

Contents lists available at ScienceDirect

Bioorganic & Medicinal Chemistry Letters

journal homepage: www.elsevier.com/locate/bmcl



Targeting G-quadruplexes to achieve antiviral activity

Emanuela Ruggiero^a, Sara N. Richter^{a,b,*}

^a Department of Molecular Medicine, University of Padua, Italy
 ^b Microbiology and Virology Unit, Padua University Hospital, Padua, Italy

ARTICLE INFO

Keywords: G-quadruplex Virus Antiviral activity Non-canonical nucleic acids Ligands

ABSTRACT

With the emergence of new viruses in the human population and the fast mutation rates of existing viruses, new antiviral targets and compounds are needed. Most existing antiviral drugs are active against proteins of a handful of viruses. Most of these proteins in the end affect viral nucleic acid processing, but direct nucleic acid targeting is less represented due to the difficulty of selectively acting at the nucleic acid of interest. Recently, nucleic acids have been shown to fold in structures alternative to the classic double helix and Watson and Crick base-pairing. Among these non-canonical structures, G-quadruplexes (G4s) have attracted interest because of their key biological roles that are being discovered. Molecules able to selectively target G4s have been developed and since G4s have been investigated as targets in several human pathologies, including viral infections. Here, after briefly introducing viruses, G4s and the G4-binding molecules with antiviral properties, we comment on the mechanisms at the base of the antiviral activity reported for G4-binding molecules. Understanding how G4-ligands act in infected cells will possibly help designing and developing next-generation antiviral drugs.

Introduction

Virus structure and life cycle

Viruses are microorganisms that must exploit the host organism to replicate. The two basic structural components of viruses are the nucleic acid genome, formed by double- or single-stranded DNA or RNA, and the capsid, a shell made by multiple copies of few viral proteins which protect the viral nucleic acid inside it (Fig. 1).¹ The capsid also serves for cell-entry, genome uncoating and intracellular trafficking.^{2–3} Some viruses have an additional external layer, the envelope, which is derived from the cell membrane of the infected host and modified with inserted and exposed viral glycoproteins. The viral genome may also encode for proteins that promote gene expression and facilitate replication and assembly of the virus particles; usually the simplest and smallest viruses exploit these functions from the host cell machinery.⁴.

The viral genome is composed of either DNA or RNA in different shapes (circular, linear), arrangements (single- or double-stranded) and numerosity (one copy, two copies or fragmented). Based on their genome type, viruses are classified in seven groups according to the Baltimore classification.⁵.

Eight steps are generally necessary for a virus to successfully infect a

host cell, produce new infectious viral particles, the virions, and release them ⁶ (Fig. 2): 1) the virus attaches to a host cell (*attachment*) and 2) if viral proteins recognize cell receptors, the virus genetic material is delivered inside the cell (*entry*) and 3) exposed outside the capsid (*uncoating*); 4) during replication the viral DNA or RNA is replicated (*replication*) and 5),6) viral proteins expressed (*transcription, translation*); 7) new virions are assembled from the newly formed viral proteins and nucleic acids (*assembly*) and 8) released outside the host cell (*release*).

Antiviral drugs

There are 130 viruses reported to infect the human species and cause diseases, which span from benign symptoms that include common cold, respiratory and gastrointestinal manifestations, fever, skin lesions, to severe or fatal outcomes, such as encephalitis, hemorrhagic fever, hepatitis, AIDS, and are accountable for millions of deaths worldwide.⁷.

Hence, development of antiviral drugs has been a major aim since the era of antimicrobial drugs and since the US Food and Drug Administration (FDA) approved the first antiviral drug 'idoxuridine' in 1963 to treat the infection caused by herpes simplex virus.⁷ Emergence of the human immunodeficiency retrovirus (HIV) and consequent acquired immune deficiency syndrome (AIDS) epidemic around the globe during

https://doi.org/10.1016/j.bmcl.2022.129085

Received 1 September 2022; Received in revised form 9 November 2022; Accepted 16 November 2022 Available online 21 November 2022 0960-894X/© 2022 The Author(s). Published by Elsevier Ltd. This is an open access article under the CC BY license (http://creativecommons.org/licenses/by/4.0/).

Abbreviations: G-quadruplex, G4.

^{*} Corresponding author at: Department of Molecular Medicine, University of Padua, Italy. *E-mail address:* sara.richter@unipd.it (S.N. Richter).



Fig. 1. General virus structure with the main viral components shown.

1980s, triggered solid research efforts that led to several antiretroviral drugs becoming available as well as the identification of basic viral molecular mechanisms. Even though successful antiviral drugs have been developed, the intimate interaction of viruses with the host cells has made it difficult to design drugs that are at the same time safe and effective. As a results, to date, about 120 antiviral compounds are available against only 10 human viruses, i.e. HIV, Hepatitis C virus (HCV), Hepatitis B virus (HBV), Influenza virus, Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2), Herpes simplex virus (HSV), Human cytomegalovirus (HCMV), Varicella-zoster virus (VZV), Human papillomavirus (HPV) and Respiratory syncytial virus (RSV).⁸.

Most antiviral drugs are small molecules but proteins, peptides and oligonucleotides are also present. Most of them target viral mechanisms that include nucleic acid replication, entry/fusion, uncoating, protease activity, but compounds that target host cell pathways also exist, either in monotherapies or in combination with drugs that target viral components. The antivirals directed against the host are mainly immuno-modulatory compounds, such as Interferon proteins in different types and formulations that enhance the immune response and elicit an antiviral state. Dexamethasone, an immunosuppressant and anti-inflammatory drug, is used to avoid the severe respiratory complications of SARS-CoV-2 infected and hospitalized patients.⁹ Some drugs inhibit host cytochrome protein CYP3A which in turn would metabolize and inactivate direct antiviral drugs given in combination. One anti-HPV compound inhibits cell division and two other anti-HIV drugs target cell receptors that are required for virus entry into the cell.

A major challenge in antiviral drug development is resistance, which is a commonly reported issue affecting approved antiviral drugs that directly act against viral targets or virus-host interaction. Drug resistance is particularly common in viruses with RNA-dependent RNA polymerase or retrotranscriptase activity due to the error-prone nature of these enzymes (up to 1 mutated nucleotide over 10^4 replicated bases), the rapid rate of viral replication (e.g., 10.3×10^9 and 10^{13} new HIV-1 and HBV virions, respectively, in the human host in 24 h) and frequent recombination events.^{10–11}.

Because of the described issues, new antiviral targets in the virus or in the host cell are highly sought to expand the antiviral drug armamentarium in case of resistant virus strains and towards a larger number of viruses.



Fig. 2. Simplified virus life cycle. 1) Attachment: the virus recognizes and binds to host cell's surface receptors or proteins to enter the cell. 2) Viral entry: according to different pathway, the viral particle enters the cell. 3) Uncoating: upon penetration, the viral genome is uncoated and released into the host cell's cytoplasm. 4) The viral genome nature determines the location and the strategy employed for genome replication: most of DNA viruses replicate in the cell nucleus, while the majority of RNA viruses replicate in the cytoplasm. For retroviruses, retrotranscription takes place in the cytoplasm, while integration occurs in the cell nucleus. 5–6) Transcription and translation: viral genes are transcribed to provide the viral proteins necessary for further steps. 7) Assembly: all newly synthesized viral genomes and proteins are assembled to form new virions. 8) Release: the viral progeny is released outside the host cell by several mechanisms so to infect new cells and propagate.

Non canonical nucleic acid structures: G-quadruplexes

Non-canonical nucleic acid structures form when base pairing other than the classic Watson and Crick is involved. Among them, G-quadruplexes (G4s) are structures that form in G-rich sequences of DNA or RNA(Fig. 3A),¹² when four Gs base pair via Hoogsteen-type hydrogen bonds to yield planar square structures called G-quartets (Fig. 3B). The π - π interactions between the aromatic systems of the G-quartets leads to formation of the G4, when stacking of at least two G-quartets occurs (Fig. 3C). Monovalent cations, usually potassium (K⁺) in the physiological environment, specifically support G4 formation, stability and topology by relieving the repulsion among oxygen atoms that arises in the central cavity. The same sequence can adopt different G4 conformations (Fig. 3C).¹³ Intramolecular (i.e. monomolecular) G4s can form from the following general sequences $G_m X_n G_m X_o G_m X_p G_m$, where m is the number of G residues in each G-tract, which are directly involved in G-quartet Hoogsteen interactions. Xn, Xo and Xp are the linker or loop sequences connecting the G-rich tracts involved in G4 formation and can be any combination of residues, including G. G4 structures with discontinuities in G-stretches causing bulges have also been reported.¹⁴ The recent availability of large datasets on G4 formation has enabled the application of machine learning to predict G4 forming propensity.¹⁵.

Computational as well as deep-sequencing approaches have demonstrated that in the human genome over 700,000 regions exist that could potentially fold into G4 structures.¹⁶ The physiological relevance of G4 structures is further supported by the existence of proteins that are able to bind and unfold G4s (see G4 Interacting Proteins Database).,^{12,1} ¹⁸ Mutations and/or deletions of these proteins lead to modulation in G4 formation.¹⁹ Based on their interaction with proteins and localization in the cell genome, G4s have been implicated in several key biological processes, such as telomere protection and telomerase recruitment,²⁰ genomic and epigenetic instability,^{21–22}²³ obstacles to the replication machinery,²⁴ and triggers of initiation of DNA replication..²⁵ G4s are strongly enriched at promoters, especially of oncogenes: more than 40 % of human promoter regions harbor at least one G4 motif.²⁶. Recent evidence has pointed to G4s as modulators of transcription, where their formation corresponds to increased transcription levels and interaction with transcription factors.^{27–28,29–3}

G-quadruplexes in viruses

Besides mammalian cells, G4s have been described also in yeast, plants, bacteria, archea and viruses. Because of the recent SARS-CoV-2 pandemic, viruses in general have rightfully gained much more

attention than in the past years. Formation of G4s has now been predicted in all human viruses ³¹ and experimentally shown in all the main virus groups. These have been recently reviewed in ^{32,33, 34} Briefly, G4s have been described in double-stranded DNA viruses: in the Aphaherpesvirinae herpes simplex virus types 1 and 2 (HSV-1 and 2) and varicella zoster virus,^{35–37 38} in the *Betaherpesvirinae* human cytomegalovirus (CMV)³⁹⁻⁴⁰ and human herpesvirus 6 (HHV-6),⁴¹ in the Gammaherpesvirinae Epstein-Barr virus (EBV)⁴²⁻⁴³ and Kaposi's sarcoma herpesvirus (KSHV)⁴⁴⁻⁴⁵; in Papillomaviridae⁴⁶ and Adenoviridae.⁴⁷ In viruses with single-strand RNA genome with positive polarity, G4s have been found in Coronaviridae, including SARS-CoV-2,48 Flaviviridae as hepatitis C virus (HCV)⁴⁹ and those transmitted by mosquitos, such as Zika virus (ZIKV)^{50–51} and West Nile virus (WNV).⁵² In the same group, Chikungunya virus (CHIKV)⁵³ belonging to *Togaviridae* and rhinovirus (RhV)⁵⁴ belonging to *Picornaviridae* were shown to present G4s. In viruses with single-stranded RNA genome with negative polarity, G4s have been reported in Ebola (EBOV)⁵⁵ and Marburg (MARV)⁵⁶ viruses belonging to *Filoviridae* and in Influenza virus belonging to Orthomyxoviridae.^{57–58} Finally, G4s have been described in two viruses that present reverse transcriptase activity, i.e. the human immunodeficiency virus (HIV) belonging to Retroviridae, 59-64 where also animal viruses have been shown to display the same G4 arrangement,^{65–66} and in the hepatitis B virus,⁶⁷ belonging to *Hepadnaviridae*.

The viral G4s have been shown to modulate virus replication and transcription in most cases, to affect translation and integration of the viral genome in the host chromosome, ^{41,48} modulate antigen presentation, ⁴³ be involved in virus compartmentalization into vesicles. ⁶⁸ In some cases, viral proteins able to specifically recognize viral and host G4s have been reported; ^{36,69,70-71} in other cases, cell proteins that interact with viral G4s and affect viral replication have been described. ⁷²⁻⁷⁴

G-quadruplex ligands

Because of the relevance of G4s in cell biology, many G4 ligands have been developed to date. Most of them display an aromatic surface that stacks with the external G-tetrads, one positive charge or basic groups that bind to the G4 grooves or loops, and steric hindrance to avoid intercalation with the double-stranded DNA.⁷⁵ Even though these G4 ligands lack traditional 'drug-like' properties, one of them has shown significant accumulation and efficacy in tumor xenografts of human cancers.⁷⁶ Some compounds have reduced planarity while maintaining G4 binding due to their interaction with the groove and backbone phosphates.



Fig. 3. G-quadruplex structure. A) A G4 formed within double-stranded DNA; B) Hoogsteen H-bonds among four guanines which form a G-tetrad; C) Different topologies of intramolecular G4 structures.

To date, around 2800 small molecules targeting G4 structures have been reported (see G-Quadruplex Ligands Database 2.2 https://www. g4ldb.com/).⁷⁷ Two of these ligands have entered clinical trials (Fig. 4A). CX-3543, also named quarfloxin, is a fluoroquinolone that disrupts the binding between ribosomal G4s and nucleolin in the nucleolus, thus inhibiting ribosome biogenesis.78 Although CX-3543 advanced to phase I clinical trials as a candidate therapeutic agent against several tumors, it lacked sufficient efficacy to warrant further clinical development. Another fluoroquinolone, CX-5461, has been recently proved to inhibit topoisomerase II in G4-containing regions, by selectively binding to G4s.⁷⁹ Both CX-3543 and CX-5461 bind and stabilize a broad spectrum of G4 structures, including those at oncogene promoters, e.g. *c-MYC*, *c-KIT*, and telomeres.⁸⁰ From 2016, CX-5461 is in phase I clinical trials for patients with BRCA1/2 deficient tumors, constituting the most advanced G4 ligand in the clinics at the moment.⁸¹.

G4 ligands tested against viruses include BRACO-19, TMPyP4, Pyridostatin and its derivative PDP, PhenDC and its derivatives, and quindoline derivatives (Fig. 4B). These, albeit interacting with both cell and viral G4s, have been shown to exert antiviral activity,^{32,34,38} In some instances, G4 ligands that targets the viral G4s with a certain degree of selectivity have also been developed: ThT-NE, which serves as an indicator of HCV G4s,⁸² and more proper antiviral agents such as the naphthalene diimide derivative c-exNDI,⁸³ gamma-PNAs ⁵² and benzoselenoxanthenes ⁸⁴ (Fig. 4C).

Proposed antiviral modes of action of G-quadruplex-binding compounds

Because of the less than perfect specificity of the employed G4 ligands to the viral G4s, how comes that good antiviral activities have been observed?

Different aspects can be evoked to explain the observed effect (Fig. 5):

1) when a virus infects the host cells, it produces many genomes that are then assembled into new virions to be released from the cell. The number of virions released from the cell is called burst size and comprises both infectious and non-infectious virions. It has been estimated that thousands of new virions are formed and released in a short time within a single eukaryotic cell.^{85–86}.

The number of sequences actually folded in G4 in human cells at any given moment is about 10000.⁸⁷ In the HSV-1 genome, G4s may form

both in about 1,000 bases at repeats ^{31,88} and in the promoters of immediate early genes, i.e. genes that control the viral life cycle. ^{37,36} There has been no count of the G4s that are actually folded in the HSV-1 genome yet, but considering that the typical HSV-1 burst size is about 1,000 virions per infected cell ^{89,90} and having observed that the G4 signal measured by immunofluorescence highly increased upon cell infection and followed the viral genome movement in the host cell, ³⁵ we can safely assume that the number of viral G4s largely exceeds that of host cell G4s, at least in the highly replicating HSV-1 genome. The same types of considerations can be applied to other viruses, especially those with high G/C rich content genomes and fast replications rates.

2) The replicating and newly formed viral genomes are typically devoid of interacting and protecting proteins that vice versa are present in the host cell genome, with its heavy chromatinization. Thus, even the genome of double-stranded viruses during replication is more exposed and more prone to G4 formation. Consequently, when a general G4 ligand is employed, assuming that the affinity towards the different G4 conformations is similar, the antiviral effects can be likely ascribed to the increased viral G4 targets being available upon virus infection and replication in the host cell, paralleled by the lower availability of the ligand for the host cell G4s.

3) A third mechanism can be proposed: all viruses need to exploit the host cell synthetic machinery for their own replication. The simplest viruses heavily rely on the host cell, the more complex viruses synthesize several own enzymes but still need elements of the host cell. Many human viruses achieve their fast replication by manipulating the host cell cycle, subverting it to their advantage by interfering with specific steps of the cycle.⁹¹ For instance, some viruses induce quiescent cells to enter the cell cycle or G1-to-S phase transition to increase the available pools of deoxynucleotides and replicate their genomes concomitantly with the synthesis of cellular chromosomes, others arrest the progression from the G2 phase, a period of rapid cell growth and protein synthesis, to the M phase during which cells divide.⁹².

G4-forming sequences are prevalent in genes that trigger the cell cycle 93 and G4 folding at gene promoters has been associated with gene transcription. $^{27-28}$ Thus, an increase in host cell G4s can be expected upon virus infection. The use of G4 ligands could block those genes that are transcriptionally modulated, rendering the cell less permissive to viral infection and, in the end, exerting indirect antiviral activity. For instance, HSV-1 has been reported to modulate the amount of MYC, $^{94-95}$ the promoter of which contains G4s that are normally targeted by G4



Fig. 4. Chemical structures of the discussed G4 ligands.



Fig. 5. Proposed G4-mediated antiviral mechanisms. 1) The abundance of viral vs host genome, occurring upon virus infection, drives G4-ligands towards viral G4s with respect to the cellular ones. 2) While the host genome is present inside the cell nucleus in a highly condensed (chromatin) form, the viral genome is more accessible to G4 ligands, especially right after being newly synthesized. 3) Since viruses exploit the host transcription machinery to replicate their genome and propagate, targeting transcriptionally active host G4s might indirectly impair virus replication, thus providing a general antiviral effect. 4) When viral replication occurs within the cytoplasm, antiviral G4 ligands that accumulate in the cell nucleus most likely exert their activity trough modulation of host G4s. The blue "DRUG" label indicates a generic G4 ligand.

ligands. In other instances, G4s may be linked to cell receptors that mediate virus entry, as recently shown in SARS-CoV-2 infection.⁴⁸.

4) A final aspect to be considered is the cell compartment of the G4 target. In RNA viruses, for instance, the G4s are produced and maintained in the cytoplasm, therefore G4 ligands that concentrate in the nucleus would not be effective or could exert indirect antiviral activity as proposed above. In light of this, some reported antiviral activities may be rather ascribed to indirect activity on host cell G4s, especially when a virus, such as RNA viruses, replicates in the cytoplasm while the G4 ligand concentrates in the cell nucleus.

The four proposed mechanisms are not exclusive and can be present at the same time, thus further amplifying the antiviral effects of G4 ligands.

Conclusion and outlook

Considering the emergence of new viruses, the fast mutation rates of the existing ones and the lack of general antiviral drugs, new targets that can be exploited for antiviral purposes are highly sought. G4s could be resourceful elements to be evaluated to this end, since effective antiviral activity can be obtained also when a molecule is not tailored against one single G4. *In vivo* and clinical data will be needed to assess whether G4binding compounds can effectively be developed into next-generation antiviral drugs.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

No data was used for the research described in the article.

Acknowledgments

This work was funded by the Bill and Melinda Gates Foundation [grant numbers OPP1035881, OPP1097238] and the Italian Ministry of Education, University and Research [PRIN-2020 2020KSY3KL].

References

- 1 E. Domingo Introduction to virus origins and their role in biological evolution In: Virus as Populations. Elsevier 2020: 1 –33 10.1016/B978-0-12-816331-3.00001-5.
- 2 Freire JM, Santos NC, Veiga AS, Da Poian AT, Castanho MARB. Rethinking the capsid proteins of enveloped viruses: multifunctionality from genome packaging to genome transfection. *FEBS J.* 2015;282:2267–2278. https://doi.org/10.1111/febs.13274.
- 3 Mateu MG. Assembly, stability and dynamics of virus capsids. Arch Biochem Biophys. 2013;531:65–79. https://doi.org/10.1016/j.abb.2012.10.015.
- 4 Banerjee N, Mukhopadhyay S. Viral glycoproteins: biological role and application in diagnosis. VirusDisease. 2016;27:1–11. https://doi.org/10.1007/s13337-015-0293-5.
- Mahmoudabadi G, Phillips R. A comprehensive and quantitative exploration of thousands of viral genomes. *Elife*. 2018;7:e31955.
- 6 Ryu WS. Virus life cycle. In: Molecular virology of human pathogenic viruses. Elsevier; 2017:31-45. doi:10.1016/B978-0-12-800838-6.00003-5.
- 7 Tompa DR, Immanuel A, Srikanth S, Kadhirvel S. Trends and strategies to combat viral infections: A review on FDA approved antiviral drugs. *Int J Biol Macromol.* 2021; 172:524–541. https://doi.org/10.1016/j.ijbiomac.2021.01.076.
- 8 Ianevski A, Yao R, Simonsen RM, et al. Mono- and combinational drug therapies for global viral pandemic preparedness. *iScience*. 2022;25, 104112. https://doi.org/ 10.1016/j.isci.2022.104112.
- 9 The RECOVERY Collaborative Group. Dexamethasone in Hospitalized Patients with Covid-19. N Engl J Med. 2021;384:693–704. https://doi.org/10.1056/ NEJMoa2021436.
- 10 Perelson AS, Neumann AU, Markowitz M, Leonard JM, Ho DD. HIV-1 dynamics in vivo: virion clearance rate, infected cell life-span, and viral generation time. *Science*. 1996;271:1582–1586. https://doi.org/10.1126/science.271.5255.1582.
- 11 Whalley SA, Murray JM, Brown D, et al. Kinetics of acute hepatitis B virus infection in humans. J Exp Med. 2001;193:847–854. https://doi.org/10.1084/jem.193.7.847.

- 12 Varshney D, Spiegel J, Zyner K, Tannahill D, Balasubramanian S. The regulation and functions of DNA and RNA G-quadruplexes. *Nat Rev Mol Cell Biol.* 2020;21:459–474. https://doi.org/10.1038/s41580-020-0236-x.
- 13 Bhattacharyya D, Mirihana Arachchilage G, Basu S. Metal cations in G-quadruplex folding and stability. Front Chem. 2016:4. https://doi.org/10.3389/ fchem.2016.00038.
- 14 Mukundan VT, Phan AT. Bulges in G-quadruplexes: broadening the definition of Gquadruplex-forming sequences. J Am Chem Soc. 2013;135:5017–5028. https://doi. org/10.1021/ja310251r.
- 15 Sahakyan AB, Chambers VS, Marsico G, Santner T, Di Antonio M, Balasubramanian S. Machine learning model for sequence-driven DNA G-quadruplex formation. *Sci Rep.* 2017;7:14535. https://doi.org/10.1038/s41598-017-14017-4.
- Chambers VS, Marsico G, Boutell JM, Di Antonio M, Smith GP, Balasubramanian S.
 High-throughput sequencing of DNA G-quadruplex structures in the human genome. Nat Biotechnol. 2015;33:877–881. https://doi.org/10.1038/nbt.3295.
- 17 Brázda V, Háronfková L, Liao J, Fojta M. DNA and RNA quadruplex-binding proteins. Int J Mol Sci. 2014;15:17493–17517. https://doi.org/10.3390/ijms151017493.
- 18 Mishra SK, Tawani A, Mishra A, Kumar A. G4IPDB: A database for G-quadruplex structure forming nucleic acid interacting proteins. *Sci Rep.* 2016;6:38144. https:// doi.org/10.1038/srep38144.
- 19 Sauer M, Paeschke K. G-quadruplex unwinding helicases and their function in vivo. Biochem Soc Trans. 2017;45:1173–1182. https://doi.org/10.1042/BST20170097.
- 20 Moye AL, Porter KC, Cohen SB, et al. Telomeric G-quadruplexes are a substrate and site of localization for human telomerase. *Nat Commun.* 2015;6:7643. https://doi. org/10.1038/ncomms8643.
- 21 Paeschke K, Bochman ML, Garcia PD, et al. Pif1 family helicases suppress genome instability at G-quadruplex motifs. *Nature*. 2013;497:458–462. https://doi.org/ 10.1038/nature12149.
- 22 De Magis A, Manzo SG, Russo M, et al. DNA damage and genome instability by Gquadruplex ligands are mediated by R loops in human cancer cells. *Proc Natl Acad Sci.* 2019;116:816–825. https://doi.org/10.1073/pnas.1810409116.
- 23 Puig Lombardi E, Holmes A, Verga D, Teulade-Fichou MP, Nicolas A, Londoño-Vallejo A. Thermodynamically stable and genetically unstable G-quadruplexes are depleted in genomes across species. *Nucleic Acids Res.* 2019;47:6098–6113. https:// doi.org/10.1093/nar/gkz463.
- 24 Guilbaud G, Murat P, Recolin B, et al. Local epigenetic reprogramming induced by Gquadruplex ligands. Nat Chem. 2017;9:1110–1117. https://doi.org/10.1038/ nchem.2828.
- 25 Besnard E, Babled A, Lapasset L, et al. Unraveling cell type-specific and reprogrammable human replication origin signatures associated with G-quadruplex consensus motifs. *Nat Struct Mol Biol.* 2012;19:837–844. https://doi.org/10.1038/ nsmb.2339.
- 26 Huppert JL, Balasubramanian S. G-quadruplexes in promoters throughout the human genome. Nucleic Acids Res. 2007;35:406–413. https://doi.org/10.1093/nar/gkl1057.
- 27 Lago S, Nadai M, Cernilogar FM, et al. Promoter G-quadruplexes and transcription factors cooperate to shape the cell type-specific transcriptome. *Nat Commun.* 2021; 12:3885. https://doi.org/10.1038/s41467-021-24198-2.
- 28 Spiegel J, Cuesta SM, Adhikari S, Hänsel-Hertsch R, Tannahill D, Balasubramanian S. G-quadruplexes are transcription factor binding hubs in human chromatin. *Genome Biol.* 2021;22:117. https://doi.org/10.1186/s13059-021-02324-z.
- 29 Robinson J, Raguseo F, Nuccio SP, Liano D, Di Antonio M. DNA G-quadruplex structures: more than simple roadblocks to transcription? *Nucleic Acids Res.* 2021;49: 8419–8431. https://doi.org/10.1093/nar/gkab609.
- 30 Li C, Wang H, Yin Z, et al. Ligand-induced native G-quadruplex stabilization impairs transcription initiation. *Genome Res.* 2021;31:1546–1560. https://doi.org/10.1101/ gr.275431.121.
- 31 Lavezzo E, Berselli M, Frasson I, et al. G-quadruplex forming sequences in the genome of all known human viruses: A comprehensive guide. Lexa M, ed. PLOS Comput Biol. 2018;14(12):e1006675. doi:10.1371/journal.pcbi.1006675.
- 32 Ruggiero E, Zanin I, Terreri M, Richter SN. G-Quadruplex Targeting in the Fight against Viruses: An Update. Int J Mol Sci. 2021;22:10984. https://doi.org/10.3390/ ijms222010984.
- 33 Ruggiero E, Richter SN. Viral G-quadruplexes: New frontiers in virus pathogenesis and antiviral therapy. In: Annual Reports in Medicinal Chemistry. Vol 54. Elsevier; 2020:101-131. doi:10.1016/bs.armc.2020.04.001.
- 34 Abiri A, Lavigne M, Rezaei M, et al. Unlocking G-quadruplexes as antiviral targets. Touyz R, ed. *Pharmacol Rev.* 2021;73(3):897-923. doi:10.1124/ pharmrev.120.000230.
- 35 Artusi S, Perrone R, Lago S, et al. Visualization of DNA G-quadruplexes in herpes simplex virus 1-infected cells. *Nucleic Acids Res.* 2016;gkw968. doi:10.1093/nar/ gkw968.
- 36 Frasson I, Soldà P, Nadai M, Lago S, Richter SN. Parallel G-quadruplexes recruit the HSV-1 transcription factor ICP4 to promote viral transcription in herpes virusinfected human cells. *Commun Biol.* 2021;4:510. https://doi.org/10.1038/s42003-021-02035-y.
- 37 Frasson I, Nadai M, Richter SN. Conserved G-quadruplexes regulate the immediate early promoters of human alphaherpesviruses. *Molecules*. 2019;24:2375. https://doi. org/10.3390/molecules24132375.
- 38 Frasson I, Soldà P, Nadai M, et al. Quindoline-derivatives display potent Gquadruplex-mediated antiviral activity against herpes simplex virus 1. Antiviral Res. 2022;208, 105432. https://doi.org/10.1016/j.antiviral.2022.105432.
- 39 Kumar S, Ramamurthy C, Choudhary D, et al. Contrasting roles for G-quadruplexes in regulating human Bcl-2 and virus homologues KSHV KS-Bcl-2 and EBV BHRF1. *Sci Rep.* 2022;12:5019. https://doi.org/10.1038/s41598-022-08161-9.

- 40 Bohálová N, Cantara A, Bartas M, et al. Analyses of viral genomes for G-quadruplex forming sequences reveal their correlation with the type of infection. *Biochimie*. 2021;186:13–27. https://doi.org/10.1016/j.biochi.2021.03.017.
- 41 Gilbert-Girard S, Gravel A, Artusi S, et al. Stabilization of Telomere G-Quadruplexes Interferes with Human Herpesvirus 6A Chromosomal Integration. Longnecker RM, ed. J Virol. 2017;91(14):e00402-17. doi:10.1128/JVI.00402-17.
- 42 Granzhan A, Martins RP, Fåhraeus R, Blondel M, Teulade-Fichou MP. Quadruplexinteracting compounds for regulating the translation of the Epstein–Barr virus nuclear antigen 1 (EBNA1) mRNA: A new strategy to prevent and treat EBV-related cancers. In: Annual Reports in Medicinal Chemistry. Vol 54. Elsevier; 2020:243-286. doi:10.1016/bs.armc.2020.05.001.
- 43 Murat P, Zhong J, Lekieffre L, et al. G-quadruplexes regulate Epstein-Barr virus-encoded nuclear antigen 1 mRNA translation. *Nat Chem Biol.* 2014;10: 358–364. https://doi.org/10.1038/nchembio.1479.
- 44 Biswas B, Kandpal M, Jauhari UK, Vivekanandan P. Genome-wide analysis of Gquadruplexes in herpesvirus genomes. *BMC Genomics*. 2016;17:949. https://doi.org/ 10.1186/s12864-016-3282-1.
- 45 Kumar S, Choudhary D, Patra A, Bhavesh NS, Vivekanandan P. Analysis of Gquadruplexes upstream of herpesvirus miRNAs: evidence of G-quadruplex mediated regulation of KSHV miR-K12–1-9,11 cluster and HCMV miR-US33. BMC Mol Cell Biol. 2020;21:67. https://doi.org/10.1186/s12860-020-00306-w.
- 46 Marušič M, Hošnjak L, Krafčikova P, Poljak M, Viglasky V, Plavec J. The effect of single nucleotide polymorphisms in G-rich regions of high-risk human papillomaviruses on structural diversity of DNA. *Biochim Biophys Acta BBA - Gen Subj.* 2017;1861:1229–1236. https://doi.org/10.1016/j.bbagen.2016.11.007.
- 47 Majee P, Shankar U, Pasadi S, Muniyappa K, Nayak D, Kumar A. Genome-wide analysis reveals a regulatory role for G-quadruplexes during Adenovirus multiplication. *Virus Res.* 2020;283, 197960. https://doi.org/10.1016/j. virusres.2020.197960.
- 48 Liu G, Du W, Sang X, et al. RNA G-quadruplex in TMPRSS2 reduces SARS-CoV-2 infection. Nat Commun. 2022;13:1444. https://doi.org/10.1038/s41467-022-29135-5.
- 49 Jaubert C, Bedrat A, Bartolucci L, et al. RNA synthesis is modulated by G-quadruplex formation in Hepatitis C virus negative RNA strand. *Sci Rep.* 2018;8:8120. https:// doi.org/10.1038/s41598-018-26582-3.
- 50 Fleming AM, Ding Y, Alenko A, Burrows CJ. Zika virus genomic RNA possesses conserved G-quadruplexes characteristic of the flaviviridae family. ACS Infect Dis. 2016;2:674–681. https://doi.org/10.1021/acsinfecdis.6b00109.
- 51 Majee P, Pattnaik A, Sahoo BR, et al. Inhibition of Zika virus replication by Gquadruplex-binding ligands. *Mol Ther - Nucleic Acids*. 2021;23:691–701. https://doi. org/10.1016/j.omtn.2020.12.030.
- 52 Sarkar S, Armitage BA. Targeting a potential G-quadruplex forming sequence found in the west nile virus genome by complementary gamma-peptide nucleic acid oligomers. ACS Infect Dis. 2021;7:1445–1456. https://doi.org/10.1021/ acsinfecdis.0e00793.
- 53 Lv L, Cui H, Chen Z, Zhou Y, Zhang L. G-quadruplex ligands inhibit chikungunya virus replication. J Med Virol. 2022;94:2519–2527. https://doi.org/10.1002/ jmv.27622.
- 54 Real-Hohn A, Blaas D. Rhinovirus inhibitors: including a new target, the Viral RNA. Viruses. 2021;13:1784. https://doi.org/10.3390/v13091784.
- 55 Wang SR, Zhang QY, Wang JQ, et al. Chemical targeting of a G-quadruplex RNA in the ebola virus L gene. *Cell Chem Biol.* 2016;23:1113–1122. https://doi.org/ 10.1016/j.chembiol.2016.07.019.
- 56 Krafčíková P, Demkovičová E, Víglaský V. Ebola virus derived G-quadruplexes: Thiazole orange interaction. *Biochim Biophys Acta BBA - Gen Subj.* 2017;1861: 1321–1328. https://doi.org/10.1016/j.bbagen.2016.12.009.
- 57 Brázda V, Porubiaková O, Cantara A, et al. G-quadruplexes in H1N1 influenza genomes. BMC Genomics. 2021;22:77. https://doi.org/10.1186/s12864-021-07377-9.
- 58 Tomaszewska M, Szabat M, Zielińska K, Kierzek R. Identification and structural aspects of G-quadruplex-forming sequences from the influenza A virus genome. Int J Mol Sci. 2021;22:6031. https://doi.org/10.3390/ijms22116031.
- 59 Perrone R, Nadai M, Frasson I, et al. A Dynamic G-quadruplex region regulates the HIV-1 long terminal repeat promoter. J Med Chem. 2013;56:6521–6530. https://doi. org/10.1021/jm400914r.
- 60 R. Perrone M. Nadai J.A. Poe et al. Formation of a unique cluster of G-quadruplex structures in the HIV-1 nef Coding Region: implications for Antiviral Activity. Qiu J, ed PLoS ONE. 2013;8(8):e73121. 10.1371/journal.pone.0073121.
- 61 De Nicola B, Lech CJ, Heddi B, et al. Structure and possible function of a Gquadruplex in the long terminal repeat of the proviral HIV-1 genome. *Nucleic Acids Res.* 2016;44:6442–6451. https://doi.org/10.1093/nar/gkw432.
- 62 Butovskaya E, Heddi B, Bakalar B, Richter SN, Phan AT. Major G-quadruplex form of HIV-1 LTR reveals a (3 + 1) folding topology containing a stem-loop. *J Am Chem Soc.* 2018;140:13654–13662. https://doi.org/10.1021/jacs.8b05332.
- 63 Piekna-Przybylska D, Sullivan MA, Sharma G, Banbara RA. U3 region in the HIV-1 genome adopts a G-quadruplex structure in its RNA and DNA sequence. *Biochemistry*. 2014;53:2581–2593. https://doi.org/10.1021/bi4016692.
- 64 Piekna-Przybylska D, Bambara RA, Maggirwar SB, Dewhurst S. G-quadruplex ligands targeting telomeres do not inhibit HIV promoter activity and cooperate with latency reversing agents in killing latently infected cells. *Cell Cycle*. 2020;19:2298–2313. https://doi.org/10.1080/15384101.2020.1796268.
- 65 Ruggiero E, Tassinari M, Perrone R, Nadai M, Richter SN. Stable and conserved G-Quadruplexes in the long terminal repeat promoter of retroviruses. ACS Infect Dis. 2019;5:1150–1159. https://doi.org/10.1021/acsinfecdis.9b00011.

E. Ruggiero and S.N. Richter

- 66 Perrone R, Lavezzo E, Palù G, Richter SN. Conserved presence of G-quadruplex forming sequences in the Long Terminal Repeat Promoter of Lentiviruses. *Sci Rep.* 2017;7:2018. https://doi.org/10.1038/s41598-017-02291-1.
- 67 Meier-Stephenson V, Badmalia MD, Mrozowich T, et al. Identification and characterization of a G-quadruplex structure in the pre-core promoter region of hepatitis B virus covalently closed circular DNA. J Biol Chem. 2021;296, 100589. https://doi.org/10.1016/j.jbc.2021.100589.
- 68 Artusi S, Ruggiero E, Nadai M, et al. Antiviral activity of the G-quadruplex ligand TMPyP4 against herpes simplex virus-1. Viruses. 2021;13:196. https://doi.org/ 10.3390/v13020196.
- 69 Lavigne M, Helynck O, Rigolet P, et al. SARS-CoV-2 Nsp3 unique domain SUD interacts with guanine quadruplexes and G4-ligands inhibit this interaction. *Nucleic Acids Res.* 2021;49:7695–7712. https://doi.org/10.1093/nar/gkab571.
- 70 Butovskaya E, Soldà P, Scalabrin M, Nadai M, Richter SN. HIV-1 Nucleocapsid Protein Unfolds Stable RNA G-Quadruplexes in the Viral Genome and Is Inhibited by G-Quadruplex Ligands. ACS Infect Dis. 2019;5:2127–2135. https://doi.org/10.1021/ acsinfecdis.9b00272.
- 71 Rajendran A, Endo M, Hidaka K, et al. HIV-1 nucleocapsid proteins as molecular chaperones for tetramolecular antiparallel G-quadruplex formation. J Am Chem Soc. 2013;135:18575–18585. https://doi.org/10.1021/ja409085j.
- 72 Tosoni E, Frasson I, Scalabrin M, et al. Nucleolin stabilizes G-quadruplex structures folded by the LTR promoter and silences HIV-1 viral transcription. *Nucleic Acids Res.* 2015;43:8884–8897. https://doi.org/10.1093/nar/gkv897.
- 73 Scalabrin M, Frasson I, Ruggiero E, et al. The cellular protein hnRNP A2/B1 enhances HIV-1 transcription by unfolding LTR promoter G-quadruplexes. *Sci Rep.* 2017;7: 45244. https://doi.org/10.1038/srep45244.
- 74 Ruggiero E, Frasson I, Tosoni E, et al. Fused in liposarcoma protein, a new player in the regulation of HIV-1 transcription, binds to known and newly identified LTR Gquadruplexes. ACS Infect Dis. 2022;8:958–968. https://doi.org/10.1021/ acsinfecdis.1e00508.
- 75 Monchaud D, Teulade-Fichou MP. A hitchhiker's guide to G-quadruplex ligands. Org Biomol Chem. 2008;6:627–636. https://doi.org/10.1039/B714772B.
- 76 Marchetti C, Zyner KG, Ohnmacht SA, et al. Targeting multiple effector pathways in pancreatic ductal adenocarcinoma with a G-quadruplex-binding small molecule. J Med Chem. 2018;61:2500–2517. https://doi.org/10.1021/acs.jmedchem.7b01781.
- 77 Li Q, Xiang JF, Yang QF, Sun HX, Guan AJ, Tang YL. G4LDB: a database for discovering and studying G-quadruplex ligands. *Nucleic Acids Res.* 2013;41: D1115–D1123. https://doi.org/10.1093/nar/gks1101.
- 78 Drygin D, Siddiqui-Jain A, O'Brien S, et al. Anticancer activity of CX-3543: a direct inhibitor of rRNA biogenesis. *Cancer Res.* 2009;69:7653–7661. https://doi.org/ 10.1158/0008-5472.CAN-09-1304.
- 79 Xu H, Hurley LH. A first-in-class clinical G-quadruplex-targeting drug. The bench-tobedside translation of the fluoroquinolone QQ58 to CX-5461 (Pidnarulex). *Bioorg Med Chem Lett.* 2022;77, 129016. https://doi.org/10.1016/j.bmcl.2022.129016.
- 80 Xu H, Di Antonio M, McKinney S, et al. CX-5461 is a DNA G-quadruplex stabilizer with selective lethality in BRCA1/2 deficient tumours. *Nat Commun.* 2017;8:14432. https://doi.org/10.1038/ncomms14432.
- 81 Alessandrini I, Recagni M, Zaffaroni N, Folini M. On the road to fight cancer: the potential of G-quadruplex ligands as novel therapeutic agents. *Int J Mol Sci.* 2021;22: 5947. https://doi.org/10.3390/ijms22115947.

- 82 Luo X, Xue B, Feng G, et al. Lighting up the native viral RNA genome with a fluorogenic probe for the live-cell visualization of virus infection. J Am Chem Soc. 2019;141:5182–5191. https://doi.org/10.1021/jacs.8b10265.
- 83 Perrone R, Doria F, Butovskaya E, et al. Synthesis, binding and antiviral properties of potent core-extended naphthalene diimides targeting the HIV-1 long terminal repeat promoter G-quadruplexes. J Med Chem. 2015;58:9639–9652. https://doi.org/ 10.1021/acs.jmedchem.5b01283.
- 84 Shen LW, Qian MQ, Yu K, et al. Inhibition of Influenza A virus propagation by benzoselenoxanthenes stabilizing TMPRSS2 Gene G-quadruplex and hence downregulating TMPRSS2 expression. *Sci Rep.* 2020;10:7635. https://doi.org/10.1038/ s41598-020-64368-8.
- 85 Milo R, Jorgensen P, Moran U, Weber G, Springer M. BioNumbers—the database of key numbers in molecular and cell biology. *Nucleic Acids Res.* 2010;38(suppl_1): D750–D753. https://doi.org/10.1093/nar/gkp889.
- 86 Sender R, Bar-On YM, Gleizer S, et al. The total number and mass of SARS-CoV-2 virions. Proc Natl Acad Sci. 2021;118(25):e2024815118. doi:10.1073/ pnas.2024815118.
- 87 Hänsel-Hertsch R, Beraldi D, Lensing SV, et al. G-quadruplex structures mark human regulatory chromatin. Nat Genet. 2016;48:1267–1272. https://doi.org/10.1038/ ng.3662.
- 88 Artusi S, Nadai M, Perrone R, et al. The Herpes Simplex Virus-1 genome contains multiple clusters of repeated G-quadruplex: Implications for the antiviral activity of a G-quadruplex ligand. *Antiviral Res.* 2015;118:123–131. https://doi.org/10.1016/j. antiviral.2015.03.016.
- 89 Alandijany T, Roberts APE, Conn KL, et al. Distinct temporal roles for the promyelocytic leukaemia (PML) protein in the sequential regulation of intracellular host immunity to HSV-1 infection. Hutt-Fletcher L, ed. *PLOS Pathog.* 2018;14(1): e1006769. doi:10.1371/journal.ppat.1006769.
- 90 Shipley MM, Renner DW, Ott M, et al. Genome-wide surveillance of genital herpes simplex virus type 1 from multiple anatomic sites over time. *J Infect Dis.* 2018;218: 595–605. https://doi.org/10.1093/infdis/jiy216.
- 91 Bagga S, Bouchard MJ. Cell Cycle Regulation During Viral Infection. In: Noguchi E, Gadaleta MC, eds. Cell Cycle Control. Vol 1170. Methods in Molecular Biology. Springer New York; 2014:165-227. doi:10.1007/978-1-4939-0888-2 10.
- 92 Fan Y, Sanyal S, Bruzzone R. Breaking bad: how viruses subvert the cell cycle. Front Cell Infect Microbiol. 2018;8:396. https://doi.org/10.3389/fcimb.2018.00396.
- 93 Kosiol N, Juranek S, Brossart P, Heine A, Paeschke K. G-quadruplexes: a promising target for cancer therapy. *Mol Cancer*. 2021;20:40. https://doi.org/10.1186/s12943-021-01328-4.
- 94 Birkenheuer CH, Danko CG, Baines JD. Herpes Simplex Virus 1 Dramatically Alters Loading and Positioning of RNA Polymerase II on Host Genes Early in Infection. Sandri-Goldin RM, ed. J Virol. 2018;92(8):e02184-17. doi:10.1128/JVI.02184-17.
- 95 Polpitiya Arachchige S, Henke W, Pramanik A, Kalamvoki M, Stephens EB. Analysis of Select Herpes Simplex Virus 1 (HSV-1) Proteins for Restriction of Human Immunodeficiency Virus Type 1 (HIV-1): HSV-1 gM Protein Potently Restricts HIV-1 by Preventing Intracellular Transport and Processing of Env gp160. Sandri-Goldin RM, ed. J Virol. 2018;92(2):e01476-17. doi:10.1128/JVI.01476-17.