

Selenoprotein Gene Nomenclature

Vadim N. Gladyshev^{1,2,#}, Elias S. Arnér³, Marla J. Berry⁴, Regina Brigelius-Flohé⁵, Elspeth A. Bruford⁶, Raymond F. Burk⁷, Bradley A. Carlson⁸, Sergi Castellano⁹, Laurent Chavatte¹⁰, Marcus Conrad¹¹, Paul R. Copeland¹², Alan M. Diamond¹³, Donna M. Driscoll¹⁴, Ana Ferreira^{15,16,17,18}, Leopold Flohé^{19,20}, Fiona R. Green²¹, Roderic Guigo^{22,23}, Diane E. Handy²⁴, Dolph L. Hatfield⁸, John Hesketh^{25,26,27}, Peter R. Hoffmann⁴, Arne Holmgren³, Robert J. Hondal²⁸, Michael T. Howard²⁹, Kaixun Huang³⁰, Hwa-Young Kim³¹, Ick Young Kim³², Josef Köhrlé³³, Alain Krol³⁴, Gregory V. Kryukov³⁵, Byeong Jae Lee³⁶, Byung Cheon Lee³², Xin Gen Lei³⁷, Qiong Liu³⁸, Alain Lescure^{34,39}, Alexei V. Lobanov¹, Joseph Loscalzo⁴⁰, Matilde Maiorino²⁰, Marco Mariotti¹, K. Sandeep Prabhu⁴¹, Margaret P. Rayman⁴², Sharon Rozovsky⁴³, Gustavo Salinas⁴⁴, Lutz Schomburg³³, Ulrich Schweizer⁴⁵, Miljan Simonović⁴⁶, Roger A. Sunde⁴⁷, Petra A. Tsuji⁴⁸, Susan Tweedie^{6,49}, Fulvio Ursini²⁰, Yan Zhang³⁸

¹ Division of Genetics, Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, 02115, USA

² Broad Institute of Harvard and MIT, Cambridge, MA 02142, USA

³ Division of Biochemistry, Department of Medical Biochemistry and Biophysics (MBB), Karolinska Institutet, SE-171 77, Stockholm, SWEDEN

⁴ Department of Cell and Molecular Biology, John A. Burns School of Medicine, University of Hawaii at Manoa, Honolulu, HI, 96813, USA

⁵ German Institute of Human Nutrition Potsdam-Rehbruecke, 14558, Nuthetal, GERMANY

⁶ HUGO Gene Nomenclature Committee (HGNC), European Bioinformatics Institute-European Molecular Biology Laboratory (EMBL-EBI), Hinxton, CB10 1SD, UK

⁷ Division of Gastroenterology, Hepatology, and Nutrition, Department of Medicine, Vanderbilt University School of Medicine, Nashville, TN, 37232, USA

⁸ Molecular Biology of Selenium Section, Mouse Cancer Genetics Program, Center for Cancer Research, National Institutes of Health, Bethesda, MD, 20892, USA

⁹ Department of Evolutionary Genetics, Max Planck Institute for Evolutionary Anthropology, 04103, Leipzig, GERMANY

¹⁰ Centre International de Recherche en Infectiologie, CIRI, INSERM U1111, and CNRS/ENS UMR5308, 69007, Lyon, FRANCE

¹¹ Helmholtz Zentrum München, Institute of Developmental Genetics, 85764, Neuherberg, GERMANY

¹² Department of Biochemistry and Molecular Biology, Rutgers-Robert Wood Johnson Medical School, Piscataway, NJ, 08854, USA

¹³ Department of Pathology, University of Illinois at Chicago, Chicago, IL, 60607, USA

¹⁴ Department of Cellular and Molecular Medicine, Lerner Research Institute, Cleveland Clinic Foundation, Cleveland, Ohio, 44195, USA

¹⁵ Pathophysiology of striated muscles laboratory, Unit of Functional and Adaptive Biology (BFA), University Paris Diderot, Sorbonne Paris Cité, BFA, UMR CNRS 8251, 75250, Paris, FRANCE

¹⁶ Inserm U787, Myology group, Institut de Myologie, Groupe Hospitalier Pitié-Salpêtrière, 75013, Paris, FRANCE

¹⁷ UPMC, UMR787, 75013, Paris, FRANCE

¹⁸ AP-HP, Centre de Référence Maladies Neuromusculaires Paris-Est, Groupe Hospitalier Pitié-Salpêtrière, 75013, Paris, FRANCE

¹⁹ Universidad de la República, Facultad de Medicina, Departamento de Bioquímica, 11800, Montevideo, URUGUAY

²⁰ Department of Molecular Medicine, University of Padova, I-35121, Padova, ITALY

²¹ Department of Biochemistry and Physiology, Faculty of Health and Medical Sciences, University of Surrey, Guildford, GU2 7XH, UK

- ²² Centre for Genomic Regulation (CRG), 08003, Barcelona, SPAIN
- ²³ Universitat Pompeu Fabra (UPF), 08002, Barcelona, SPAIN
- ²⁴ Cardiovascular Division, Department of Medicine, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, 02115, USA
- ²⁵ Institute for Cell and Molecular Biosciences, Newcastle University, NE1 7RU, Newcastle-upon-Tyne, UK
- ²⁶ Human Nutrition Research Centre, Newcastle University, NE1 7RU, Newcastle-upon-Tyne, UK
- ²⁷ The Medical School, Newcastle University, Newcastle-upon-Tyne, NE2 4HH, UK
- ²⁸ Department of Biochemistry, University of Vermont, Burlington, VT, 05405, USA
- ²⁹ Department of Human Genetics, University of Utah, Salt Lake City, Utah, 84112, USA
- ³⁰ Hubei Key Laboratory of Bioinorganic Chemistry & Materia Medica, School of Chemistry and Chemical Engineering, Huazhong University of Science and Technology, Wuhan, 430074, P. R. CHINA
- ³¹ Department of Biochemistry and Molecular Biology, Yeungnam University College of Medicine, Daegu, 42415, SOUTH KOREA
- ³² College of Life Sciences and Biotechnology, Korea University, Seoul, 02841, SOUTH KOREA
- ³³ Institute for Experimental Endocrinology, Charité -Universitaetsmedizin Berlin, D-13353, Berlin, GERMANY
- ³⁴ Architecture et Réactivité de l'ARN, Université de Strasbourg, Centre National de la Recherche Scientifique, Institut de Biologie Moléculaire et Cellulaire, 67084, Strasbourg, FRANCE
- ³⁵ KSQ Therapeutics, Cambridge, MA, 02139, USA
- ³⁶ School of Biological Sciences, Seoul National University, Seoul, 151-742, SOUTH KOREA
- ³⁷ Department of Animal Science, Cornell University, Ithaca, NY, 14853, USA
- ³⁸ Shenzhen Key Laboratory of Marine Biotechnology and Ecology, College of Life Science, Shenzhen University, Shenzhen, 518060, Guangdong Province, P. R. CHINA
- ³⁹ Centre National de la Recherche Scientifique, 75794, Paris, FRANCE
- ⁴⁰ Department of Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, MA, 02115, USA
- ⁴¹ Department of Veterinary and Biomedical Sciences, The Pennsylvania State University, University Park, PA, 16802, USA
- ⁴² Department of Nutritional Sciences, Faculty of Health and Medical Sciences, University of Surrey, Guildford, GU2 7XH, UK
- ⁴³ Department of Chemistry and Biochemistry, University of Delaware, Newark, DE, 19716, USA
- ⁴⁴ Cátedra de Inmunología, Facultad de Química, Instituto de Higiene, CP11600, Montevideo, URUGUAY
- ⁴⁵ Rheinische Friedrich-Wilhelms Universität Bonn, Institut für Biochemie und Molekularbiologie, 53115, Bonn, GERMANY
- ⁴⁶ Department of Biochemistry and Molecular Genetics, University of Illinois at Chicago, Chicago, IL 60607, USA
- ⁴⁷ Department of Nutritional Sciences, University of Wisconsin, Madison, WI, 53706, USA
- ⁴⁸ Department of Biological Sciences, Towson University, Towson, MD, 21252, USA
- ⁴⁹ Department of Genetics, University of Cambridge, Cambridgeshire, CB10 1SD, UK

Running Title: Selenoprotein gene nomenclature

To whom correspondence should be addressed: vgladyshev@rics.bwh.harvard.edu

Keywords: selenium; selenocysteine; selenoprotein; gene nomenclature

ABSTRACT

The human genome contains 25 genes coding for selenocysteine-containing proteins (selenoproteins). These proteins are involved in a variety of functions, most notably redox homeostasis. Selenoprotein enzymes with known functions are designated according to these functions: TXNRD1, TXNRD2, and TXNRD3 (thioredoxin reductases), GPX1, GPX2, GPX3, GPX4 and GPX6 (glutathione peroxidases), DIO1, DIO2, and DIO3 (iodothyronine deiodinases), MSRB1 (methionine-*R*-sulfoxide reductase 1) and SEPHS2 (selenophosphate synthetase 2). Selenoproteins without known functions have traditionally been denoted by SEL or SEP symbols. However, these symbols are sometimes ambiguous and conflict with the approved nomenclature for several other genes. Therefore, there is a need to implement a rational and coherent nomenclature system for selenoprotein-encoding genes. Our solution is to use the root symbol SELENO followed by a letter. This nomenclature applies to SELENOF (selenoprotein F, the 15 kDa selenoprotein, SEP15), SELENOH (selenoprotein H, SELH, C11orf31), SELENOI (selenoprotein I, SELI, EPT1), SELENOK (selenoprotein K, SELK), SELENOM (selenoprotein M, SELM), SELENON (selenoprotein N, SEP1, SELN), SELENOO (selenoprotein O, SELO), SELENOP (selenoprotein P, SeP, SEPP1, SELP), SELENOS (selenoprotein S, SELS, SEPS1, VIMP), SELENOT (selenoprotein T, SELT), SELENOV (selenoprotein V, SELV) and SELENOW (selenoprotein W, SELW, SEP1). This system, approved by the HUGO Gene Nomenclature Committee, also resolves conflicting, missing and ambiguous designations for selenoprotein genes and is applicable to selenoproteins across vertebrates.

INTRODUCTION

Selenium is an essential trace element in humans, which is present in proteins in the form of the 21st proteinogenic amino acid, selenocysteine (Sec). Sec is co-translationally inserted into a polypeptide chain in response to in-frame UGA codons directed by the Sec insertion sequence element, a stem-loop structure

in the 3'-UTRs of selenoprotein mRNAs. The human genome contains 25 selenoprotein genes (1), and selenoproteins are essential for embryo development and human health (2, 3). Among the selenoproteins, 13 have known functions; at least 12 of them serve as oxidoreductases, wherein Sec is the catalytic redox-active residue. The redox theme is also common for selenoproteins in other organisms (4).

The remaining 12 selenoproteins either have no known function, or their functions are only partially established. One of the selenoproteins, selenoprotein P (5), requires special mention as it has more than one Sec. It is a major plasma selenoprotein that delivers selenium primarily from the liver to other organs (6,7), and is involved in selenium transport and metabolism within organs. However, this protein also has an N-terminal Sec-containing thioredoxin domain similar to that found in most selenoproteins with known functions, which points to a potential redox function. Several other selenoproteins, including selenoproteins H, M, T, V, W and Sep15, also possess thioredoxin-like domains, suggesting redox-related functions (8).

Selenoproteins are not all homologous, but are characterized by their incorporation of Sec. Historically they have been given designations by the groups that discovered them, *e.g.*, owing to its presence in plasma the respective selenoprotein was named Selenoprotein P (9,10), or because of its size another protein was called the 15 kDa selenoprotein or Sep15 (11). However, some selenoproteins were identified independently by two or more groups, which created confusion and discrepancies in the field. For example, the same protein was named Selenoprotein R by one group (12), but discovered concurrently and designated by another group as Selenoprotein X (13). This protein was then functionally characterized (14) and renamed MsrB1 (for methionine-*R*-sulfoxide reductase 1) (15), but all three designations persist in the literature and/or databases. Another problematic example is the nomenclature used for thioredoxin reductases. The names for the first thioredoxin reductase, which had been known decades before its selenoprotein nature was discovered (16), are generally internally consistent, although they differ in the abbreviations used, *e.g.*, TR1 and TrxR1 (17). The second and third thioredoxin reductases

discovered, however, were named inconsistently by the authors, wherein the mitochondrial thioredoxin reductase was designated as TrxR2 (18) and TR3 (19), and the testis-specific thioredoxin-glutathione reductase has been alternatively labeled as TR2 (19), TrxR3 or TGR.

Designations are also confusing for several other selenoproteins. For example, Selenoprotein S was named SelS (1), but a later paper introduced the designation VIMP (20). Similarly, Selenoprotein H was named SelH (1), but also C11orf31, and Selenoprotein I was named SelI (1), but also called EPT1 (21). To avoid confusion, and at the instigation of the HUGO Gene Nomenclature Committee (HGNC), we describe a new standardized designation system for human (and other vertebrate) selenoproteins.

RESULTS AND DISCUSSION

Resolving the nomenclature of selenoprotein genes Human gene designations are approved by the HUGO Gene Nomenclature Committee (HGNC), and genes in other mammals follow the same designations. Selenoproteins have traditionally been published using SEL or SEP symbols followed by a letter or a number. Unfortunately, for naming the genes encoding these proteins, the SEL root was not an option as it was already approved for the selectin gene family; for example, *SELP* is the approved gene symbol for selectin P (P-selectin) and not Selenoprotein P. Some selenoprotein genes had been approved using the root SEP (i.e., *SEPN1*, *SEPP1*, *SEPWI*) but this could not be utilized for all selenoproteins as Selenoprotein T gene would then be *SEPT* or *SEPT1*, and *SEPT#* is already used for the septin genes. HGNC does not use the same root for unrelated groups of genes (e.g., SEL for selectins and selenoproteins) and does not endorse the use of multiple root symbols for genes sharing a common name (e.g., SEP and SEL for selenoprotein). With a view to solving these issues, HGNC approached selenoprotein researchers to propose a new unifying root symbol for all selenoprotein genes.

Proposal for a new nomenclature We propose that all selenoproteins (except those that have been functionally characterized, e.g., with enzymatic activity) use the root symbol SELENO

followed by a letter. This gene nomenclature is designed to highlight selenium, the key functional site in these proteins, and to provide a new and unambiguous root for these genes. The new nomenclature applies to 12 human selenoprotein genes as detailed in Table 1. Selenoproteins with known functions will continue to use the same designations (Table 2). Once functions are established for other selenoproteins, they may be renamed, as required. The proposed designations apply to the selenoprotein genes; although the same designations may be used for many of the encoded proteins, traditional names of selenoproteins, e.g. Selenoprotein P, may also be used.

Selenoprotein gene designation in other species The new HGNC nomenclature will automatically be used to designate orthologous selenoprotein genes in other vertebrates and extended to accommodate selenoprotein genes with no orthologs in human (22) (Table 3). Where vertebrate gene duplications have occurred, the additional paralogs will be named in line with the human genes, but with suffixes on the symbols, e.g., zebrafish *selenot1a*, *selenot1b* and *selenot2*. Selenoproteins are widespread in all three domains of life. Despite the fact that land plants, yeast, and some other species have lost selenoprotein biosynthesis pathways, a unifying nomenclature beyond vertebrates might be desirable. We suggest using the human nomenclature described in this paper for orthologs of vertebrate selenoprotein genes. This nomenclature may also be extended to accommodate additional selenoprotein genes as they are discovered. While we use human designations in this paper, we note that, according to HGNC guidelines, most vertebrates use all uppercase letters for genes and proteins (italics for genes), rodents use title case for genes (uppercase for proteins), *Xenopus* and zebrafish use lower case for genes and title case for proteins, and *Anolis* use lowercase for genes and uppercase for proteins.

Designations of proteins that do not contain selenocysteine There exists another class of selenium-containing proteins, those which contain a bound atom of selenium but do not

contain a UGA-encoded Sec, for which there is also ambiguous nomenclature. For example, the Selenium Binding Protein 1 (SBP1), also referred to as SELENBP1 or hSP56, is one such protein (23). The naming of such proteins will not be included in the new nomenclature as they lack Sec. Similarly, the machinery for Sec biosynthesis and insertion will not be renamed.

Implementation The new selenoprotein gene nomenclature has been approved by the HGNC, can be found on their website (<http://www.genenames.org/cgi->

[bin/genefamilies/set/890](http://www.genenames.org/cgi-bin/genefamilies/set/890)), and will be found in all major genomic resources in due course. We recommend that future publications primarily use the new SELENO designations, but supplement them (as secondary designations/synonyms) with the names previously used by the community. Once the new nomenclature is consistently used, the old designations will no longer be needed. We hope that other researchers in the field will join us in implementing this new nomenclature.

Conflict of Interest: The authors declare that they have no conflicts of interest with the contents of this article.

Author Contributions: The manuscript was drafted by VNG in consultation with other authors. All authors contributed to revisions and discussion.

REFERENCES

1. Kryukov, G. V., Castellano, S., Novoselov, S. V., Lobanov, A. V., Zehtab, O., Guigo, R., and Gladyshev, V. N. (2003) Characterization of mammalian selenoproteomes. *Science* **300**, 1439-1443.
2. Bösl, M. R., Takaku, K., Oshima, M., Nishimura, S., and Taketo, M. M. (1997) Early embryonic lethality caused by targeted disruption of the mouse selenocysteine tRNA gene (Trsp). *Proc. Natl. Acad. Sci. USA* **94**, 5531-5534.
3. Schweizer, U., and Fradejas-Villar, N. (2016) Why 21? The significance of selenoproteins for human health revealed by inborn errors of metabolism. *FASEB J.*, In Press.
4. Fomenko, D. E., Xing, W., Adair, B. M., Thomas, D. J., and Gladyshev, V. N. (2007) High-throughput identification of catalytic redox-active cysteine residues. *Science* **315**, 387-389.
5. Burk, R. F., and Hill, K. E. (2015) Regulation of selenium metabolism and transport. *Annu. Rev. Nutr.* **35**, 109-134.
6. Carlson, B. A., Novoselov, S. V., Kumaraswamy, E., Lee, B. J., Anver, M. R., Gladyshev, V. N., and Hatfield, D. L. (2004) Specific excision of the selenocysteine tRNA[Ser]Sec (Trsp) gene in mouse liver demonstrates an essential role of selenoproteins in liver function. *J. Biol. Chem.* **279**, 8011-8017.
7. Schomburg, L., Schweizer, U., Holtmann, B., Flohé, L., Sendtner, M., and Köhrle, J. (2003) Gene disruption discloses role of selenoprotein P in selenium delivery to target tissues. *Biochem. J.* **370**, 397-402.
8. Dikiy, A., Novoselov, S. V., Fomenko, D. E., Sengupta, A., Carlson, B. A., Cerny, R. L., Ginalski, K., Grishin, N. V., Hatfield, D. L., and Gladyshev, V. N. (2007) SelT, SelW, SelH, and Rdx12: Genomics and molecular insights into the functions of selenoproteins of a novel thioredoxin-like Family. *Biochemistry* **46**, 6871-6882.
9. Motsenbocker, M. A., and Tappel, A. L. (1982) A selenocysteine-containing selenium-transport protein in rat plasma. *Biochim. Biophys. Acta* **719**, 147-153.
10. Burk, R. F., and Gregory, P. E. (1982) Some characteristics of ⁷⁵Se-P, a selenoprotein found in rat liver and plasma, and comparisons of it with selenogluthione peroxidase. *Arch. Biochem. Biophys.* **213**, 73-80.

11. Gladyshev, V. N., Jeang, K. T., Wootton, J. C., and Hatfield, D. L. (1998) A new human selenium-containing protein. Purification, characterization, and cDNA sequence. *J. Biol. Chem.* **273**, 8910-8915.
12. Kryukov, G. V., Kryukov, V.M., and Gladyshev, V. N. (1999) New mammalian selenocysteine-containing proteins identified with an algorithm that searches for selenocysteine insertion sequence elements. *J. Biol. Chem.* **274**, 33888-33897.
13. Lescure, A., Gautheret, D., Carbon, P., and Krol, A. (1999) Novel selenoproteins identified in silico and in vivo by using a conserved RNA structural motif. *J. Biol. Chem.* **274**, 38147-38154.
14. Kryukov, G. V., Kumar, R. A., Koc, A., Sun, Z., and Gladyshev, V. N. (2002) Selenoprotein R is a zinc-containing stereo-specific methionine sulfoxide reductase. *Proc. Natl. Acad. Sci. USA* **99**, 4245-4250.
15. Kim, H. Y., and Gladyshev, V. N. (2004) Methionine sulfoxide reduction in mammals: characterization of methionine-R-sulfoxide reductases. *Mol. Biol. Cell* **15**, 1055-1064.
16. Holmgren, A. (1977) Bovine thioredoxin system. Purification of thioredoxin reductase from calf liver and thymus and studies of its function in disulfide reduction. *J. Biol. Chem.* **252**, 4600-4606.
17. Arnér, E. S., and Holmgren, A. (2000) Physiological functions of thioredoxin and thioredoxin reductase. *Eur. J. Biochem.* **267**, 6102-6109.
18. Lee, S. R., Kim, J. R., Kwon, K. S., Yoon, H. W., Levine, R. L., Ginsburg, A., and Rhee, S. G. (1999) Molecular cloning and characterization of a mitochondrial selenocysteine-containing thioredoxin reductase from rat liver. *J. Biol. Chem.* **274**, 4722-4734.
19. Sun, Q. A., Wu, Y., Zappacosta, F., Jeang, K. T., Lee, B. J., Hatfield, D. L., and Gladyshev, V. N. (1999) Redox regulation of cell signaling by selenocysteine in mammalian thioredoxin reductases. *J. Biol. Chem.* **274**, 24522-24530.
20. Ye, Y., Shibata, Y., Yun, C., Ron, D., and Rapoport, T. A. (2004) A membrane protein complex mediates retro-translocation from the ER lumen into the cytosol. *Nature* **429**, 841-847.
21. Horibata, Y., and Hirabayashi, Y. (2007) Identification and characterization of human ethanolaminephosphotransferase1. *J. Lipid Res.* **48**, 503-508.
22. Mariotti, M., Ridge, P. G., Zhang, Y., Lobanov, A. V., Pringle, T. H., Guigo, R., Hatfield, D. L., and Gladyshev, V. N. (2012) Composition and evolution of the vertebrate and mammalian selenoproteomes. *PLoS One* **7**, e33066.
23. Ansong, E., Yang, W., and Diamond, A.M. (2014) Molecular cross-talk between members of distinct families of selenium containing proteins. *Mol. Nutr. Food Res.* **58**, 117-123.
24. Gladyshev, V. N., Jeang, K. T., Wootton, J. C., and Hatfield, D. L. (1998) A new human selenium-containing protein: purification, characterization and cDNA sequence. *J. Biol. Chem.* **273**, 8910-8915.
25. Korotkov, K. V., Novoselov, S. V., Hatfield, D. L., and Gladyshev, V. N. (2002) Mammalian selenoprotein in which selenocysteine (Sec) incorporation is supported by a new form of Sec insertion sequence element. *Mol. Cell. Biol.* **22**, 1402-1411.
26. Hill, K. E., Lloyd, R. S., and Burk, R. F. (1993) Conserved nucleotide sequences in the open reading frame and 3' untranslated region of selenoprotein P mRNA. *Proc. Natl. Acad. Sci. USA* **90**, 537-541.
27. Vendeland, S. C., Beilstein, M. A., Yeh, J. Y., Ream, W., and Whanger, P. D. (1995) Rat skeletal muscle selenoprotein W: cDNA clone and mRNA modulation by dietary selenium. *Proc. Natl. Acad. Sci. USA* **92**, 8749-53.
28. Gasdaska, P. Y., Gasdaska, J. R., Cochran, S., and Powis, G. (1995) Cloning and sequencing of a human thioredoxin reductase. *FEBS Lett.* **373**, 5-9.
29. Gladyshev, V. N., Jeang, K. T., and Stadtman, T. C. (1996) Selenocysteine, identified as the penultimate C-terminal residue in human T-cell thioredoxin reductase, corresponds to TGA in the human placental gene. *Proc. Natl. Acad. Sci. USA* **93**, 6146-6151.

30. Rotruck, J. T., Pope, A. L., Ganther, H. E., Swanson, A. B., Hafeman, D. G., and Hoekstra, W. G. (1973) Selenium: biochemical role as a component of glutathione peroxidase. *Science* **179**, 588-590.
31. Flohé, L., Günzler, W. A., and Schock, H. H. (1973) Glutathione peroxidase: a selenoenzyme. *FEBS Lett.* **32**, 132-134.
32. Forstrom, J. W., Zakowski, J. J., and Tappel, A. L. (1978) Identification of the catalytic site of rat liver glutathione peroxidase as selenocysteine. *Biochemistry* **17**, 2639-2644.
33. Chambers, I., Frampton, J., Goldfarb, P., Affara, N., McBain, W., and Harrison, P. R. (1986) The structure of the mouse glutathione peroxidase gene: the selenocysteine in the active site is encoded by the 'termination' codon, TGA. *EMBO J.* **5**, 1221-1227.
34. Mills, G. C. (1957) Hemoglobin catabolism. I. Glutathione peroxidase, an erythrocyte enzyme which protects hemoglobin from oxidative breakdown. *J. Biol. Chem.* **229**, 189-197.
35. Günzler, W. A., Steffens, G. J., Grossmann, A., Kim, S. M., Ötting, F., Wendel, A., and Flohé, L. (1984) The amino-acid sequence of bovine glutathione peroxidase. *Hoppe-Seyler's Zeitschrift für physiologische Chemie* **365**, 195-212.
36. Chu, F. F., Doroshov, J. H., and Esworthy, R. S. (1993) Expression, characterization, and tissue distribution of a new cellular selenium-dependent glutathione peroxidase, GSHPx-GI. *J. Biol. Chem.* **268**, 2571-2576.
37. Takahashi, K., Akasaka, M., Yamamoto, Y., Kobayashi, C., Mizoguchi, J., and Koyama, J. (1990) Primary structure of human plasma glutathione peroxidase deduced from cDNA sequences. *J. Biochem.* **108**, 145-148.
38. Ursini, F., Maiorino, M., and Gregolin, C. (1985) The selenoenzyme phospholipid hydroperoxide glutathione peroxidase. *Biochim. Biophys. Acta* **839**, 62-70.
39. Brigelius-Flohé, R., Aumann, K. D., Blöcker, H., Gross, G., Kiess, M., Klöppel, K. D., Maiorino, M., Roveri, A., Schuckelt, R., Ursini, F., Wingender, E. and Flohé, L. (1994) Phospholipid Hydroperoxide Glutathione Peroxidase: Genomic DNA, cDNA and deduced amini acid sequence. *J. Biol. Chem.* **269**, 7342-7348.
40. Berry, M. J., Banu, L., and Larsen, P. R. (1991) Type I iodothyronine deiodinase is a selenocysteine-containing enzyme. *Nature* **349**, 438-440.
41. Behne, D., Kyriakopoulos, A., Meinhold, H., and Köhrle, J. (1990) Identification of type I iodothyronine 5'-deiodinase as a selenoenzyme. *Biochem. Biophys. Res. Commun.* **173**, 1143-1149.
42. Davey, J. C., Becker, K. B., Schneider, M. J., St Germain, D. L., and Galton, V. A. (1995) Cloning of a cDNA for the type II iodothyronine deiodinase. *J. Biol. Chem.* **270**, 26786-26789.
43. St Germain, D. L., Schwartzman, R. A., Croteau, W., Kanamori, A., Wang, Z., Brown, D. D., and Galton, V. A. (1994) A thyroid hormone-regulated gene in *Xenopus laevis* encodes a type III iodothyronine 5-deiodinase. *Proc. Natl. Acad. Sci. USA* **91**, 7767-7771.
44. Guimarães, M. J., Peterson, D., Vicari, A., Cocks, B. G., Copeland, N. G., Gilbert, D. J., Jenkins, N. A., Ferrick, D. A., Kastelein, R. A., Bazan, J. F., and Zlotnik, A. (1996) Identification of a novel selD homolog from eukaryotes, bacteria, and archaea: is there an autoregulatory mechanism in selenocysteine metabolism? *Proc. Natl. Acad. Sci. USA* **93**, 15086-15091.
45. Castellano, S., Lobanov, A. V., Chapple, C., Novoselov, S. V., Albrecht, M., Hua, D., Lescure, A., Lengauer, T., Krol, A., Gladyshev, V. N., and Guigo, R. (2005) Diversity and functional plasticity of eukaryotic selenoproteins: Identification and characterization of the SelJ family. *Proc. Natl. Acad. Sci. USA* **102**, 16188-16193.
46. Castellano, S., Novoselov, S. V., Kryukov, G. V., Lescure, A., Blanco, E., Krol, A., Gladyshev, V. N., and Guigo, R. (2004) Reconsidering the evolution of eukaryotic selenoproteins: a novel nonmammalian family with scattered phylogenetic distribution. *EMBO Rep.* **7**, 71-77.
47. Shchedrina, V. A., Novoselov, S.V., Malinouski, M. Y., and Gladyshev, V. N. (2007) Identification and characterization of a selenoprotein family containing a diselenide bond in a redox motif. *Proc. Natl. Acad. Sci. USA* **104**, 13919-13924.

48. Novoselov, S. V., Hua, D., Lobanov, A. V., and Gladyshev, V. N. (2006) Identification and characterization of Fep15, a new selenocysteine-containing member of the Sep15 protein family. *Biochem. J.* **394**, 575-579.
49. Kryukov, G.V., Gladyshev, V. N. (2000) Selenium metabolism in zebrafish: multiplicity of selenoprotein genes and expression of a protein containing 17 selenocysteine residues. *Genes Cells* **5**, 1049-1060.
50. Sunde, R. A., Sunde, G. R., Sunde, C. L., Sunde, M. L., Evenson, J. K. (2015) Cloning, sequencing, and expression of selenoprotein transcripts in the turkey (*Meleagris gallopavo*). *PLoS ONE* **10**, e0129801.

FOOTNOTES

The abbreviations used are: Sec, selenocysteine; TXNRD, thioredoxin reductases; GPX, glutathione peroxidase; DIO, iodothyronine deiodinase; MSRB, methionine-*R*-sulfoxide reductase; SEPHS2, selenophosphate synthetase 2; SELENOF, 15 kDa selenoprotein; SELENOH, selenoprotein H; SELENOI, selenoprotein I; SELENOK, selenoprotein K; SELENOM, selenoprotein M; SELENON, selenoprotein N; SELENOO, selenoprotein O; SELENOP, selenoprotein P; SELENOS, selenoprotein S; SELENOT, selenoprotein T; SELENOV, selenoprotein V; SELENOW, selenoprotein W; SBP1, Selenium Binding Protein 1; HGNC, HUGO Gene Nomenclature Committee.

Table 1. Selenoprotein genes using the new SELENO root. New HGNC selenoprotein gene nomenclature is indicated in the column “symbol”. Previous HGNC symbols (shown with *) will become synonyms, along with other previously used designations.

Symbol	Name	Synonyms	Refs
SELENOF	Selenoprotein F	selenoprotein 15, SEP15	24
SELENOH	Selenoprotein H	SELH, C11orf31*	1
SELENOI	Selenoprotein I	SELI, EPT1*	1,21
SELENOK	Selenoprotein K	SELK	1
SELENOM	Selenoprotein M	SELM, SEPM	25
SELENON	Selenoprotein N	SEPN1*, SELN	11
SELENOO	Selenoprotein O	SELO	1
SELENOP	Selenoprotein P	SEPP1*, SeP, SELP, SEPP	26
SELENOS	Selenoprotein S	SELS, SEPS1, VIMP*	1
SELENOT	Selenoprotein T	SELT	12
SELENOV	Selenoprotein V	SELV	1
SELENOW	Selenoprotein W	SELW, SEPW1*	27

Table 2. Selenoprotein genes named based on encoded enzymatic activity. Note that the nomenclature of these genes will not be changing to use the SELENO root.

Symbol	Name	Synonyms	Refs
TXNRD1	Thioredoxin reductase 1	TR1, TRXR1	16,28,29
TXNRD2	Thioredoxin reductase 2	TRXR2, TR3, mitochondrial Thioredoxin reductase	13,14
TXNRD3	Thioredoxin-glutathione reductase	TGR, TRXR3, TR2	14
GPX1	Glutathione peroxidase 1	Cytosolic glutathione Peroxidase, GSHPX1	30-35
GPX2	Glutathione peroxidase 2	GSHPX-GI	36
GPX3	Glutathione peroxidase 3	Plasma glutathione peroxidase	37
GPX4	Glutathione peroxidase 4	Phospholipid hydroperoxide glutathione peroxidase, PHGPX	38,39
GPX6	Glutathione peroxidase 6		1
DIO1	Iodothyronine deiodinase 1	D1	40, 41
DIO2	Iodothyronine deiodinase 2	D2	42
DIO3	Iodothyronine deiodinase 3	D3	43
MSRB1	Methionine- <i>R</i> -sulfoxide reductase 1	SELR, SELX, SEPX1	12-14
SEPHS2	Selenophosphate synthetase 2	SPS2	44

Table 3. Vertebrate selenoproteins absent in human and mouse. New HGNC selenoprotein gene nomenclature is indicated in the column “symbol”.

Symbol	Name	Synonyms	Refs
SELENOJ	Selenoprotein J	SELJ	45
SELENOU	Selenoprotein U	SELU	46
SELENOL	Selenoprotein L	SELL	47
SELENOE	Fish selenoprotein 15	FEP15	48
SELENOP2	Selenoprotein P2	SEPP2, SELPb	49,50

Selenoprotein Gene Nomenclature

Vadim N. Gladyshev, Elias S. Arnér, Marla J. Berry, Regina Brigelius-Flohé, Elspeth A. Bruford, Raymond F. Burk, Bradley A. Carlson, Sergi Castellano, Laurent Chavatte, Marcus Conrad, Paul R. Copeland, Alan M. Diamond, Donna M. Driscoll, Ana Ferreiro, Leopold Flohé, Fiona R. Green, Roderic Guigó, Diane E. Handy, Dolph L. Hatfield, John Hesketh, Peter R. Hoffmann, Arne Holmgren, Robert J. Hondal, Michael T. Howard, Kaixun Huang, Hwa-Young Kim, Ick Young Kim, Josef Köhrle, Alain Krol, Gregory V. Kryukov, Byeong Jae Lee, Byung Cheon Lee, Xin Gen Lei, Qiong Liu, Alain Lescure, Alexei V. Lobanov, Joseph Loscalzo, Matilde Maiorino, Marco Mariotti, K. Sandeep Prabhu, Margaret P. Rayman, Sharon Rozovsky, Gustavo Salinas, Lutz Schomburg, Ulrich Schweizer, Miljan Simonovic, Roger A. Sunde, Petra A. Tsuji, Susan Tweedie, Fulvio Ursini and Yan Zhang

J. Biol. Chem. published online September 19, 2016

Access the most updated version of this article at doi: [10.1074/jbc.M116.756155](https://doi.org/10.1074/jbc.M116.756155)

Alerts:

- [When this article is cited](#)
- [When a correction for this article is posted](#)

[Click here](#) to choose from all of JBC's e-mail alerts

This article cites 0 references, 0 of which can be accessed free at <http://www.jbc.org/content/early/2016/09/19/jbc.M116.756155.full.html#ref-list-1>