they could exert an inhibiting effect on microbial activities 356
 [10]. In this experiment the total polyphenols concentration 357
 was not significantly correlated to BMP. The presence of a 358
 VS fraction containing lignocellulosic molecules recalci- 359
 trant to digestion may explain the only partial removal of 360
 VS during the AD process. 361

Fig. 1 Biomethanation potential (BMP) of biomass samples from selected wetland species. Error bars are standard deviations

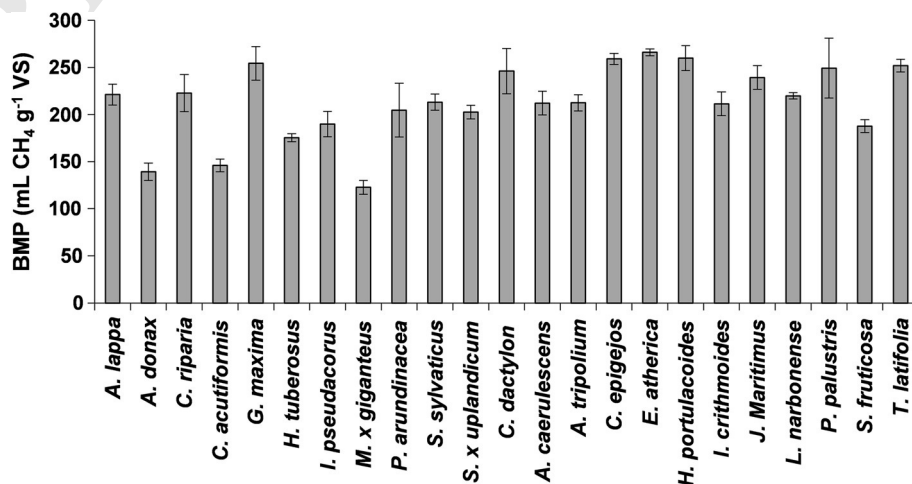


Table 3 Average composition of wetland biomass samples, and minimum, maximum and coefficient of variability (CV) (n = 23)

Parameter	Mean	Minimum value	Maximum value	CV (%)
pH of the reaction mixture	6.7	6.0	7.7	7.7
Total solids, %	94.7	91.5	96.1	1.1
Volatile solids, %	89.0	78.8	96.5	5.5
Ashes, %	11.0	3.5	21.2	44.6
Total N, g kg ⁻¹	23.4	5.0	40	50.7
Total P, g kg ⁻¹	2.9	0.7	6.3	54.9
Organic C, g kg ⁻¹	429	368	465	5.6
C/N	26	11	85	73.4
Hemicellulose, % ^a	19.2	5.9	38.7	47.3
Cellulose, %	23.6	4.1	45.1	54.0
Lignin, %	13.4	5.3	33.4	54.1
Total polyphenols (mg tannic acid g ⁻¹)	15.4	6.1	48.1	56.4
Soluble carbohydrates (SC, mg glucose g ⁻¹)	61	25	279	85.5
Starch (Sta, mg glucose g ⁻¹)	63	26	207	67.4
Total carbohydrates (TC = SC + Sta, mg glucose g ⁻¹)	123	57	354	62.9

All the concentration values are referred to the total solids content

362 Biogas Production by Wetland Biomasses 363 in Codigestion with Pig Slurry

364 The codigestion with pig slurry reduced the AD lag phase
365 (Fig. 2a) while increasing the R (Fig. 2b) and Mmax values
366 (Fig. 2c). The lag phase duration varied from 0, in the
367 majority of cases, to 0.6 days in the PS- reactors (higher
368 than 0 in 4 out 23 cases). In the PS+ reactors it varied
369 between 0 and 0.31 days (higher than 0 in only 2 cases).
370 Lag phase duration depends on several factors including
371 the level of recalcitrance of the substrate. As in this
372 experiment the lag phase duration was 0 or very short, no
373 negative effects of the substrate on microbial activities
374 could be deduced.

375 The R values averaged 9.2 mL CH₄ day⁻¹, in the PS-,
376 and 16.0 mL CH₄ day⁻¹, in the PS+ reactors (Tukey value
377 for the difference between the PS- and the PS+ treat-
378 ments, at $P < 0.01$: 1.53 mL CH₄ day⁻¹), with a 25 %
379 reduction on average of the time needed to reach Mmax
380 (from 27.7 to 20.7 days) in PS+. The average Mmax was
381 255 mL CH₄ g⁻¹ VS, in the PS- reactors, and 332 mL g⁻¹
382 VS, in the PS+ reactors (Tukey value for the difference
383 between the PS- and the PS+ treatments, at $P < 0.01$:
384 16.7 mL of CH₄), with a 30 % increase in methane pro-
385 duction, for the same amount of initial VS content. In this
386 experiment the Mmax values (i.e., cumulated mL CH₄)
387 coincided with BMP values (cumulated mL CH₄ g⁻¹ VS),
388 because the substrate of all the reactors contained 1 g of
389 VS.

390 Besides the general improvement of methane produc-
391 tion rate and yield, codigestion reduced the differences in
392 AD performances between plant materials. The CV of R

and Mmax values in the PS+ treatment (9.7 and 5.0 %, 393
394 respectively) was lower than in the PS- treatments (25.3
395 and 10.0 %, respectively). The increase of Mmax for the
396 plant materials in codigestion with pig slurry, in com-
397 parison with monodigestion, was particularly high for
398 those which had given the worst results in monodigestion
399 such as *S. silvaticus*, *I. crithmoides* and *P. australis*.
400 Codigestion has been reported to be advantageous
401 because it results in a substrate better balanced and
402 assorted in terms of nutrients [13, 18]. Positive effects of
403 codigestion with pig slurry could be attributed in par-
404 ticular to an enrichment in mineral salts and to an
405 increase in N availability for microorganisms. In fact, the
406 initial ash concentration was 67 % higher in codigestion
407 than in monodigestion (Table 4), for the same initial
408 amount of volatile solids, whereas the total N content
409 was 48 % higher and the concentration of ammonium N
410 was nearly 6 times higher than that in monodigestion. As
411 pig slurry is particularly rich in methanogenic micror-
412 ganisms [27] a possible contribution of the pig slurry
413 microbial populations to the methanogenic activity could
414 also be hypothesised. However, it is also known that the
415 type and relative richness of the various microbial groups
416 in the anaerobic digesters is driven by the substrate
417 characteristics [40]. Therefore, the quantitative and
418 qualitative relationship between the initial and the con-
419 solidated microbial populations in batch reactors is not
420 obvious.

421 The average residual VS content of the digestates was
422 47.7 % of the initial VS content in the PS- treatment and
423 44.8 % in the PS+ treatment, without significant differ-
424 ences between PS- and PS+ treatments and no significant

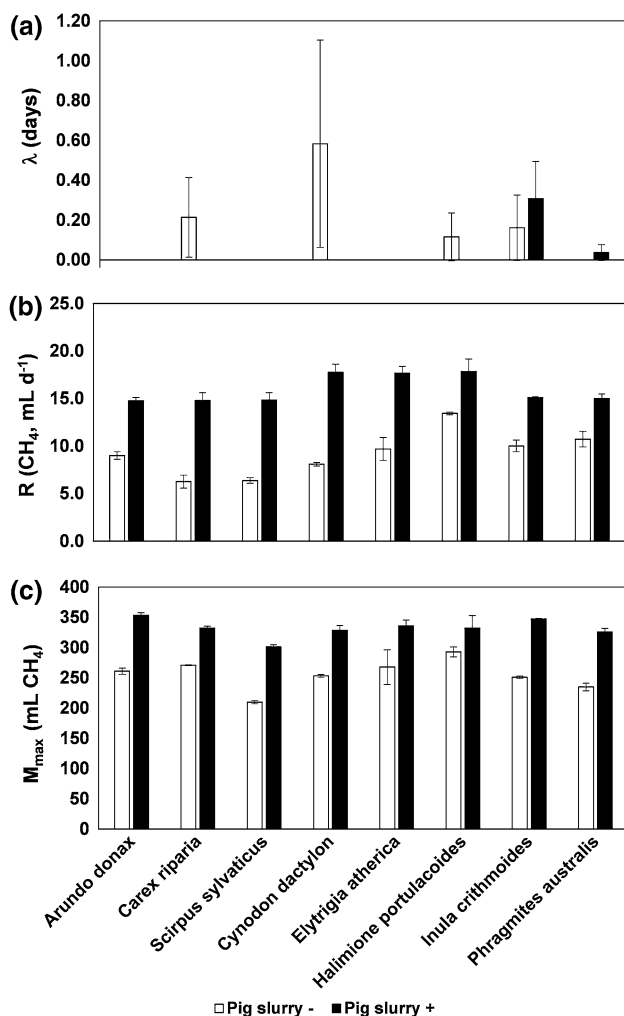


Fig. 2 Values of the Gompertz parameters for the curves of methane accumulation from wetland biomasses digested with (Pig slurry +) or without pig slurry (Pig slurry -). **a** lag phase duration (λ); **b** maximum CH_4 accumulation rate (R); **c** maximum potential methane production (M_{max}). Error bars are standard deviations

Table 4 Changes in substrate composition during anaerobic digestion of wetland plant samples digested with or without pig slurry

Parameter	Digestion without pig slurry		Digestion with pig slurry	
	Mean	CV	Mean	CV
Total solids (TS), %				
Input material	2.54	2.77	2.77	1.30
Digestate	1.8	10.4	1.6	8.7
Volatile solids (VS), % initial ^a				
Digestate	47.7	14.8	44.8	10.8
Ashes, % TS				
Input material	0.45	15.7	0.68	5.3
Digestate	0.75	9.3	0.64	7.3
Total N, % TS				
Input material	3.0	35.0	4.4	10.9

correlation between residual VS content of the digestates and BMP. 425 426

The better biomethanation performances observed when 427
plant materials were in codigestion with pig slurry, for the 428
same starting amount of VS, can be related to differences 429
in the quality of these VS. The most productive plant 430
materials were those having higher N concentrations in 431
their tissues (*E. atherica* and *H. portulacoides*, Table 4). 432
Pig slurry further increased N availability while lowering 433
the C to N ratio. Codigestion with lignocellulosic plant 434
material has been suggested for animal effluents, in order 435
to increase the carbon amount available for AD [4] and to 436
adjust the C to N ratio at levels suitable for AD [39]. 437
According to our results, the opposite is also true: as lig- 438
nocellulosic materials supply high amounts of carbon we 439
can improve AD performances by adding animal manure 440
rich in nitrogen that compensates for these high amounts of 441
C, lowering and improving the C to N ratio. The amount of 442
ammonia-N supplied in the reactors by pig slurry (0.033 % 443
fresh weight; 1.18 % TS, on average, as the difference 444
between PS+ and PS- mean values; Table 4) was not so 445
high as to inhibit the methanogenic activity [7]. It as been 446
reported that ammonia supply to lignocellulosic manure 447
can even enhance biogas yield [20]. 448

Conclusions 449

Interesting BMP levels were associated to anaerobic 450
digestion of wetland biomass. The variability of BMP 451
among wetland samples was linked to their nutrient con- 452
tent. An important role was played by the C to N ratio. The 453
time for the joining of the maximum methane production 454
was on average 25 % shorter and the amount of methane 455
was 30 % higher for wetland biomass in codigestion with 456
pig slurry than in monodigestion. The advantage of 457

Table 4 continued

Parameter	Digestion without pig slurry		Digestion with pig slurry	
	Mean	CV	Mean	CV
Digestate	5.1	33.4	9.7	16.2
NH ₄ -N, % TS				
Input material	0.24	2.72	1.42	1.29
Digestate	2.7	55.5	6.6	20.2
Organic C, % TS				
Input material	40.8	2.4	40.1	1.2
Digestate	33.1	10.0	31.9	3.8
C/N				
Input material	15.7	40.9	9.2	11.6
Digestate	7.2	31.3	3.4	14.7

^a Initial volatile solids content in the reactors (substrate + inoculum): 2.09 g L⁻¹

458 codigestion with pig slurry was particularly evident
459 (35–43 % more CH₄ production) when using plant materi-
460 als that had not given the best results in monodigestion,
461 such as *A. donax* and *P. australis*. Pig slurry in codigestion
462 with wetland biomass modified the N content and the C to
463 N ratio of the methanogenic substrate, with an overall
464 improvement of the methane production rate and yield.
465 Liquid animal manure is therefore a better hydration
466 medium for AD of wetland biomass, in comparison with
467 water, because its supply permits to adjust the C to N ratio
468 in favour of higher methane production rates and yields.

469 The joint evaluation of attitude to biomethanation and
470 agronomic performance will allow the selection of the
471 wetland species most advantageous as substrates for
472 biomethanation. These materials could represent a more
473 valid and environmentally sustainable alternative for AD
474 than energy crops.

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