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TOPIC HIGHLIGHT

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Neoplastic disease after liver transplantation: Focus on *de novo* neoplasms

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Abstract

De novo neoplasms account for almost 30% of deaths 10 years after liver transplantation and are the most common cause of mortality in patients surviving at least 1 year after transplant. The risk of malignancy is two to four times higher in transplant recipients than in an age- and sex-matched population, and cancer is expected to surpass cardiovascular complications as the primary cause of death in transplanted patients within the next 2 decades. Since exposure to immunosuppression is associated with an increased frequency of developing neoplasm, long-term immunosuppression should be therefore minimized. Promising results in the prevention of hepatocellular carcinoma (HCC) recurrence have been reported with the use of mTOR inhibitors including everolimus and sirolimus and the ongoing open-label prospective randomized controlled SILVER. Study will provide more information on whether sirolimus-containing vs mTOR-inhibitorfree immunosuppression is more efficacious in reducing HCC recurrence.

Key words: Liver transplantation; *De novo* neoplasms; Immunosuppression; mTOR inhibitors; Hepatocellular carcinoma

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Core tip: With the notable increase in life expectancy after liver transplantation, together with the lengthy exposure to immunosuppression, transplant recipients are at risk of developing neoplastic disease, which accounts for almost 30% of deaths 10 years after liver transplantation. The risk of malignancy is two to four times higher in transplant recipients than in an age-



and sex-matched population, and cancer is expected to surpass cardiovascular complications as the primary cause of death in transplanted patients within the next 2 decades, making this an important topic for clinicians to consider.

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INTRODUCTION

With excellent long-term survival rates, the causes of morbidity and mortality of liver transplant (LT) recipients are primarily cardiovascular diseases, renal insufficiency, and de novo neoplasm, the latter of which account for almost 30% of deaths at 10 years post transplantation. Apart from hepatic causes, neoplasm has been reported as the most common cause of death in patients surviving at least 1 year after LT, and is responsible for approximately 40% of deaths^[1,2]. Overall, it is estimated that in LT recipients the incidence of neoplasms is between 3.1% and 14.4%, and the cancer-related mortality rate is between 0.6% and $8.0\%^{[3,4]}$.

Although the risk of some neoplasms including breast cancer (1.9 times lower) and genitourinary cancer (1.5 times lower) in women seem to be reduced compared to those of the general population^[5], in general terms, the status of transplant recipient is associated with an increased risk of developing *de novo* neoplasm. As shown in a study analyzing 1000 consecutive LT recipients in Pittsburgh and comparing this population's incidence of neoplasms compared to the general population, the former have a significantly elevated risk for developing neoplasm, which is 7.6 times higher for oropharyngeal cancer and 1.7 times higher for respiratory malignancies (Table 1).

Since a more prolonged exposure to immunosuppression is associated with an increased frequency of developing neoplasms, the cumulative risk of developing *de novo* malignancy rises from 20% at 10 years to 55% at 15 years after transplant^[6]. In an Italian study analyzing 313 LT recipients who survived more than 12 mo after transplant, during a total followup time of 1753 person-years, *de novo* malignancies were diagnosed in 40 (12.8%) subjects, with a median time from transplantation to diagnosis of 54 mo (range, 2-159 mo)^[7]. Other studies have reported a slightly lower mean interval between LT and diagnosis of nonlymphoid malignancies (36.2 mo, range, 5.8-74.1)^[5].

Not only are malignant neoplasms more frequent in transplant recipients, but they also have a more aggressive behavior, present at an earlier age compared to the non-transplant population, and take

Table 1 Estimated standardized incidence ratios for *de novo* malignancies after liver transplantation (data according to [7,9,15,46-48,61,72,174-182])

Cancer site/type	Estimated incidence (%)	SIR
All cancers	5-6	1.94-3
Kaposi's sarcoma	0.14-2.8	> 100
Skin (non melanoma)	0.9-3.2	> 30
PTLD	0.9-2.6	6-20
Gastrointestinal and		
oropharyngeal sites		
Lip/oropharyngeal/head and	0.1-2.0	5-14
neck cancers		
Esophagus ¹	0.5-1.19	12-18.7
Colorectal overall	0.0-0.65	1.41
Colorectal in IBD/PSC	0.7-7.9	3-5
Stomach	0.25	3
Vulva	0.25	8-23.8
Lung	0.6-1.2	2-8
Renal	0.35	2-2.65
Thyroid	0.20	4.60
Prostate	0.25-0.6	1 (risk not increased)
Breast	0.40	1 (risk not increased)
Colorectal in non-IBD/PSC	0.30	1 (risk not increased)

¹Although there are no population-based SIR estimates showing an increased risk of esophageal cancer after LT, an Italian study reported an SIR of 23.4 on the basis of cases ascertained by medical record reviews^[178]. This association may be related to prior alcohol exposure; 2 of 3 patients diagnosed with esophageal cancer in a US cohort underwent LT for ALD^[1]. IBD: Inflammatory bowel disease; PSC: Primary sclerosing cholangitis; PTLD: Posttransplant lymphoproliferative disease; SIR: Standardized incidence ratio.

a higher toll on survival^[8]. Mortality after diagnosis of *de novo* malignant neoplasms is particularly elevated, with reported rates as high as 55% and a median survival of 54 mo after diagnosis^[7]. Overall, estimated survival rates for all types of *de novo* malignancies are reportedly 70%, 56%, 48%, and 39% after 1, 3, 5, and 10 years, respectively. For certain types of cancer, mortality is particularly high, reaching 100% for lung cancer, 62.5% for esophageal and gastric cancers, 57% for head and neck cancer, 50% for post-transplant lymphoproliferative disorder (PTLD), and 50% for Kaposi Sarcoma (KS)^[7].

TYPES OF *DE NOVO* NEOPLASMS

De novo malignancies are neoplasms that develop after transplantation, including solid tumors such as pancreatic cancer, lung cancer, colorectal cancer, gastric cancer, esophageal cancer, renal cell carcinoma, bladder cancer, thyroid cancer, oral cancer, brain tumors and laryngeal cancer, as well as non-solid tumors, primarily PTLD/non-Hodgkin Lymphoma (NHL) and leukemia. According to a large German study analyzing the frequency and distribution of de novo neoplasms after LT^[9], 1 de novo malignancy is to be expected approximately every 120 person-years after LT (120 de novo malignancies/14490 person-years). It was also shown that cancer incidence rates for LT



recipients are almost twice as high as those for an age- and sex-matched general population. To quantify the risk that the status of transplant recipient conveys, cancer site-specific incidence rates in the transplant population are compared against the general population, with standardized incidence ratios (SIRs). Estimated SIRs for each malignancy, as well as the reported incidence are shown in Table 1. PTLD is the most frequent de novo malignancy after LT, accounting for approximately 20% of cases^[7]. Other common types of de novo malignant tumors include KS (17%), head and neck cancer (17%), esophageal tumors (12%), lung cancer (10%), gastric adenocarcinoma (7%), melanoma (5%), colorectal cancer (5%), cervical cancer (5%), and breast cancer (2%), as shown in a study from Northern Italy^[7].

Skin cancer

In a series of LT recipients with nonlymphoid de novo malignancies, skin cancer was reportedly the most common type of malignancy (22/57 patients with de novo cancer, representing 33.3%), including squamous cell carcinomas in 50%, basal cell carcinomas in 40.9%, and melanomas in 9.1%. Neoplasms were most frequent on the skin of the head, face, and neck (in 14 subjects), but there were also several cases of multiple site involvement, and the mean time to onset was 36.4 mo (range, 8.2-75.1 mo)^[5]. Another study demonstrated that the prevalence of pre-malignant and neoplastic cutaneous lesions increased with time, with a frequency of premalignant lesions of 5% at 2-3 years, 12% at 3-5 years, 28% beyond 5 years, and frequency of malignant lesions of 0% at 2-3 years, 9% at 3-5 years, and 12% beyond 5 years of follow-up after transplantation. Furthermore, in that same study, the cumulative incidence of cutaneous lesions was significantly higher in patients treated with cyclosporine compared to recipients on tacrolimus^[10]. One-year survival after diagnosis of skin cancer in LT recipients is reportedly 90.9%^[5]. Several factors have been identified as being considered high risk for developing skin cancer, including increased age, increased intensity and longer duration of immunosuppressive therapy, infection with human papillomavirus, history of increased ultraviolet exposure, easily burned skin, history of actinic keratosis, CD4 lymphocytopenia, and blue or hazel eyes^[11,12]. Primary sclerosing cholangitis^[13] as well as alcohol-related liver disease as indications for LT are associated with a higher risk of skin malignancies compared to other etiologies of liver disease^[14,15]. Other risk factors for the development of skin malignancy after LT include male sex, age over 55 years, Caucasian background, and monoclonal antibody induction therapy[11], while the use of polyclonal or interleukin (IL)-2 receptor antibody induction therapy, treatment for rejection, and non-cholestatic etiologies of liver disease as indications for LT, seem not to be associated with an increased risk.

PTLDs

PTLD encompasses a heterogeneous group of diseases characterized by excessive proliferation of lymphoid cells and it commonly results from de novo infection or reactivation of latent Epstein-Barr virus (EBV)[16,17], especially in the case of EBV seronegative recipients of organs from EBV seropositive donors. LT carries an intermediate risk of PTLD, in contrast with intestinal transplantation, which has the highest rates[18,19]. An increased intensity of immunosuppression^[5,20-23] and the use of certain types of immunosuppressive agents, in particular T-cell depleting antibodies such as Muromonab-CD3 (OKT3) or anti-thymocyte globulin (ATG), cyclosporine, and belatacept (in renal transplant recipients) constitute additional risk factors for PTLD development^[24-26]. In an Italian study, 15 cases of PTLD were described in 1011 solid organ transplant recipients; in 13/15 patients, induction immunosuppressive therapy with OKT3 was used, and EBV was detected in 10 of 13 patients in whom neoplastic tissue was available for analysis. Moreover, in 2 of the 3 patients who were negative for EBV, hepatitis C virus (HCV) was present, and positivity for HCV was significantly more frequent in patients who developed PTLD compared to those who did not, suggesting a possible role of HCV in the development of PTLD^[19]. Other studies have also shown a correlation between the presence of HCV and the development of PTLD^[27-29].

In the pediatric population, PTLD is the most common tumor in solid organ recipients, with an overall incidence rate of 5% to 15% in different series or 298/100000 posttransplantation years of follow-up^[30,31]. Reported mortality is unfortunately very high, of up to 60%, especially in infants who develop PTLD as a result of primary EBV transmission from EBV-positive allograft transplant^[32-35].

The most important risk factors for PTLD development in the pediatric population include high levels of immunosuppression (especially associated with tacrolimus-based regimens^[36]), young age, time from transplant (related to longer exposure time to immunosuppression), EBV seronegativity before transplant, and primary EBV transmission. Fukushima and collaborators, in a recently published study on 32 infants younger than 2 years who had undergone livingdonor liver transplantation and were on tacrolimusbased immunosuppression, found that deteriorated tacrolimus metabolism (with elevated plasmatic levels) accompanied by an increase in Epstein-Barr viral load was more frequently associated with PTLD^[36]. In a recently published paper by the Studies of Pediatric Transplantation Research Group^[37] analyzing a large multicenter cohort of pediatric patients who underwent LT, transplants performed in the era 1995-2001 (vs those performed between 2002 and 2007), recipient EBV status, and frequent rejection episodes were associated with symptomatic EBV infection and

PTLD. The subgroup at a highest risk is constituted by younger infants with multiple rejection episodes. Importantly, the incidence of both symptomatic EBV infection and PTLD are seemingly decreasing in pediatric LT recipients, concomitantly with a reduction in immunosuppression^[37].

In a recent study, Khedmat and Taheri^[38] reviewed 250 cases of PTLD after liver transplantation published in the literature, of whom 212 were pediatric cases (18 years of age or less). PTLD was diagnosed at a mean age of 9.9 years and the mean \pm SD interval between LT and diagnosis of PTLD was 28.7 mo (35.1 mo). Organs/areas involved included: orbit, skin, stomach, genitalia, central nervous system, spleen, kidneys, respiratory system, liver, bone marrow, small intestine, and colon; in comparison with their adult counterparts, histopathological features of PTLD were significantly of more benign types.

Analogous to management strategies in adults, a sequential approach is employed, starting with reduction or complete withdrawal of immunosuppression, initiation of inferferon-alpha, various chemotherapic regimens, surgery, and radiotherapy, escalating strategies if the previous alternative proves inefficacious^[39]. Moreover, long-term withdrawal of immunosuppression has been shown to be feasible without graft rejection^[40]. The use of the anti-B-cell monoclonal antibody rituximab has brought about improved results, and more recently, Gupta and collaborators reported on satisfactory outcomes employing a dual combination of rituximab and reduced dose chemotherapy, with two-year failure-free survival of 57% in liver transplant recipients^[39].

Kaposi's sarcoma

KS is a multifocal angioproliferative mucocutaneous neoplasm driven by HHV-8 infection and represents approximately 4% of all post-transplant tumors. The risk of developing this neoplasm is increased 500-fold in solid organ transplant recipients compared with the general population^[41,42]. In a large study on 2705 recipients of solid organs, amongst whom 159 LT recipients, KS was diagnosed in 1.44% of all transplant recipients, including 12.8% of LT recipients^[43]. Contrary to most other neoplasms, the incidence of KS seems to decrease significantly with time after solid organ transplantation^[44]. In the presence of infection with HHV-8, the most important risk factor for the development of this neoplasm is the intensity of immunosuppression, and its therapy is based on immunosuppression tapering, as well as the use of chemotherapeutic agents. Moreover, evidence is mounting on the usefulness of mTOR inhibitors in treating this tumor while at the same time providing effective immunosuppression^[45].

Solid tumors

Lung cancer: The incidence of lung cancer among

LT recipients is increased compared to the general population, and reportedly accounted for 15.7% of nonlymphoid neoplasms in a series of LT recipients, in whom it was diagnosed, on average, 48.5 mo (range, 11.2 to 64.3 mo) after LT, and a one-year survival of 37.5%^[5]. In large case series of LT recipients, the mean time to diagnosis ranges from 42 to 50 mo^[5,46-48]. Akin to the association between smoking observed in the general population, this carcinogen is correlated with an increased risk of lung cancer in transplant recipients^[5,46]. Although probably representing an epidemiological association, as smokers are also frequently heavy drinkers, a study showed that patients with alcohol-related cirrhosis as an indication for LT had higher rates of lung cancer than those who underwent LT for other indications^[49].

Head and neck cancers: Head and neck neoplasms are more frequent in the LT population than in the general population, and mean time to diagnosis is reportedly between 34.3 mo and 61.2 mo^[5,15,47,50,51]. Oropharyngeal cancer is 25.5 times more frequent in patients transplanted for alcohol-related cirrhosis vs those transplanted for other indications^[52]. Moreover, upper aerodigestive squamous carcinomas are more frequent in patients with alcohol-related cirrhosis as the main indication for LT^[53]. Moreover, another study showed that whereas the incidence of oropharyngeal cancer was 16.7% in patients who underwent LT for alcohol-related liver disease, none of the patients who underwent LT for indications other than alcoholrelated cirrhosis developed oropharyngeal malignant neoplasms $(P = 0.001)^{[50]}$. Notably, there was not one case of oropharyngeal cancer in a small, single-center study involving patients without a history of smoking or alcohol use^[54]. Likewise, tongue cancer and laryngeal cancer have been reported in smokers^[5,46], and the carcinogenic effects of tobacco observed in the general population also applies for transplant recipients. It is difficult to establish the weight of alcohol compared to tobacco use as contributing risk factors for head and neck neoplasms, as alcohol is known to potentiate the carcinogenic effects of smoking^[55], and also since patients who are heavy smokers also tend to be heavy drinkers^[56].

Esophageal and gastric cancer: Although their incidence is increased with respect to the general population^[57], gastric and esophageal cancers are reported infrequently in most series of LT recipients^[58]. As well as for several other types of cancer, notably those of the oropharynx/larynx, alcohol is a wellestablished risk factor for esophageal malignant neoplasms^[59], and this neoplasm occurs at a higher rate after LT in patients with alcohol-related liver disease^[15,27,60]. In an Italian study on 313 LT recipients followed during a 15-year period, of 40 patients with *de novo* malignancy, esophageal cancer was diagnosed

in 12%, with a mortality (combined for esophageal and gastric cancer of 62.5%) being second only to that of lung cancer^[7]. A German study analyzing 1,926 LT recipients found that 9 patients (0.5%) developed a de novo esophageal cancer and 1 patient developed cancer of the cardia (0.05%), diagnosed on average 51 mo after LT. The histological type of tumor was squamous cell carcinoma in 7/10 and adenocarcinoma in 3/10. Of note, 9/10 patients had undergone LT due to alcohol-related cirrhosis^[61]. A predisposing lesion, Barrett's esophagus, has been demonstrated to rapidly evolve into adenocarcinoma after LT, which is why surveillance endoscopy with aggressive endoscopic treatment of Barrett's mucosa is paramount in these patients to prevent death from cancer^[62-66]. In a Korean study of 6491 patients who underwent solid organ transplantation, 30 patients (0.46%) with 31 lesions were diagnosed with gastric cancer^[67]. In another series, 36 cases of gastric cancer were identified among 7000 transplant-related malignant neoplasms, and 3 of the 34 were observed in LT recipients^[68]. Moreover, another study reported 3 cases of gastric cancer amongst 329 cases of malignant neoplasms in LT recipients^[69].

Genitourinary cancer: Although the incidence of prostate cancer does not seem to be increased in LT recipients, all other genitourinary cancers (including bladder and renal cancer) seem to be higher than that of the general population^[5,15,27,46,47]. Mean time to diagnosis of non-prostate genitourinary cancer ranges from 20 to 55.3 mo, while in cases of prostate cancer the diagnosis is often performed between 5.8 and 18.4 mo after LT^[5,15,47,48]. In LT recipients, prostate cancer is more often diagnosed at earlier stages and has a good prognosis, whereas renal and bladder cancers have a poor prognosis^[5].

Gynecological cancer: Although it seems that breast cancer is no more frequent in LT compared to the general population^[3], non-breast gynecological cancers (cervical and ovarian) are more frequent in LT recipients than in the general population^[15,46,47]. It has been hypothesized that rigorous screening before LT has contributed to a tendency, albeit not statistically significant, for a lower incidence of breast cancer in LT recipients^[5]. However, other studies have documented that breast cancer incidence is in fact elevated in the transplant population, with the advantage, however, that early detection is more common, and this has also resulted in decreased mortality compared to that of the general population upon similar diagnoses^[46].

Colorectal cancer: The incidence of colorectal cancer seems to be higher in the LT recipient population *vs* the general population^[46,47], although most of this difference in incidence, if not all, can be accounted for by the increased risk of colorectal cancer associated

with LT for primary sclerosing cholangitis, probably due to the association with ulcerative colitis^[70-72]. More frequently diagnosed between 16 and 50 mo after transplant, colorectal cancer in transplant recipients tends to be detected at an earlier age and has been associated with a worse prognosis compared to the general population^[73,74].

De novo hepatocellular carcinoma: A search performed by Trevisani et al^[75] identified 14 cases of de novo hepatocellular carcinoma (HCC) which have been reported in the literature. Although until now a relatively rare occurrence, truly de novo HCC, that is, neoplasms arising from the liver graft and not recurrences of recipient HCC, might be seen more often in the future, due to the increased use of extended criteria grafts, especially those from older donors, donors carrying HCV or HBV infection, or alcoholic liver disease^[76,77]. One of the principal risk factors for de novo HCC is recurrence of liver disease in the allograft, and especially the development of cirrhosis^[75], and reported cases have been diagnosed on average 2 years after LT. As for non-transplant recipients, post transplant exposure to hepatocarcinogens like aflatoxin B1, nitrosamine, aromatic amines, vinyl chloride, azo-dyes, pesticides, arsenic, organic solvents, and cigarette smoking, can theoretically trigger the development of HCC, although no case has yet been reported in association with any of these factors. Immunosuppression regimens used in the 14 reported cases include OKT3, azathioprine, cyclosporine, corticosteroids, mycophenolate mofetil, basiliximab, and tacrolimus^[78-82].

Prognosis seems dismal according to reported cases, despite tapering of immunosuppression, transarterial chemoembolization, radiofrequency ablation, hepatic resection, or retransplantation. Strategies for preventing this neoplasm include avoidance of recurrent graft damage as well as a judicious immunosuppression after LT^[75]. While HCC recurrence is considered a contraindication for retransplantation, this therapeutic option could be contemplated in the setting of *de novo* HCC and has been reported in a case with development of this *de novo* malignancy 14 years after primary LT^[82].

RISK FACTORS FOR THE DEVELOPMENT OF *DE NOVO* MALIGNANCIES

In a study analyzing risk factors for the development of solid neoplasms after LT, multivariate analyisis demonstrated that primary sclerosing cholangitis (HR = 2.62, 95%CI: 1.50-4.56), alcohol-related cirrhosis (HR = 2.14, 95%CI: 1.22-3.73), smoking (HR = 1.72, 95%CI: 1.06-2.79), and increasing age in decades (HR = 1.33, 95%CI: 1.05-1.66) were all significantly associated with *de novo* neoplasms^[1]. A summary of the most important risk factors is provided in Table 2.



Table 2 Risk factors for the development of *de novo* malignancies according to tumor location/type (data according to [5,14-17,20-22,25,26,46,48,50,53,54,61,62,64,75,130,181,183,184])

Tumor location/type	Risk factor	
Skin	Age > 40 yr	
	Male gender	
	Skin type	
	Sun exposure	
	Smoking	
	Alcoholic cirrhosis	
	Primary sclerosing cholangitis as indication	
	for LT	
	Cyclosporine-based immunosuppression	
KS	Increased intensity of immunosuppression	
	Infection with HHV-8	
PTLD	Age > 50 yr	
	Infection with EBV (especially seronegative	
	recipients of organs from EBV seropositive	
	donors)	
	Increased intensity of immunosuppression	
	OKT3 or anti-thymocyte globulin	
	Cyclosporine-based immunosuppression	
	Hepatitis C virus	
Lung cancer	Cigarette smoking	
	LT for alcohol-related liver disease	
Head and neck cancers	Cigarette smoking	
	LT for alcohol-related liver disease	
Esophageal and gastric	LT for alcohol-related liver disease	
cancers	Barrett's Esophagus	
Colorectal cancer	Primary sclerosing cholangitis	
	Inflammatory bowel disease	
De novo HCC	Recurrence of liver disease in the allograft	
Gynecologic cancers	Insufficient evidence	
Genitourinary cancers	Insufficient evidence	

EBV: Epstein-Barr virus; HCC: Hepatocellular carcinoma; HHV-8: Human herpesvirus 8; KS: Kaposi's sarcoma; LT: Liver transplantation; PTLD: Post-transplant lymphoproliferative disorder.

DONOR-TRANSMITTED MALIGNANCIES

The role of immunosuppression in reactivating dormant neoplasms is supported by the fact that transplant recipients who have received organs from donors with previously cured neoplasms may develop the donor' s malignancy^[83,84]. Reportedly, 0.5% to 3% of donors have a history of malignancy, and transmission from these donors to the recipients has been demonstrated in 0.02%-6% of cases^[85-89], the risk being higher in LT recipients as compared to recipients of other organs^[90,91]. According to the time elapsed from clinical remission of the neoplasm in the donor to the moment of donation, tumor site, and risk of transmission, recommendations for specific tumor types have been issued by the Malignancy Subcommittee of the Disease Transmission Advisory Committee of the Organ Procurement and Transplantation Network/United Network for Organ Sharing (OPTN/UNOS). Organ shortage, a low risk of transmission of malignancy to the recipient, and the need for a life-saving transplant in cases of urgent LT may drive the decision of using organs from extended criteria donors, including donors with a neoplasm. It is important, however, to quantify the risk, based on the type of neoplasm. Thus, an organ from a donor with basal cell carcinoma is considered to be associated with a minimal risk (< 0.01%) of transmission and may be used as a graft, whereas at the other end of the spectrum, the history or presence of melanoma, lung cancer, or active breast cancer > stage 0 are considered at high risk of transmission (> 10%) and their use is discouraged^[92]. Allegedly, organs from donors with central nervous system malignancies may be safely transplanted; in a study analyzing 62 recipients of organs from donors with a history of or active central nervous system neoplasm, 8 transmissions were identified, occurring 2-15 mo after transplant, with seven patients dying as the result of metastatic disease. The presence of one or more risk factors, identified as: high-grade tumors, ventriculoperitoneal or ventriculoatrial shungs, prior craniotomy and systemic chemiotherapy, entailed a risk of 53% of tumor transmission, whereas the rate was significantly lower (7%, P < 0.01) if no risk factor was present^[93]. However, a more recent and larger study performed in the United Kingdom concluded that organs from donors who died as a consequence of primary intracranial malignancy, including those with high-grade tumors, should be considered for transplantation due to the small risk of tumor transmission. Identification of 448 recipients of 495 organs from 177 donors with primary intracranial malignancy, including 33 with high-grade malignancy (9 medulloblastomas and 24 grade IV gliomas amongst 179 donors) demonstrated not one single case of tumor transmission^[94]. As in all medical interventions, a risk-benefit evaluation must be performed, the patient should be informed of the possibility of receiving one such organ, and this must be weighed against the risk of dying on the waiting list, which is much higher.

The recommendations for screening in the donor so as to reduce the risk of undiagnosed neoplasm and subsequent transmission to the recipient include execution of complete medical history specifically inquiring on previous diagnosis of malignancy, radiological imaging, complete physical examination to rule out possible skin cancer, laboratory analysis for the detection of tumor markers, pathology examination of extracted organs, and in cases of unexplained intracranial hemorrhage and in women with menstral disorders, underlying neoplasms must be excluded [95,96].

IMMUNOSUPPRESSION

Immunosuppression plays a fundamental role in the development of neoplasms, acting through several different mechanisms including decreased immune surveillance, increased susceptibility to infections, induction of insulin resistance, and a direct carcinogenic effect which has been described in the case of some immunosuppressive agents. The association between

alterations in the immune system and the development of neoplasms is also reflected in the elevated incidence of cancer in most medical conditions associated with immunosuppression^[97,98] and the fact that the length of exposure and intensity of immunosuppression correlate with the incidence of malignant neoplasms^[99,100]. Whereas in immunocompetent subjects there is continuous ongoing surveillance that acts as tumor suppressor, keeping in check possible accumulated cell damage resulting in neoplasms, immunosuppression in organ transplant recipients results in a lower threshold for immunosurveillance, allowing neoplastic cells to proliferate.

Moreover, chronic immunosuppression renders transplant recipients more vulnerable to viral infections, some of which have oncogenic potential. Although not all neoplasms are the result of viral triggers, the ones that are tend to be those that show the greatest rise in frequency amongst transplant recipients including B-cell lymphoma and PTLD (EBV), squamous cell skin carcinoma (HPV), Kaposi's sarcoma (HHV8), anogenital cancers (HPV), Merkel skin cancer (polyomavirus), and HCC (HBV, HCV)[97]. The viral oncogenic potential may be enhanced by the action of some immunosuppressants. Calcineurin inhibitors in particular, can favor the expression of EBV growth and virus-inducing factors including IL-1, IL-6, and transforming growth factor (TGF-β), can promote EBV replication, and can augment immunoresistance by favoring the expression of anti-apoptotic genes^[101].

Aside from these indirect effects, several immunosuppressive drugs seem to have direct oncogenic effects, either by provoking damage to DNA or through other mechanisms not linked to immunosuppression. Azathioprine, for instance, induces chromosomal aberrations and increases skin cell sensitivity to photodamage^[97].

Calcineurin inhibitors: There is evidence of direct prooncogenic activity in the case of calcineurin inhibitors, which induce tumorigenesis and tumor growth by inducing cancer cell invasiveness^[102], hampering DNA repair mechanisms^[103,104] and apoptosis^[103], inducing tumor angiogenesis *via* the stimulation of vascular endothelial growth factor (VEGF)^[105], and promoting the transcription and functional expression of the TGF- β 1 gene which results in tumor cell invasion and metastatic potential^[106]. In LT recipients, it has been shown than exposure to elevated concentrations of tacrolimus (> 20 ng/mL) in the weeks immediately after transplantation increases long-term mortality due to infections, cardiovascular events and development of neoplasms^[107-110].

Furthermore, both calcineurin inhibitors and steroids exert a diabetogenic effect, causing impaired insulin secretion and inducing pancreatic beta cell apoptosis^[111-113]. As many as 5%-27% of LT recipients develop neo-onset diabetes mellitus, and it is associated with a negative

impact on patient and graft survival^[114-116], diabetes being a recognized risk factor for neoplasms, playing an important role especially in HCC^[117]. Calcineurin inhibitors, especially tacrolimus, have in fact been shown to increase the risk of developing new-onset diabetes mellitus after transplantation.

Other immunosuppressant agents: The use of other immunosuppressant agents, including OKT3 and ATG, has also been associated with an increased risk for the development of neoplasms after solid organ transplantation. Early PTLD has been shown to occur shortly after administration of OKT3, with an average of 7 mo from transplantation and/or administration to diagnosis of PTLD[118]. In other series, high total doses of OKT3, especially in individuals in whom a second course of therapy was administered, were associated with a higher frequency of lymphomas^[119,120]. In contrast, a single-center study reporting on 1570 LT of whom 125 patients developed de novo tumors, did not show any relationship between OKT3 and the development of de novo neoplasms; the authors note that this is consistent with the concept that chronic maintenance immunosuppression is more important than short albeit intense periods of immunosuppression (treated with OKT3)^[47]. A recently published Cochrane Database Systematic Review evaluated the benefits and harms of immunosuppressive T-cell specific antibody induction compared with placebo, no induction, or another type of T-cell specific antibody induction for prevention of acute rejection in LT recipients, and included studies using T-cell specific antibodies polyclonal antibodies [rabbit of horse antithymocyte globulin (ATG), or antilymphocyte globulin (ALG)], monoclonal antibodies (OKT3, anti-CD2, or alemtuzumab), and IL-2 receptor antagonists (daclizumab, basiliximab, BT563, or Lo-Tact-1). The authors concluded that there were no statistically significant differences in terms of malignancy[121].

Mammalian target of rapamycin inhibitors:

Mammalian target of rapamycin (mTOR) is a serine/threonine protein kinase downstream of the phophoinositide-3-kinase-related kinase family, which plays a fundamental role as regulator of various oncogenic processes including cell growth, proliferation, metabolism, and angiogenesis[122]. The combination of anti-tumoral as well as immunosuppressive properties render this family of drugs very attractive in the posttransplantation setting. There is growing evidence that the incidence of neoplastic disease is inferior in patients with gradual reduction of CNI with the introduction of mTOR inhibitors, vs those subjects treated with standard-dose CNI^[123]. An anti-neoplastic activity has been demonstrated for everolimus with regard to various solid tumors, and a potential role in HCC and cholangiocarcinoma are being increasingly reported[105,124-127]

Table 3 Intensive screening protocols for tumor surveillance in liver transplant recipients (data according to [128-130])

Traditional screening	Intensive screening	
Annual chest X-ray	Annual chest and abdominal CT	
Annual abdominal ultrasound	Annual abdominal ultrasound	
Chest and abdominal CT	Annual urologic screening with PSA	
	determination	
Mammography and urologic	Annual Pap smear and mammography	
screening (with timing	(every 1-2 yr)	
according to standard of care)	Annual skin examination	
	Colonoscopy 1 year after LT in patients	
	with adenoma on pre-LT colonoscopy,	
	and repeated every 2-4 yr if more	
	adenomas are found. Colonoscopy	
	repetition every 10 yr in patients	
	> 50-yr-old	
	Ears, nose and throat clinic visit in	
	patients with > 20 pack year smoking	

CT: Computed tomography; PSA: Prostatic specific antigen; LT: Liver transplant.

PREVENTION

As most neoplasms are favored by immunosuppression, the long-term use of the lowest effective dose of immunosuppression to avoid rejection are recommended, as well as the avoidance of excessive sun exposure, treatment of premalignant lesions including warts and actinic keratoses, and avoidance of exposure to confirmed carcinogenic substances including those present in tobacco smoke.

Screening protocols are recommended in order to detect malignancies in early states, increasing the probability of opportune treatment and improving prognosis^[128,129]. Some recommended strategies include monthly skin autoexam, annual dermatological visit, annual Pap smear, mammography every 2 years, annual digital rectal exam and prostate-specific antigen determination, annual fecal occult blood test, colonoscopy every 10 years, annual chest X-ray, abdominal ultrasound, chest and abdominal CT scan^[130-135]. A summary of preventive measures is provided in Table 3.

Management of neoplastic disease in LT recipients: In general terms, management of malignant neoplasms in LT recipients is similar to that of the immunocompetent patient in terms of surgery, chemotherapy and radiotherapy, but, in contrast, one of the main pillars of the approach to a neoplasm in transplant recipients is represented by modification of immunosuppression, especially in tumors which are highly susceptible to immunosuppression, such as KS and PTLD.

Owing to their strong anti-angiogenic effects which result in inhibition of tumor growth, as well as their direct action on cancer cells by the inhibition of their dependence on the mTOR pathway for cell growth and survival, mTOR inhibitors are increasingly being

used in the management of neoplasms in transplant recipients^[105,136,137]. Since cyclosporine favors an invasive and aggressive tumor cell behavior, the combination with mTORi was hypothesized to be beneficial, adequately avoiding rejection while also providing malignancy control. This has been proven to be true, with significantly better survival times with mTORi plus cyclosporine treatment vs cyclosporine-only treatment in mice injected with tumor cells^[138]. In fact, mTORi alone or mTORi plus cyclosporine impairs immunity and promotes allograft survival in experimental models, and the combination of sirolimus and everolimus with cyclosporine is effective in clinical transplantation, being approved by the Food and Drug Administration (FDA) for use in transplant recipients^[139]. Specifically, everolimus is indicated for immunosuppression kidney heart and liver transplantation, while sirolimus has been approved for kidney transplantation. Malignancy rates post-conversion to sirolimus-based, CNI-free, immunosuppression regimen were significantly lower with respect to the CNI-based immunosuppression protocol in the RMR study and the CONVERT trials^[140,141]. Moreover, experience in renal transplant recipients has demonstrated that the risk of de novo malignancies is significantly lower in patients treated with mTOR inhibitors (with or without CNIs) compared to patients on CNI-based regimens^[142]. Thus, one of the recommended strategies in the management of post-transplant neoplasms is the conversion from CNIs to mTOR inhibitors or inclusion of mTOR inhibitors in a CNI-based immunosuppressive regimen^[143-145]. Furthermore, in another study reporting on 10 LT recipients who had developed de novo neoplasms after LT, everolimus treatment significantly increased the probability of survival from 14% (in a similar historical cohort of patients not treated with everolimus) to 72% at 20 mo^[146]. Moreover, in a recently published retrospective study analyzing prognostic factors for patients transplanted for alcohol-related cirrhosis who developed non-cutaneous de novo solid organ neoplasms, conversion to everolimus improved prognosis, with one- and five-year survival rates of 77.4% and 35.2% in patients converted to everolimus vs 47.2% and 19.4% in patients not treated with everolimus, respectively $(P = 0.003)^{[147]}$.

RECURRENCE OF NON-HEPATIC NEOPLASMS

With the broadening of eligibility criteria for LT, older patients are now being transplanted, increasing the probability of patients with past medical history of malignancy to be evaluated for LT, waitlisted, and transplanted. The risk of neoplastic recurrence upon commencement and maintenance of immunosuppression and its derived mortality must be weighed against the probability of survival without a transplant. Recurrence of a preexistent neoplasm can occur after



LT, and according to the risk of recurrence, neoplasms can be classified as low recurrence risk (0%-10%) as in the case of cervical carcinoma, endometrial carcinoma, myeloproliferative disorders, and lymphomas; intermediate recurrence rate (11%-25%) as in the case of colorectal cancer, non-melanoma skin cancer, and thyroid carcinoma; and neoplasms with a high recurrence rate (> 26%) as in the case of oral squamous carcinoma and breast cancer^[148]. There is consensus that the tumor type and stage of the disease must be carefully evaluated, and according to this, recommendations have been made regarding the waiting time between achieving clinical "cure" or disease control and LT^[149-151]. According to American^[151] and European^[95] guidelines, proposed malignancyfree delay periods before transplantation vary from no delay in cases of basal-cell skin cancers and incidental renal cell carcinoma, to less than 2 years in cases of small single focal neoplasms, low-grade bladder cancer, excised squamous cell carcinoma, 2 years in cases off testicular and thyroid neoplasms, to 2-5 years or more for malignant melanomas, breast cancer, invasive cervical cancer, and colorectal cancer. Nevertheless, since many patients being evaluated for LT are too sick to endure a long waiting period, provided that the neoplasm is adequately controlled and the stage of the neoplasm itself is not associated with a poor prognosis, LT may be considered before completion of the waiting period with informed consent of the candidate^[152].

HCC RECURRENCE IN LT RECIPIENTS

In spite of the 5-year 60%-80% disease-free survival rate after LT for HCC in cases with unresectable early stages of the neoplasm, recurrence does occur in 3.5%-21% of cases, and is associated with a poor prognosis^[153]. Tumor-related established risk factors for HCC recurrence after LT include high levels of alpha-fetoprotein^[154,155], tumor grading^[156,157], tumor stage^[154,156-158], and vascular invasion^[154,157,158], while immunosuppression-related risk factors for HCC recurrence are primarily the level of immunosuppression^[156], mTOR- vs mTOR inhibitor-free immunosuppression regimen^[154,159]. Clinical studies have shown a CNIs dose-dependent increase in the risk of developing HCC recurrence^[102]. Elevated exposure to CNIs (mean trough concentrations of tacrolimus > 10 ng/mL or cyclosporine > 300 ng/mL) during the first postoperative period has in fact been associated with an increased risk of HCC recurrence^[160]. Moreover, it has been observed that high doses of cyclosporine are associated with a lower recurrence-free survival in patients transplanted for HCC. In fact, a study on 219 patients transplanted for HCC undertaken in Milan revealed that elevated doses of cyclosporine or tacrolimus during the first 30 d after LT almost tripled the risk of HCC recurrence^[127].

In contrast, mTOR inhibitors possess anti-angiogenic and anti-proliferative properties acting though the reduction of several growth factors and enhancing microvascular thrombosis, which correlates with lower metastatic potential^[122,161]. The antineoplastic effect of mTOR inhibitors has also been shown in several clinical studies^[162]. There is growing evidence that mTOR deregulation plays a significant role in hepatocellular carcinogenesis, and pre-clinical data indicate that deregulated expression of mTOR pathway effectors is present in 40%-50% of HCCs, and activation of the mTOR pathway is associated with less differentiated neoplasms, earlier tumor recurrence, and worse survival outcomes[163,164]. A recent meta-analysis comparing CNIs against sirolimus demonstrated a protective effect of the latter in terms of achieving a lower incidence of HCC recurrence after LT^[165]. This protective effect was confirmed in a more recent meta-analysis^[166], which demonstrated that sirolimus, compared with CNIs, was associated with lower HCC recurrence (OR = 0.30, 95%CI: 0.16-0.55, P <0.001), lower HCC recurrence-related mortality (OR = 0.29, 95%CI: 0.12-0.70, P = 0.005), and lower overall mortality (OR = 0.35, 95%CI: 0.20-0.61, P < 0.001). In addition, a recent systematic review showed that patients on CNIs developed HCC recurrence significantly more frequently compared with patients on mTORi. In addition, patients on everolimus had significantly lower HCC recurrence rates compared with those on sirolimus or CNIs, although patients treated with mTOR inhibitors tended to have less advanced stages of HCC^[167,168].

CONCLUSION

Overall, the risk of malignancy is two to four times higher in transplant recipients than in an age- and sexmatched population, and cancer is expected to surpass cardiovascular complications as the primary cause of death in transplanted patients within the next 2 decades^[4,169]. De novo malignancy is a very significant cause of mortality, particularly for long-term survivors, and minimization of long-term immunosuppression should be aimed at reducing the incidence of de novo $neoplasms^{[1,170]}$. Promising results in prevention of HCC recurrence have been reported with the use of mTOR inhibitors including everolimus and sirolimus^[154,159,171] and the ongoing open-label prospective randomized controlled SILVER Study[172] will provide more information on whether sirolimus-containing vs mTORinhibitor-free immunosuppression is more efficacious in reducing HCC recurrence. The combined use of sorafenib, a multikinase antiangiogenic inhibitor, and an mTOR inhibitor has yielded positive results in treating patients with HCC recurrence after LT, despite notable associated toxity[173].

REFERENCES

Watt KD, Pedersen RA, Kremers WK, Heimbach JK, Sanchez W, Gores GJ. Long-term probability of and mortality from de novo malignancy after liver transplantation. *Gastroenterology* 2009; 137:



- 2010-2017 [PMID: 19766646 DOI: 10.1053/j.gastro.2009.08.070] **Watt KD**, Pedersen RA, Kremers WK, Heimbach JK, Charlton
- **Watt KD**, Pedersen RA, Kremers WK, Heimbach JK, Charlton MR. Evolution of causes and risk factors for mortality post-liver transplant: results of the NIDDK long-term follow-up study. *Am J Transplant* 2010; **10**: 1420-1427 [PMID: 20486907 DOI: 10.1111/j.1600-6143.2010.03126.x]
- 3 Chandok N, Watt KD. Burden of de novo malignancy in the liver transplant recipient. *Liver Transpl* 2012; 18: 1277-1289 [PMID: 22887956 DOI: 10.1002/lt.23531]
- 4 Sanchez W, Talwalkar JA, Gores GJ. "Will all liver transplantation patients eventually die from cancer?". *J Hepatol* 2006; 44: 13-18 [PMID: 16297490 DOI: 10.1016/j.jhep.2005.10.007]
- Jain AB, Yee LD, Nalesnik MA, Youk A, Marsh G, Reyes J, Zak M, Rakela J, Irish W, Fung JJ. Comparative incidence of de novo nonlymphoid malignancies after liver transplantation under tacrolimus using surveillance epidemiologic end result data. *Transplantation* 1998; 66: 1193-1200 [PMID: 9825817]
- 6 Haagsma EB, Hagens VE, Schaapveld M, van den Berg AP, de Vries EG, Klompmaker IJ, Slooff MJ, Jansen PL. Increased cancer risk after liver transplantation: a population-based study. *J Hepatol* 2001; 34: 84-91 [PMID: 11211912]
- Baccarani U, Adani GL, Serraino D, Lorenzin D, Gambato M, Buda A, Zanus G, Vitale A, Piselli P, De Paoli A, Bresadola V, Risaliti A, Toniutto P, Cillo U, Bresadola F, Burra P. De novo tumors are a major cause of late mortality after orthotopic liver transplantation. *Transplant Proc* 2009; 41: 1303-1305 [PMID: 19460546 DOI: 10.1016/j.transproceed.2009.03.079]
- 8 Verran DJ, Mulhearn MH, Dilworth PJ, Balderson GA, Munn S, Chen JW, Fink MA, Crawford MD, McCaughan GW. Nature and outcomes of the increased incidence of colorectal malignancy after liver transplantation in Australasia. *Med J Aust* 2013; 199: 610-612 [PMID: 24182227]
- 9 Schrem H, Kurok M, Kaltenborn A, Vogel A, Walter U, Zachau L, Manns MP, Klempnauer J, Kleine M. Incidence and long-term risk of de novo malignancies after liver transplantation with implications for prevention and detection. *Liver Transpl* 2013; 19: 1252-1261 [PMID: 24106037 DOI: 10.1002/lt.23722]
- Belloni-Fortina A, Piaserico S, Bordignon M, Gambato M, Senzolo M, Russo FP, Peserico A, De Matteis G, Perissinotto E, Cillo U, Vitale A, Alaibac M, Burra P. Skin cancer and other cutaneous disorders in liver transplant recipients. *Acta Derm Venereol* 2012; 92: 411-415 [PMID: 22377797 DOI: 10.2340/00015555-1316]
- Berg D, Otley CC. Skin cancer in organ transplant recipients: Epidemiology, pathogenesis, and management. *J Am Acad Dermatol* 2002; 47: 1-17; quiz 18-20 [PMID: 12077575]
- 12 Carroll RP, Ramsay HM, Fryer AA, Hawley CM, Nicol DL, Harden PN. Incidence and prediction of nonmelanoma skin cancer post-renal transplantation: a prospective study in Queensland, Australia. Am J Kidney Dis 2003; 41: 676-683 [PMID: 12612993 DOI: 10.1053/ajkd.2003.50130]
- Otley CC, Cherikh WS, Salasche SJ, McBride MA, Christenson LJ, Kauffman HM. Skin cancer in organ transplant recipients: effect of pretransplant end-organ disease. *J Am Acad Dermatol* 2005; 53: 783-790 [PMID: 16243126]
- Mithoefer AB, Supran S, Freeman RB. Risk factors associated with the development of skin cancer after liver transplantation. Liver Transpl 2002; 8: 939-944 [PMID: 12360438 DOI: 10.1053/jlts.2002.35551]
- Saigal S, Norris S, Muiesan P, Rela M, Heaton N, O'Grady J. Evidence of differential risk for posttransplantation malignancy based on pretransplantation cause in patients undergoing liver transplantation. *Liver Transpl* 2002; 8: 482-487 [PMID: 12004349 DOI: 10.1053/jlts.2002.32977]
- Shroff R, Rees L. The post-transplant lymphoproliferative disorder-a literature review. *Pediatr Nephrol* 2004; 19: 369-377 [PMID: 14986084 DOI: 10.1007/s00467-003-1392-x]
- Dierickx D, Tousseyn T, De Wolf-Peeters C, Pirenne J, Verhoef G. Management of posttransplant lymphoproliferative disorders following solid organ transplant: an update. *Leuk Lymphoma* 2011; 52: 950-961 [PMID: 21338285 DOI: 10.3109/10428194.2011.557453]

- 18 Aversa SM, Stragliotto S, Marino D, Calabrese F, Rigotti P, Marchini F, Gambino A, Feltrin G, Boso C, Canova F, Soldà C, Mazzarotto R, Burra P. Post-transplant lymphoproliferative disorders after heart or kidney transplantation at a single centre: presentation and response to treatment. *Acta Haematol* 2008; 120: 36-46 [PMID: 18797163 DOI: 10.1159/000155234]
- Burra P, Buda A, Livi U, Rigotti P, Zanus G, Calabrese F, Caforio A, Menin C, Canova D, Farinati F, Luciana Aversa SM. Occurrence of post-transplant lymphoproliferative disorders among over thousand adult recipients: any role for hepatitis C infection? Eur J Gastroenterol Hepatol 2006; 18: 1065-1070 [PMID: 16957512 DOI: 10.1097/01.meg.0000231752.50587.ae]
- 20 Winkelhorst JT, Brokelman WJ, Tiggeler RG, Wobbes T. Incidence and clinical course of de-novo malignancies in renal allograft recipients. *Eur J Surg Oncol* 2001; 27: 409-413 [PMID: 11417989 DOI: 10.1053/ejso.2001.1119]
- 21 Opelz G, Henderson R. Incidence of non-Hodgkin lymphoma in kidney and heart transplant recipients. *Lancet* 1993; 342: 1514-1516 [PMID: 7902900]
- 22 Libertiny G, Watson CJ, Gray DW, Welsh KI, Morris PJ. Rising incidence of post-transplant lymphoproliferative disease in kidney transplant recipients. *Br J Surg* 2001; 88: 1330-1334 [PMID: 11578286 DOI: 10.1046/j.0007-1323.2001.01924.x]
- 23 Angel LF, Cai TH, Sako EY, Levine SM. Posttransplant lymphoproliferative disorders in lung transplant recipients: clinical experience at a single center. *Ann Transplant* 2000; 5: 26-30 [PMID: 11147026]
- Kasiske BL, Snyder JJ, Gilbertson DT, Wang C. Cancer after kidney transplantation in the United States. Am J Transplant 2004; 4: 905-913 [PMID: 15147424 DOI: 10.1111/j.1600-6143.2004.00450.x]
- Vincenti F, Larsen CP, Alberu J, Bresnahan B, Garcia VD, Kothari J, Lang P, Urrea EM, Massari P, Mondragon-Ramirez G, Reyes-Acevedo R, Rice K, Rostaing L, Steinberg S, Xing J, Agarwal M, Harler MB, Charpentier B. Three-year outcomes from BENEFIT, a randomized, active-controlled, parallel-group study in adult kidney transplant recipients. *Am J Transplant* 2012; 12: 210-217 [PMID: 21992533 DOI: 10.1111/j.1600-6143.2011.03785.x]
- 26 Dantal J, Soulillou JP. Immunosuppressive drugs and the risk of cancer after organ transplantation. N Engl J Med 2005; 352: 1371-1373 [PMID: 15800234 DOI: 10.1056/NEJMe058018]
- 27 Benlloch S, Berenguer M, Prieto M, Moreno R, San Juan F, Rayón M, Mir J, Segura A, Berenguer J. De novo internal neoplasms after liver transplantation: increased risk and aggressive behavior in recent years? Am J Transplant 2004; 4: 596-604 [PMID: 15023152 DOI: 10.1111/j.1600-6143.2004.00380.x]
- Duvoux C, Pageaux GP, Vanlemmens C, Roudot-Thoraval F, Vincens-Rolland AL, Hézode C, Gaulard P, Miguet JP, Larrey D, Dhumeaux D, Cherqui D. Risk factors for lymphoproliferative disorders after liver transplantation in adults: an analysis of 480 patients. *Transplantation* 2002; 74: 1103-1109 [PMID: 12438954 DOI: 10.1097/01.TP.0000031543.16208.1C]
- McLaughlin K, Wajstaub S, Marotta P, Adams P, Grant DR, Wall WJ, Jevnikar AM, Rizkalla KS. Increased risk for posttransplant lymphoproliferative disease in recipients of liver transplants with hepatitis C. Liver Transpl 2000; 6: 570-574 [PMID: 10980055 DOI: 10.1053/jlts.2000.7578]
- 30 Penn I. De novo malignances in pediatric organ transplant recipients. *Pediatr Transplant* 1998; 2: 56-63 [PMID: 10084762]
- 31 Smets F, Sokal EM. Epstein-Barr virus-related lymphoproliferation in children after liver transplant: role of immunity, diagnosis, and management. *Pediatr Transplant* 2002; 6: 280-287 [PMID: 12234267]
- 32 Cacciarelli TV, Reyes J, Jaffe R, Mazariegos GV, Jain A, Fung JJ, Green M. Primary tacrolimus (FK506) therapy and the long-term risk of post-transplant lymphoproliferative disease in pediatric liver transplant recipients. *Pediatr Transplant* 2001; 5: 359-364 [PMID: 11560756]
- Cox KL, Lawrence-Miyasaki LS, Garcia-Kennedy R, Lennette ET, Martinez OM, Krams SM, Berquist WE, So SK, Esquivel CO. An increased incidence of Epstein-Barr virus infection and



- lymphoproliferative disorder in young children on FK506 after liver transplantation. *Transplantation* 1995; **59**: 524-529 [PMID: 7533344]
- 34 Younes BS, McDiarmid SV, Martin MG, Vargas JH, Goss JA, Busuttil RW, Ament ME. The effect of immunosuppression on posttransplant lymphoproliferative disease in pediatric liver transplant patients. *Transplantation* 2000; 70: 94-99 [PMID: 10919581]
- 35 Guthery SL, Heubi JE, Bucuvalas JC, Gross TG, Ryckman FC, Alonso MH, Balistreri WF, Hornung RW. Determination of risk factors for Epstein-Barr virus-associated posttransplant lymphoproliferative disorder in pediatric liver transplant recipients using objective case ascertainment. *Transplantation* 2003; 75: 987-993 [PMID: 12698085 DOI: 10.1097/01.TP.0000057244.03192.BD]
- 36 Fukushima D, Sato K, Kawagishi N, Ohuchi N, Satomi S. Epstein-Barr virus--associated posttransplantation lymphoproliferative disorder with tacrolimus metabolism deterioration in infants after living-donor liver transplantation. *Transplantation* 2015; 99: 114-119 [PMID: 24846306 DOI: 10.1097/TP.000000000000000255]
- 37 Narkewicz MR, Green M, Dunn S, Millis M, McDiarmid S, Mazariegos G, Anand R, Yin W. Decreasing incidence of symptomatic Epstein-Barr virus disease and posttransplant lymphoproliferative disorder in pediatric liver transplant recipients: report of the studies of pediatric liver transplantation experience. Liver Transpl 2013; 19: 730-740 [PMID: 23696264 DOI: 10.1002/lt.23659]
- 38 Khedmat H, Taheri S. Lymphoproliferative disorders in pediatric liver allograft recipients: a review of 212 cases. *Hematol Oncol Stem Cell Ther* 2012; 5: 84-90 [PMID: 22828371]
- 39 Gupta S, Fricker FJ, González-Peralta RP, Slayton WB, Schuler PM, Dharnidharka VR. Post-transplant lymphoproliferative disorder in children: recent outcomes and response to dual rituximab/low-dose chemotherapy combination. *Pediatr Transplant* 2010; 14: 896-902 [PMID: 20642490 DOI: 10.1111/j.1399-3046.2010.01370.x]
- 40 Smets F, Vajro P, Cornu G, Reding R, Otte JB, Sokal E. Indications and results of chemotherapy in children with posttransplant lymphoproliferative disease after liver transplantation. *Transplantation* 2000; 69: 982-984 [PMID: 10755561]
- 41 Euvrard S, Kanitakis J. Skin cancers after liver transplantation: what to do? *J Hepatol* 2006; 44: 27-32 [PMID: 16290909 DOI: 10.1016/j.jhep.2005.10.010]
- 42 Berber I, Altaca G, Aydin C, Dural A, Kara VM, Yigit B, Turkmen A, Titiz MI. Kaposi's sarcoma in renal transplant patients: predisposing factors and prognosis. *Transplant Proc* 2005; 37: 967-968 [PMID: 15848593 DOI: 10.1016/j.transproceed.2004.12.034]
- 43 Serraino D, Angeletti C, Carrieri MP, Longo B, Piche M, Piselli P, Arbustini E, Burra P, Citterio F, Colombo V, Fuzibet JG, Dal Bello B, Targhetta S, Grasso M, Pozzetto U, Bellelli S, Dorrucci M, Dal Maso L, Busnach G, Pradier C, Rezza G. Kaposi's sarcoma in transplant and HIV-infected patients: an epidemiologic study in Italy and France. *Transplantation* 2005; 80: 1699-1704 [PMID: 16378064]
- 44 Piselli P, Busnach G, Citterio F, Frigerio M, Arbustini E, Burra P, Pinna AD, Bresadola V, Ettorre GM, Baccarani U, Buda A, Lauro A, Zanus G, Cimaglia C, Spagnoletti G, Lenardon A, Agozzino M, Gambato M, Zanfi C, Miglioresi L, Di Gioia P, Mei L, Ippolito G, Serraino D. Risk of Kaposi sarcoma after solidorgan transplantation: multicenter study in 4,767 recipients in Italy, 1970-2006. *Transplant Proc* 2009; 41: 1227-1230 [PMID: 19460525 DOI: 10.1016/j.transproceed.2009.03.009]
- 45 Stallone G, Schena A, Infante B, Di Paolo S, Loverre A, Maggio G, Ranieri E, Gesualdo L, Schena FP, Grandaliano G. Sirolimus for Kaposi's sarcoma in renal-transplant recipients. N Engl J Med 2005; 352: 1317-1323 [PMID: 15800227 DOI: 10.1056/NEJMoa042831]
- 46 Jonas S, Rayes N, Neumann U, Neuhaus R, Bechstein WO, Guckelberger O, Tullius SG, Serke S, Neuhaus P. De novo malignancies after liver transplantation using tacrolimus-based protocols or cyclosporine-based quadruple immunosuppression with an interleukin-2 receptor antibody or antithymocyte globulin.

- Cancer 1997; 80: 1141-1150 [PMID: 9305716]
- 47 Sanchez EQ, Marubashi S, Jung G, Levy MF, Goldstein RM, Molmenti EP, Fasola CG, Gonwa TA, Jennings LW, Brooks BK, Klintmalm GB. De novo tumors after liver transplantation: a single-institution experience. *Liver Transpl* 2002; 8: 285-291 [PMID: 11910575 DOI: 10.1053/jlts.2002.29350]
- 48 Frezza EE, Fung JJ, van Thiel DH. Non-lymphoid cancer after liver transplantation. *Hepatogastroenterology* 1997; 44: 1172-1181 [PMID: 9261620]
- 49 Jiménez C, Manrique A, Marqués E, Ortega P, Loinaz C, Gómez R, Meneu JC, Abradelo M, Moreno A, López A, Moreno E. Incidence and risk factors for the development of lung tumors after liver transplantation. *Transpl Int* 2007; 20: 57-63 [PMID: 17181654 DOI: 10.1111/j.1432-2277.2006.00397.x]
- 50 Duvoux C, Delacroix I, Richardet JP, Roudot-Thoraval F, Métreau JM, Fagniez PL, Dhumeaux D, Cherqui D. Increased incidence of oropharyngeal squamous cell carcinomas after liver transplantation for alcoholic cirrhosis. *Transplantation* 1999; 67: 418-421 [PMID: 10030289]
- 51 Tallón Aguilar L, Barrera Pulido L, Bernal Bellido C, Pareja Ciuró F, Suárez Artacho G, Alamo Martínez JM, García González I, Gómez Bravo MA, Bernardos Rodríguez A. Causes and predisposing factors of de novo tumors in our series of liver transplant recipients. *Transplant Proc* 2009; 41: 2453-2454 [PMID: 19715949 DOI: 10.1016/j.transproceed.2009.05.012]
- 52 Jain A, DiMartini A, Kashyap R, Youk A, Rohal S, Fung J. Long-term follow-up after liver transplantation for alcoholic liver disease under tacrolimus. *Transplantation* 2000; 70: 1335-1342 [PMID: 11087149]
- 53 Bellamy CO, DiMartini AM, Ruppert K, Jain A, Dodson F, Torbenson M, Starzl TE, Fung JJ, Demetris AJ. Liver transplantation for alcoholic cirrhosis: long term follow-up and impact of disease recurrence. *Transplantation* 2001; 72: 619-626 [PMID: 11544420]
- 54 Schmilovitz-Weiss H, Mor E, Sulkes J, Bar-Nathan N, Shaharabani E, Melzer E, Tur-Kaspa R, Ben-Ari Z. De novo tumors after liver transplantation: a single-center experience. *Transplant Proc* 2003; 35: 665-666 [PMID: 12644086]
- 55 Castelli E, Hrelia P, Maffei F, Fimognari C, Foschi FG, Caputo F, Cantelli-Forti G, Stefanini GF, Gasbarrini G. Indicators of genetic damage in alcoholics: reversibility after alcohol abstinence. Hepatogastroenterology 1999; 46: 1664-1668 [PMID: 10430317]
- Franceschi S, Talamini R, Barra S, Barón AE, Negri E, Bidoli E, Serraino D, La Vecchia C. Smoking and drinking in relation to cancers of the oral cavity, pharynx, larynx, and esophagus in northern Italy. *Cancer Res* 1990; 50: 6502-6507 [PMID: 2208109]
- 57 Adami J, Gäbel H, Lindelöf B, Ekström K, Rydh B, Glimelius B, Ekbom A, Adami HO, Granath F. Cancer risk following organ transplantation: a nationwide cohort study in Sweden. Br J Cancer 2003; 89: 1221-1227 [PMID: 14520450 DOI: 10.1038/sj.bjc.6601219]
- 58 Penn I. Cancers complicating organ transplantation. N Engl J Med 1990; 323: 1767-1769 [PMID: 2247108 DOI: 10.1056/ NEJM199012203232510]
- 59 Cogliano VJ, Baan R, Straif K, Grosse Y, Lauby-Secretan B, El Ghissassi F, Bouvard V, Benbrahim-Tallaa L, Guha N, Freeman C, Galichet L, Wild CP. Preventable exposures associated with human cancers. *J Natl Cancer Inst* 2011; 103: 1827-1839 [PMID: 22158127 DOI: 10.1093/jnci/djr483]
- 60 Gaglio PJ, Gaglio PJ. Complications in patients with alcoholassociated liver disease who undergo liver transplantation. *Clin Liver Dis* 2012; 16: 865-875 [PMID: 23101987 DOI: 10.1016/ j.cld.2012.08.013]
- 61 Presser SJ, Schumacher G, Neuhaus R, Thuss-Patience P, Stieler J, Neuhaus P. De novo esophageal neoplasia after liver transplantation. *Liver Transpl* 2007; 13: 443-450 [PMID: 17318861 DOI: 10.1002/lt.21058]
- 62 Oezcelik A, Kaiser GM, Dechêne A, Treckmann JW, Sotiropoulos GC, Reinhardt R, Saner FH, Paul A. Progression to adenocarcinoma in Barrett's esophagus after liver transplantation. Transplantation 2011; 91: 1250-1253 [PMID: 21464795 DOI:



- 10.1097/TP.0b013e31821841a0]
- 63 Ilan Y, Shouval D, Galun E, Goldin E, Ligumsky M, Friedman G, Tur Kaspa R. Esophageal malignancy after liver transplantation in a patient with Barrett's esophagus. *Scand J Gastroenterol* 1996; 31: 415-416 [PMID: 8726313]
- 64 Trotter JF, Brazer SR. Rapid progression to high-grade dysplasia in Barrett's esophagus after liver transplantation. *Liver Transpl Surg* 1999; 5: 332-333 [PMID: 10388506 DOI: 10.1002/ lt.500050405]
- 65 Yao F, Ahuja J, Savides T, Behling C, Li S, Hart M. Rapid progression of gastroesophageal junction adenocarcinoma after liver transplantation. *J Clin Gastroenterol* 1997; 24: 54-55 [PMID: 9013353]
- 66 Safadi R, Ilan Y, Eid A, Galun E, Ashur Y, Goldin E, Papo O, Blachar A, Jurim O. Solid tumors after liver transplantation. Transplant Proc 1999; 31: 1894-1895 [PMID: 10371989]
- 67 Na S, Lee GH, Song JH, Ahn JY, Kim SO, Park SJ, Park SE, Kim MY, Lee J, Choi KS, Kim do H, Song HJ, Choi KD, Jung HY, Kim JH. Endoscopic resection of gastric neoplasm in solid-organ transplant recipients. *Transplantation* 2014; 97: 781-787 [PMID: 24406452 DOI: 10.1097/01.TP.0000438638.29214.f4]
- 68 Buell JF, Husted T, Hanaway MJ, Peddi VR, Trofe J, Gross TG, Beebe TM, First MR, Woodle ES. Incidental diagnosis of gastric cancer in transplant recipients improves patient survival. *Surgery* 2002; 132: 754-758; discussion 758-760 [PMID: 12407362]
- 69 Penn I. Posttransplantation de novo tumors in liver allograft recipients. *Liver Transpl Surg* 1996; 2: 52-59 [PMID: 9346628]
- 70 Vera A, Gunson BK, Ussatoff V, Nightingale P, Candinas D, Radley S, Mayer A, Buckels JA, McMaster P, Neuberger J, Mirza DF. Colorectal cancer in patients with inflammatory bowel disease after liver transplantation for primary sclerosing cholangitis. Transplantation 2003; 75: 1983-1988 [PMID: 12829898 DOI: 10.1097/01.TP.0000058744.34965.38]
- 71 Bleday R, Lee E, Jessurun J, Heine J, Wong WD. Increased risk of early colorectal neoplasms after hepatic transplant in patients with inflammatory bowel disease. *Dis Colon Rectum* 1993; 36: 908-912 [PMID: 8404380]
- 72 **Fabia R**, Levy MF, Testa G, Obiekwe S, Goldstein RM, Husberg BS, Gonwa TA, Klintmalm GB. Colon carcinoma in patients undergoing liver transplantation. *Am J Surg* 1998; **176**: 265-269 [PMID: 9776156]
- 73 Buell JF, Papaconstantinou HT, Skalow B, Hanaway MJ, Alloway RR, Woodle ES. De novo colorectal cancer: five-year survival is markedly lower in transplant recipients compared with the general population. *Transplant Proc* 2005; 37: 960-961 [PMID: 15848590 DOI: 10.1016/j.transproceed.2004.12.122]
- 74 Johnson EE, Leverson GE, Pirsch JD, Heise CP. A 30-year analysis of colorectal adenocarcinoma in transplant recipients and proposal for altered screening. *J Gastrointest Surg* 2007; 11: 272-279 [PMID: 17458597 DOI: 10.1007/s11605-007-0084-4]
- 75 Trevisani F, Garuti F, Cucchetti A, Lenzi B, Bernardi M. De novo hepatocellular carcinoma of liver allograft: a neglected issue. *Cancer Lett* 2015; 357: 47-54 [PMID: 25444925 DOI: 10.1016/ j.canlet.2014.11.032]
- 76 Adam R, Karam V, Delvart V, O'Grady J, Mirza D, Klempnauer J, Castaing D, Neuhaus P, Jamieson N, Salizzoni M, Pollard S, Lerut J, Paul A, Garcia-Valdecasas JC, Rodríguez FS, Burroughs A. Evolution of indications and results of liver transplantation in Europe. A report from the European Liver Transplant Registry (ELTR). J Hepatol 2012; 57: 675-688 [PMID: 22609307 DOI: 10.1016/j.jhep.2012.04.015]
- 77 Angelico M, Cillo U, Fagiuoli S, Gasbarrini A, Gavrila C, Marianelli T, Costa AN, Nardi A, Strazzabosco M, Burra P, Agnes S, Baccarani U, Calise F, Colledan M, Cuomo O, De Carlis L, Donataccio M, Ettorre GM, Gerunda GE, Gridelli B, Lupo L, Mazzaferro V, Pinna A, Risaliti A, Salizzoni M, Tisone G, Valente U, Rossi G, Rossi M, Zamboni F. Liver Match, a prospective observational cohort study on liver transplantation in Italy: study design and current practice of donor-recipient matching. Dig Liver Dis 2011; 43: 155-164 [PMID: 21185796 DOI: 10.1016/

- j.dld.2010.11.002]
- Flemming P, Tillmann HL, Barg-Hock H, Kleeberger W, Manns MP, Klempnauer J, Kreipe HH. Donor origin of de novo hepatocellular carcinoma in hepatic allografts. *Transplantation* 2003; 76: 1625-1627 [PMID: 14702536 DOI: 10.1097/01. TP.0000086341.57778.D9]
- 79 Levitsky J, Faust TW, Cohen SM, Te HS. Group G streptococcal bacteremia and de novo hepatocellular carcinoma after liver transplantation. *Liver Transpl* 2002; 8: 572 [PMID: 12037793 DOI: 10.1002/lt.500080614]
- 80 Croitoru A, Schiano TD, Schwartz M, Roayaie S, Xu R, Suriawinata A, Fiel MI. De novo hepatocellular carcinoma occurring in a transplanted liver: case report and review of the literature. *Dig Dis Sci* 2006; 51: 1780-1782 [PMID: 16967310 DOI: 10.1007/s10620-006-9333-8]
- 81 Sotiropoulos GC, Frilling A, Molmenti EP, Brokalaki EI, Beckebaum S, Omar OS, Broelsch CE, Malagó M. De novo hepatocellular carcinoma in recurrent liver cirrhosis after liver transplantation for benign hepatic disease: is a deceased donor retransplantation justified? *Transplantation* 2006; 82: 1112 [PMID: 17060864 DOI: 10.1097/01.tp.0000230283.84633.4a]
- Kita Y, Klintmalm G, Kobayashi S, Yanaga K. Retransplantation for de novo hepatocellular carcinoma in a liver allograft with recurrent hepatitis B cirrhosis 14 years after primary liver transplantation. *Dig Dis Sci* 2007; 52: 3392-3393 [PMID: 17404871 DOI: 10.1007/s10620-006-9574-6]
- 83 Teng MW, Swann JB, Koebel CM, Schreiber RD, Smyth MJ. Immune-mediated dormancy: an equilibrium with cancer. J Leukoc Biol 2008; 84: 988-993 [PMID: 18515327 DOI: 10.1189/ jlb.1107774]
- 84 Strauss DC, Thomas JM. Transmission of donor melanoma by organ transplantation. *Lancet Oncol* 2010; 11: 790-796 [PMID: 20451456 DOI: 10.1016/S1470-2045(10)70024-3]
- 85 Ison MG, Nalesnik MA. An update on donor-derived disease transmission in organ transplantation. Am J Transplant 2011; 11: 1123-1130 [PMID: 21443676 DOI: 10.1111/j.1600-6143.2011.03493.x]
- Kauffman HM, McBride MA, Delmonico FL. First report of the United Network for Organ Sharing Transplant Tumor Registry: donors with a history of cancer. *Transplantation* 2000; 70: 1747-1751 [PMID: 11152107]
- Kauffman HM, Cherikh WS, McBride MA, Cheng Y, Hanto DW. Deceased donors with a past history of malignancy: an organ procurement and transplantation network/united network for organ sharing update. *Transplantation* 2007; 84: 272-274 [PMID: 17667822 DOI: 10.1097/01.tp.0000267919.93425.fb]
- Ison MG, Hager J, Blumberg E, Burdick J, Carney K, Cutler J, Dimaio JM, Hasz R, Kuehnert MJ, Ortiz-Rios E, Teperman L, Nalesnik M. Donor-derived disease transmission events in the United States: data reviewed by the OPTN/UNOS Disease Transmission Advisory Committee. *Am J Transplant* 2009; 9: 1929-1935 [PMID: 19538493 DOI: 10.1111/j.1600-6143.2009.02700.x]
- 89 Birkeland SA, Storm HH. Risk for tumor and other disease transmission by transplantation: a population-based study of unrecognized malignancies and other diseases in organ donors. *Transplantation* 2002; 74: 1409-1413 [PMID: 12451241 DOI: 10.1097/01.TP.0000034717.19606.B5]
- 90 Feng S, Buell JF, Chari RS, DiMaio JM, Hanto DW. Tumors and transplantation: The 2003 Third Annual ASTS State-of-the-Art Winter Symposium. Am J Transplant 2003; 3: 1481-1487 [PMID: 14629278]
- 91 Myron Kauffman H, McBride MA, Cherikh WS, Spain PC, Marks WH, Roza AM. Transplant tumor registry: donor related malignancies. *Transplantation* 2002; 74: 358-362 [PMID: 12177614]
- 92 Desai R, Neuberger J. Donor transmitted and de novo cancer after liver transplantation. World J Gastroenterol 2014; 20: 6170-6179 [PMID: 24876738 DOI: 10.3748/wjg.v20.i20.6170]
- 93 Buell JF, Trofe J, Sethuraman G, Hanaway MJ, Beebe TM, Gross TG, Alloway R, First MR, Woodle ES. Donors with central nervous system malignancies: are they truly safe? *Transplantation*



8764

- 2003; **76**: 340-343 [PMID: 12883189 DOI: 10.1097/01. TP.0000076094.64973.D8]
- 94 Watson CJ, Roberts R, Wright KA, Greenberg DC, Rous BA, Brown CH, Counter C, Collett D, Bradley JA. How safe is it to transplant organs from deceased donors with primary intracranial malignancy? An analysis of UK Registry data. Am J Transplant 2010; 10: 1437-1444 [PMID: 20486904 DOI: 10.1111/j.1600-6143.2010.03130.x]
- 95 Abramowicz D, Cochat P, Claas FH, Heemann U, Pascual J, Dudley C, Harden P, Hourmant M, Maggiore U, Salvadori M, Spasovski G, Squifflet JP, Steiger J, Torres A, Viklicky O, Zeier M, Vanholder R, Van Biesen W, Nagler E. European Renal Best Practice Guideline on kidney donor and recipient evaluation and perioperative care. Nephrol Dial Transplant 2014; Epub ahead of print [PMID: 25007790 DOI: 10.1093/ndt/gfu216]
- 96 Buell JF, Beebe TM, Trofe J, Gross TG, Alloway RR, Hanaway MJ, Woodle ES. Donor transmitted malignancies. *Ann Transplant* 2004; 9: 53-56 [PMID: 15478892]
- 97 Gutierrez-Dalmau A, Campistol JM. Immunosuppressive therapy and malignancy in organ transplant recipients: a systematic review. *Drugs* 2007; 67: 1167-1198 [PMID: 17521218]
- 98 Grulich AE, van Leeuwen MT, Falster MO, Vajdic CM. Incidence of cancers in people with HIV/AIDS compared with immunosuppressed transplant recipients: a meta-analysis. *Lancet* 2007; 370: 59-67 [PMID: 17617273 DOI: 10.1016/S0140-6736(07)61050-2]
- 99 Soulillou JP, Giral M. Controlling the incidence of infection and malignancy by modifying immunosuppression. *Transplantation* 2001; 72: S89-S93 [PMID: 11833147]
- 100 Dantal J, Hourmant M, Cantarovich D, Giral M, Blancho G, Dreno B, Soulillou JP. Effect of long-term immunosuppression in kidney-graft recipients on cancer incidence: randomised comparison of two cyclosporin regimens. *Lancet* 1998; 351: 623-628 [PMID: 9500317 DOI: 10.1016/S0140-6736(97)08496-1]
- 101 Tanner JE, Alfieri C. The Epstein-Barr virus and post-transplant lymphoproliferative disease: interplay of immunosuppression, EBV, and the immune system in disease pathogenesis. *Transpl Infect Dis* 2001; 3: 60-69 [PMID: 11395971]
- 102 Hojo M, Morimoto T, Maluccio M, Asano T, Morimoto K, Lagman M, Shimbo T, Suthanthiran M. Cyclosporine induces cancer progression by a cell-autonomous mechanism. *Nature* 1999; 397: 530-534 [PMID: 10028970 DOI: 10.1038/17401]
- 103 Yarosh DB, Pena AV, Nay SL, Canning MT, Brown DA. Calcineurin inhibitors decrease DNA repair and apoptosis in human keratinocytes following ultraviolet B irradiation. *J Invest Dermatol* 2005; 125: 1020-1025 [PMID: 16297204 DOI: 10.1111/j.0022-202X.2005.23858.x]
- 104 Herman M, Weinstein T, Korzets A, Chagnac A, Ori Y, Zevin D, Malachi T, Gafter U. Effect of cyclosporin A on DNA repair and cancer incidence in kidney transplant recipients. *J Lab Clin Med* 2001; 137: 14-20 [PMID: 11150019 DOI: 10.1067/mlc.2001.111469]
- 105 Guba M, von Breitenbuch P, Steinbauer M, Koehl G, Flegel S, Hornung M, Bruns CJ, Zuelke C, Farkas S, Anthuber M, Jauch KW, Geissler EK. Rapamycin inhibits primary and metastatic tumor growth by antiangiogenesis: involvement of vascular endothelial growth factor. *Nat Med* 2002; 8: 128-135 [PMID: 11821896 DOI: 10.1038/nm0202-128]
- 106 Maluccio M, Sharma V, Lagman M, Vyas S, Yang H, Li B, Suthanthiran M. Tacrolimus enhances transforming growth factor-betal expression and promotes tumor progression. *Transplantation* 2003; 76: 597-602 [PMID: 12923450 DOI: 10.1097/01. TP.0000081399.75231.3B]
- 107 Suthanthiran M, Hojo M, Maluccio M, Boffa DJ, Luan FL. Post-transplantation malignancy: a cell autonomous mechanism with implications for therapy. *Trans Am Clin Climatol Assoc* 2009; 120: 369-388 [PMID: 19768190]
- 108 Walsh SB, Xu J, Xu H, Kurundkar AR, Maheshwari A, Grizzle WE, Timares L, Huang CC, Kopelovich L, Elmets CA, Athar M. Cyclosporine a mediates pathogenesis of aggressive cutaneous squamous cell carcinoma by augmenting epithelial-mesenchymal

8765

- transition: role of TGFβ signaling pathway. *Mol Carcinog* 2011; **50**: 516-527 [PMID: 21308804 DOI: 10.1002/mc.20744]
- Tjon AS, Sint Nicolaas J, Kwekkeboom J, de Man RA, Kazemier G, Tilanus HW, Hansen BE, van der Laan LJ, Tha-In T, Metselaar HJ. Increased incidence of early de novo cancer in liver graft recipients treated with cyclosporine: an association with C2 monitoring and recipient age. *Liver Transpl* 2010; 16: 837-846 [PMID: 20583092 DOI: 10.1002/lt.22064]
- 110 Rodríguez-Perálvarez M, Germani G, Papastergiou V, Tsochatzis E, Thalassinos E, Luong TV, Rolando N, Dhillon AP, Patch D, O' Beirne J, Thorburn D, Burroughs AK. Early tacrolimus exposure after liver transplantation: relationship with moderate/severe acute rejection and long-term outcome. *J Hepatol* 2013; 58: 262-270 [PMID: 23023010 DOI: 10.1016/j.jhep.2012.09.019]
- 111 Øzbay LA, Smidt K, Mortensen DM, Carstens J, Jørgensen KA, Rungby J. Cyclosporin and tacrolimus impair insulin secretion and transcriptional regulation in INS-1E beta-cells. *Br J Pharmacol* 2011; 162: 136-146 [PMID: 20825407 DOI: 10.1111/j.1476-5381.2010.01018.x]
- 112 Chakkera HA, Mandarino LJ. Calcineurin inhibition and new-onset diabetes mellitus after transplantation. *Transplantation* 2013; 95: 647-652 [PMID: 23076551 DOI: 10.1097/TP.0b013e31826e592e]
- 113 Soleimanpour SA, Crutchlow MF, Ferrari AM, Raum JC, Groff DN, Rankin MM, Liu C, De León DD, Naji A, Kushner JA, Stoffers DA. Calcineurin signaling regulates human islet {beta}-cell survival. *J Biol Chem* 2010; 285: 40050-40059 [PMID: 20943662 DOI: 10.1074/jbc.M110.154955]
- 114 Navasa M, Bustamante J, Marroni C, González E, Andreu H, Esmatjes E, García-Valdecasas JC, Grande L, Cirera I, Rimola A, Rodés J. Diabetes mellitus after liver transplantation: prevalence and predictive factors. *J Hepatol* 1996; 25: 64-71 [PMID: 8836903]
- 115 Trail KC, McCashland TM, Larsen JL, Heffron TG, Stratta RJ, Langnas AN, Fox IJ, Zetterman RK, Donovan JP, Sorrell MF, Pillen TJ, Ruby EI, Shaw BW. Morbidity in patients with posttransplant diabetes mellitus following orthotopic liver transplantation. *Liver Transpl Surg* 1996; 2: 276-283 [PMID: 9346661]
- 116 Levy G, Villamil F, Samuel D, Sanjuan F, Grazi GL, Wu Y, Marotta P, Boillot O, Muehlbacher F, Klintmalm G. Results of lis2t, a multicenter, randomized study comparing cyclosporine microemulsion with C2 monitoring and tacrolimus with C0 monitoring in de novo liver transplantation. *Transplantation* 2004; 77: 1632-1638 [PMID: 15201658]
- 117 El-Serag HB, Hampel H, Javadi F. The association between diabetes and hepatocellular carcinoma: a systematic review of epidemiologic evidence. *Clin Gastroenterol Hepatol* 2006; 4: 369-380 [PMID: 16527702 DOI: 10.1016/j.cgh.2005.12.007]
- 118 **Penn I**. The changing pattern of posttransplant malignancies. *Transplant Proc* 1991; **23**: 1101-1103 [PMID: 1899153]
- Howard TK, Klintmalm GB, Stone MJ, Cofer JB, Husberg BS, Goldstein RM, Gonwa TA. Lymphoproliferative disorder masquerading as rejection in liver transplant recipients--an early aggressive tumor with atypical presentation. *Transplantation* 1992; 53: 1145-1147 [PMID: 1316652]
- 120 Swinnen LJ, Costanzo-Nordin MR, Fisher SG, O'Sullivan EJ, Johnson MR, Heroux AL, Dizikes GJ, Pifarre R, Fisher RI. Increased incidence of lymphoproliferative disorder after immunosuppression with the monoclonal antibody OKT3 in cardiac-transplant recipients. N Engl J Med 1990; 323: 1723-1728 [PMID: 2100991 DOI: 10.1056/NEJM199012203232502]
- 121 Penninga L, Wettergren A, Wilson CH, Chan AW, Steinbrüchel DA, Gluud C. Antibody induction versus placebo, no induction, or another type of antibody induction for liver transplant recipients. *Cochrane Database Syst Rev* 2014; 6: CD010253 [PMID: 24901467 DOI: 10.1002/14651858.CD010253.pub2]
- 122 Neuhaus P, Klupp J, Langrehr JM. mTOR inhibitors: an overview. Liver Transpl 2001; 7: 473-484 [PMID: 11443573 DOI: 10.1053/jlts.2001.24645]
- 23 Mathew T, Kreis H, Friend P. Two-year incidence of malignancy in sirolimus-treated renal transplant recipients: results from five



- multicenter studies. *Clin Transplant* 2004; **18**: 446-449 [PMID: 15233824 DOI: 10.1111/j.1399-0012.2004.00188.x]
- 124 Klintmalm GB, Saab S, Hong JC, Nashan B. The role of mammalian target of rapamycin inhibitors in the management of post-transplant malignancy. *Clin Transplant* 2014; 28: 635-648 [PMID: 24628264 DOI: 10.1111/ctr.12357]
- 125 Ewald F, Grabinski N, Grottke A, Windhorst S, Nörz D, Carstensen L, Staufer K, Hofmann BT, Diehl F, David K, Schumacher U, Nashan B, Jücker M. Combined targeting of AKT and mTOR using MK-2206 and RAD001 is synergistic in the treatment of cholangiocarcinoma. *Int J Cancer* 2013; 133: 2065-2076 [PMID: 23588885 DOI: 10.1002/ijc.28214]
- 126 Grabinski N, Ewald F, Hofmann BT, Staufer K, Schumacher U, Nashan B, Jücker M. Combined targeting of AKT and mTOR synergistically inhibits proliferation of hepatocellular carcinoma cells. *Mol Cancer* 2012; 11: 85 [PMID: 23167739 DOI: 10.1186/1476-4598-11-85]
- 127 **Rodríguez-Perálvarez M**, De la Mata M, Burroughs AK. Liver transplantation: immunosuppression and oncology. *Curr Opin Organ Transplant* 2014; **19**: 253-260 [PMID: 24685671 DOI: 10.1097/MOT.0000000000000009]
- 128 Finkenstedt A, Graziadei IW, Oberaigner W, Hilbe W, Nachbaur K, Mark W, Margreiter R, Vogel W. Extensive surveillance promotes early diagnosis and improved survival of de novo malignancies in liver transplant recipients. *Am J Transplant* 2009; 9: 2355-2361 [PMID: 19663894 DOI: 10.1111/j.1600-6143.2009.02766.x]
- 129 Herrero JI, Alegre F, Quiroga J, Pardo F, Iñarrairaegui M, Sangro B, Rotellar F, Montiel C, Prieto J. Usefulness of a program of neoplasia surveillance in liver transplantation. A preliminary report. *Clin Transplant* 2009; 23: 532-536 [PMID: 19681977 DOI: 10.1111/j.1399-0012.2008.00927.x]
- 130 Chak E, Saab S. Risk factors and incidence of de novo malignancy in liver transplant recipients: a systematic review. *Liver Int* 2010; 30: 1247-1258 [PMID: 20602682 DOI: 10.1111/j.1478-3231.2010.02303.x]
- 131 U.S. Preventive Services Task Force1. Screening for skin cancer: U.S. Preventive Services Task Force recommendation statement. Ann Intern Med 2009; 150: 188-193 [PMID: 19189908]
- 132 Kasiske BL, Vazquez MA, Harmon WE, Brown RS, Danovitch GM, Gaston RS, Roth D, Scandling JD, Singer GG. Recommendations for the outpatient surveillance of renal transplant recipients. American Society of Transplantation. J Am Soc Nephrol 2000; 11 Suppl 15: S1-86 [PMID: 11044969]
- 133 Mandelblatt JS, Cronin KA, Bailey S, Berry DA, de Koning HJ, Draisma G, Huang H, Lee SJ, Munsell M, Plevritis SK, Ravdin P, Schechter CB, Sigal B, Stoto MA, Stout NK, van Ravesteyn NT, Venier J, Zelen M, Feuer EJ. Effects of mammography screening under different screening schedules: model estimates of potential benefits and harms. *Ann Intern Med* 2009; 151: 738-747 [PMID: 19920274 DOI: 10.7326/0003-4819-151-10-200911170-00010]
- 134 U.S. Preventive Services Task Force. Screening for colorectal cancer: U.S. Preventive Services Task Force recommendation statement. *Ann Intern Med* 2008; 149: 627-637 [PMID: 18838716]
- 135 Aberle DR, Adams AM, Berg CD, Black WC, Clapp JD, Fagerstrom RM, Gareen IF, Gatsonis C, Marcus PM, Sicks JD. Reduced lung-cancer mortality with low-dose computed tomographic screening. N Engl J Med 2011; 365: 395-409 [PMID: 21714641 DOI: 10.1056/NEJMoa1102873]
- 136 Geissler EK, Schlitt HJ, Thomas G. mTOR, cancer and transplantation. Am J Transplant 2008; 8: 2212-2218 [PMID: 18785960 DOI: 10.1111/j.1600-6143.2008.02391.x]
- 137 Koehl GE, Andrassy J, Guba M, Richter S, Kroemer A, Scherer MN, Steinbauer M, Graeb C, Schlitt HJ, Jauch KW, Geissler EK. Rapamycin protects allografts from rejection while simultaneously attacking tumors in immunosuppressed mice. *Transplantation* 2004; 77: 1319-1326 [PMID: 15167584]
- 138 Luan FL, Hojo M, Maluccio M, Yamaji K, Suthanthiran M. Rapamycin blocks tumor progression: unlinking immunosuppression from antitumor efficacy. *Transplantation* 2002; 73: 1565-1572 [PMID: 12042641]

- 139 Kahan BD, Camardo JS. Rapamycin: clinical results and future opportunities. *Transplantation* 2001; 72: 1181-1193 [PMID: 11602840]
- 140 Campistol JM, Eris J, Oberbauer R, Friend P, Hutchison B, Morales JM, Claesson K, Stallone G, Russ G, Rostaing L, Kreis H, Burke JT, Brault Y, Scarola JA, Neylan JF. Sirolimus therapy after early cyclosporine withdrawal reduces the risk for cancer in adult renal transplantation. *J Am Soc Nephrol* 2006; 17: 581-589 [PMID: 16434506 DOI: 10.1681/ASN.2005090993]
- 141 Schena FP, Pascoe MD, Alberu J, del Carmen Rial M, Oberbauer R, Brennan DC, Campistol JM, Racusen L, Polinsky MS, Goldberg-Alberts R, Li H, Scarola J, Neylan JF. Conversion from calcineurin inhibitors to sirolimus maintenance therapy in renal allograft recipients: 24-month efficacy and safety results from the CONVERT trial. *Transplantation* 2009; 87: 233-242 [PMID: 19155978 DOI: 10.1097/TP.0b013e3181927a41]
- 142 Kauffman HM, Cherikh WS, Cheng Y, Hanto DW, Kahan BD. Maintenance immunosuppression with target-of-rapamycin inhibitors is associated with a reduced incidence of de novo malignancies. *Transplantation* 2005; 80: 883-889 [PMID: 16249734]
- 143 Kahan BD, Yakupoglu YK, Schoenberg L, Knight RJ, Katz SM, Lai D, Van Buren CT. Low incidence of malignancy among sirolimus/cyclosporine-treated renal transplant recipients. Transplantation 2005; 80: 749-758 [PMID: 16210961]
- 144 Alberú J, Pascoe MD, Campistol JM, Schena FP, Rial Mdel C, Polinsky M, Neylan JF, Korth-Bradley J, Goldberg-Alberts R, Maller ES. Lower malignancy rates in renal allograft recipients converted to sirolimus-based, calcineurin inhibitor-free immunotherapy: 24-month results from the CONVERT trial. *Transplantation* 2011; 92: 303-310 [PMID: 21792049 DOI: 10.1097/TP.0b013e3182247ae2]
- 145 Zuckermann A, Manito N, Epailly E, Fiane A, Bara C, Delgado JF, Lehmkuhl H, Ross H, Eisen H, Chapman J, Valantine H. Multidisciplinary insights on clinical guidance for the use of proliferation signal inhibitors in heart transplantation. *J Heart Lung Transplant* 2008; 27: 141-149 [PMID: 18267219 DOI: 10.1016/j.healun.2007.08.014]
- 146 Gomez-Camarero J, Salcedo M, Rincon D, Lo Iacono O, Ripoll C, Hernando A, Sanz C, Clemente G, Bañares R. Use of everolimus as a rescue immunosuppressive therapy in liver transplant patients with neoplasms. *Transplantation* 2007; 84: 786-791 [PMID: 17893613 DOI: 10.1097/01.tp.0000280549.93403.dd]
- 147 Thimonier E, Guillaud O, Walter T, Decullier E, Vallin M, Boillot O, Dumortier J. Conversion to everolimus dramatically improves the prognosis of de novo malignancies after liver transplantation for alcoholic liver disease. *Clin Transplant* 2014; 28: 1339-1348 [PMID: 25081431 DOI: 10.1111/ctr.12430]
- 148 Benten D, Sterneck M, Panse J, Rogiers X, Lohse AW. Low recurrence of preexisting extrahepatic malignancies after liver transplantation. *Liver Transpl* 2008; 14: 789-798 [PMID: 18412260 DOI: 10.1002/lt.21434]
- 149 EBPG Expert Group on Renal Transplantation. European best practice guidelines for renal transplantation. Section IV: Long-term management of the transplant recipient. IV.10. Pregnancy in renal transplant recipients. Nephrol Dial Transplant 2002; 17 Suppl 4: 50-55 [PMID: 12091650]
- Mazzaferro V, Regalia E, Doci R, Andreola S, Pulvirenti A, Bozzetti F, Montalto F, Ammatuna M, Morabito A, Gennari L. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. N Engl J Med 1996; 334: 693-699 [PMID: 8594428 DOI: 10.1056/NEJM199603143341104]
- 151 Kasiske BL, Cangro CB, Hariharan S, Hricik DE, Kerman RH, Roth D, Rush DN, Vazquez MA, Weir MR. The evaluation of renal transplantation candidates: clinical practice guidelines. Am J Transplant 2001; 1 Suppl 2: 3-95 [PMID: 12108435]
- 152 Penn I. The effect of immunosuppression on pre-existing cancers. Transplantation 1993; 55: 742-747 [PMID: 8475546]
- 153 Castroagudín JF, Molina E, Bustamante M, Tomé S, Otero E, Martínez J, Segade FR, Conde R, Varo E. Orthotopic liver transplantation for hepatocellular carcinoma: a thirteen-year single-



- center experience. *Transplant Proc* 2008; **40**: 2975-2977 [PMID: 19010164 DOI: 10.1016/j.transproceed.2008.09.006]
- 154 Chinnakotla S, Davis GL, Vasani S, Kim P, Tomiyama K, Sanchez E, Onaca N, Goldstein R, Levy M, Klintmalm GB. Impact of sirolimus on the recurrence of hepatocellular carcinoma after liver transplantation. *Liver Transpl* 2009; 15: 1834-1842 [PMID: 19938137 DOI: 10.1002/lt.21953]
- 155 DuBay D, Sandroussi C, Sandhu L, Cleary S, Guba M, Cattral MS, McGilvray I, Ghanekar A, Selzner M, Greig PD, Grant DR. Liver transplantation for advanced hepatocellular carcinoma using poor tumor differentiation on biopsy as an exclusion criterion. *Ann Surg* 2011; 253: 166-172 [PMID: 21294289]
- 156 Vivarelli M, Cucchetti A, La Barba G, Ravaioli M, Del Gaudio M, Lauro A, Grazi GL, Pinna AD. Liver transplantation for hepatocellular carcinoma under calcineurin inhibitors: reassessment of risk factors for tumor recurrence. *Ann Surg* 2008; 248: 857-862 [PMID: 18948815 DOI: 10.1097/SLA.0b013e3181896278]
- 157 Sotiropoulos GC, Molmenti EP, Lösch C, Beckebaum S, Broelsch CE, Lang H. Meta-analysis of tumor recurrence after liver transplantation for hepatocellular carcinoma based on 1,198 cases. Eur J Med Res 2007; 12: 527-534 [PMID: 18024261]
- 158 Kornberg A, Küpper B, Tannapfel A, Katenkamp K, Thrum K, Habrecht O, Wilberg J. Long-term survival after recurrent hepatocellular carcinoma in liver transplant patients: clinical patterns and outcome variables. Eur J Surg Oncol 2010; 36: 275-280 [PMID: 19857941 DOI: 10.1016/j.ejso.2009.10.001]
- 159 Vivarelli M, Dazzi A, Zanello M, Cucchetti A, Cescon M, Ravaioli M, Del Gaudio M, Lauro A, Grazi GL, Pinna AD. Effect of different immunosuppressive schedules on recurrence-free survival after liver transplantation for hepatocellular carcinoma. *Transplantation* 2010; 89: 227-231 [PMID: 20098287 DOI: 10.1097/TP.0b013e3181c3c540]
- 160 Rodríguez-Perálvarez M, Tsochatzis E, Naveas MC, Pieri G, García-Caparrós C, O'Beirne J, Poyato-González A, Ferrín-Sánchez G, Montero-Álvarez JL, Patch D, Thorburn D, Briceño J, De la Mata M, Burroughs AK. Reduced exposure to calcineurin inhibitors early after liver transplantation prevents recurrence of hepatocellular carcinoma. *J Hepatol* 2013; 59: 1193-1199 [PMID: 23867318 DOI: 10.1016/j.jhep.2013.07.012]
- 161 Chapman TM, Perry CM. Everolimus. *Drugs* 2004; 64: 861-872; discussion 873-874 [PMID: 15059040]
- 162 Toso C, Merani S, Bigam DL, Shapiro AM, Kneteman NM. Sirolimus-based immunosuppression is associated with increased survival after liver transplantation for hepatocellular carcinoma. *Hepatology* 2010; 51: 1237-1243 [PMID: 20187107 DOI: 10.1002/hep.23437]
- 163 Ashworth RE, Wu J. Mammalian target of rapamycin inhibition in hepatocellular carcinoma. World J Hepatol 2014; 6: 776-782 [PMID: 25429315 DOI: 10.4254/wjh.v6.i11.776]
- 164 Zhou L, Huang Y, Li J, Wang Z. The mTOR pathway is associated with the poor prognosis of human hepatocellular carcinoma. *Med Oncol* 2010; 27: 255-261 [PMID: 19301157 DOI: 10.1007/ s12032-009-9201-4]
- 165 Liang W, Wang D, Ling X, Kao AA, Kong Y, Shang Y, Guo Z, He X. Sirolimus-based immunosuppression in liver transplantation for hepatocellular carcinoma: a meta-analysis. *Liver Transpl* 2012; 18: 62-69 [PMID: 21964956 DOI: 10.1002/lt.22441]
- 166 Menon KV, Hakeem AR, Heaton ND. Meta-analysis: recurrence and survival following the use of sirolimus in liver transplantation for hepatocellular carcinoma. *Aliment Pharmacol Ther* 2013; 37: 411-419 [PMID: 23278125 DOI: 10.1111/apt.12185]
- 167 Cholongitas E, Mamou C, Rodríguez-Castro KI, Burra P. Mammalian target of rapamycin inhibitors are associated with lower rates of hepatocellular carcinoma recurrence after liver transplantation: a systematic review. *Transpl Int* 2014; 27: 1039-1049 [PMID: 24943720 DOI: 10.1111/tri.12372]
- 168 Cholongitas E, Burra P. Reply to: Time to resize the role of everolimus as treatment of hepatocellular carcinoma recurrence after liver transplant. *Transpl Int* 2015; 28: 503-504 [PMID: 25440876 DOI: 10.1111/tri.12496]

- 169 Pruthi J, Medkiff KA, Esrason KT, Donovan JA, Yoshida EM, Erb SR, Steinbrecher UP, Fong TL. Analysis of causes of death in liver transplant recipients who survived more than 3 years. Liver Transpl 2001; 7: 811-815 [PMID: 11552217 DOI: 10.1053/jlts.2001.27084]
- 170 Schoening WN, Buescher N, Rademacher S, Andreou A, Kuehn S, Neuhaus R, Guckelberger O, Puhl G, Seehofer D, Neuhaus P. Twenty-year longitudinal follow-up after orthotopic liver transplantation: a single-center experience of 313 consecutive cases. *Am J Transplant* 2013; 13: 2384-2394 [PMID: 23915357 DOI: 10.1111/ajt.12384]
- 171 Zhou J, Wang Z, Wu ZQ, Qiu SJ, Yu Y, Huang XW, Tang ZY, Fan J. Sirolimus-based immunosuppression therapy in liver transplantation for patients with hepatocellular carcinoma exceeding the Milan criteria. *Transplant Proc* 2008; 40: 3548-3553 [PMID: 19100435 DOI: 10.1016/j.transproceed.2008.03.165]
- 172 Schnitzbauer AA, Zuelke C, Graeb C, Rochon J, Bilbao I, Burra P, de Jong KP, Duvoux C, Kneteman NM, Adam R, Bechstein WO, Becker T, Beckebaum S, Chazouillères O, Cillo U, Colledan M, Fändrich F, Gugenheim J, Hauss JP, Heise M, Hidalgo E, Jamieson N, Königsrainer A, Lamby PE, Lerut JP, Mäkisalo H, Margreiter R, Mazzaferro V, Mutzbauer I, Otto G, Pageaux GP, Pinna AD, Pirenne J, Rizell M, Rossi G, Rostaing L, Roy A, Turrion VS, Schmidt J, Troisi RI, van Hoek B, Valente U, Wolf P, Wolters H, Mirza DF, Scholz T, Steininger R, Soderdahl G, Strasser SI, Jauch KW, Neuhaus P, Schlitt HJ, Geissler EK. A prospective randomised, open-labeled, trial comparing sirolimus-containing versus mTOR-inhibitor-free immunosuppression in patients undergoing liver transplantation for hepatocellular carcinoma. BMC Cancer 2010; 10: 190 [PMID: 20459775 DOI: 10.1186/1471 -2407-10-190]
- 173 Gomez-Martin C, Bustamante J, Castroagudin JF, Salcedo M, Garralda E, Testillano M, Herrero I, Matilla A, Sangro B. Efficacy and safety of sorafenib in combination with mammalian target of rapamycin inhibitors for recurrent hepatocellular carcinoma after liver transplantation. *Liver Transpl* 2012; 18: 45-52 [PMID: 21932373 DOI: 10.1002/lt.22434]
- 174 McCaughan GW, Vajdic CM. De novo malignant disease after liver transplantation? Risk and surveillance strategies. *Liver Transpl* 2013; 19 Suppl 2: S62-S67 [PMID: 24019077 DOI: 10.1002/lt.23738]
- 175 Collett D, Mumford L, Banner NR, Neuberger J, Watson C. Comparison of the incidence of malignancy in recipients of different types of organ: a UK Registry audit. Am J Transplant 2010; 10: 1889-1896 [PMID: 20659094 DOI: 10.1111/j.1600-6143.2010.03181.x]
- 176 Aberg F, Pukkala E, Höckerstedt K, Sankila R, Isoniemi H. Risk of malignant neoplasms after liver transplantation: a populationbased study. *Liver Transpl* 2008; 14: 1428-1436 [PMID: 18825704 DOI: 10.1002/lt.21475]
- 177 Engels EA, Pfeiffer RM, Fraumeni JF, Kasiske BL, Israni AK, Snyder JJ, Wolfe RA, Goodrich NP, Bayakly AR, Clarke CA, Copeland G, Finch JL, Fleissner ML, Goodman MT, Kahn A, Koch L, Lynch CF, Madeleine MM, Pawlish K, Rao C, Williams MA, Castenson D, Curry M, Parsons R, Fant G, Lin M. Spectrum of cancer risk among US solid organ transplant recipients. *JAMA* 2011; 306: 1891-1901 [PMID: 22045767 DOI: 10.1001/jama.2011.1592]
- 178 Baccarani U, Piselli P, Serraino D, Adani GL, Lorenzin D, Gambato M, Buda A, Zanus G, Vitale A, De Paoli A, Cimaglia C, Bresadola V, Toniutto P, Risaliti A, Cillo U, Bresadola F, Burra P. Comparison of de novo tumours after liver transplantation with incidence rates from Italian cancer registries. *Dig Liver Dis* 2010; 42: 55-60 [PMID: 19497797 DOI: 10.1016/j.dld.2009.04.017]
- 179 Antinucci F, Anders M, Orozco F, Mella J, Cobos M, McCormack L, Mastai R. [De novo malignant tumors following liver transplantation. A single-center experience in Argentina]. *Medicina* (B Aires) 2015; 75: 18-22 [PMID: 25637895]
- 180 Yao FY, Gautam M, Palese C, Rebres R, Terrault N, Roberts JP, Peters MG. De novo malignancies following liver transplantation: a case-control study with long-term follow-up. *Clin Transplant* 2006; 20: 617-623 [PMID: 16968488 DOI: 10.1111/j.1399-0012.2006.00527.x]



- 181 Jiménez C, Rodríguez D, Marqués E, Loinaz C, Alonso O, Hernández-Vallejo G, Marín L, Rodríguez F, García I, Moreno E. De novo tumors after orthotopic liver transplantation. *Transplant Proc* 2002; 34: 297-298 [PMID: 11959293]
- 182 **Sint Nicolaas J**, de Jonge V, Steyerberg EW, Kuipers EJ, van Leerdam ME, Veldhuyzen-van Zanten SJ. Risk of colorectal carcinoma in post-liver transplant patients: a systematic review and meta-analysis. *Am J Transplant* 2010; **10**: 868-876 [PMID: 20420641 DOI: 10.1111/j.1600-6143.2010.03049.x]
- 183 **Jiménez-Romero** C, Manrique Municio A, Marqués Medina E, Colina F, Ortega Domene P, Gómez Sanz R, Meneu Diaz
- JC, Abradelo de Usera M, Moreno Elola A, Moreno Gonzalez E. Incidence of de novo nonmelanoma skin tumors after liver transplantation for alcoholic and nonalcoholic liver diseases. *Transplant Proc* 2006; **38**: 2505-2507 [PMID: 17097982 DOI: 10.1016/j.transproceed.2006.08.065]
- 184 Jain A, Nalesnik M, Reyes J, Pokharna R, Mazariegos G, Green M, Eghtesad B, Marsh W, Cacciarelli T, Fontes P, Abu-Elmagd K, Sindhi R, Demetris J, Fung J. Posttransplant lymphoproliferative disorders in liver transplantation: a 20-year experience. *Ann Surg* 2002; 236: 429-436; discussion 436-437 [PMID: 12368671 DOI: 10.1097/01.SLA.0000033429.89424.F8]

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