

Cite this: *Dalton Trans.*, 2022, **51**, 11135

Cationic palladium(II)-indenyl complexes bearing phosphines as ancillary ligands: synthesis, and study of indenyl amination and anticancer activity†

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The reactivity of palladium(II) indenyl derivatives and their applications are topics relatively less studied, though in recent times these compounds have been used as pre-catalysts able to promote challenging cross-coupling processes. Herein, we propose the first systematic study concerning the nucleophilic attack on the palladium(II) coordinated indenyl fragment and, for this purpose, we have prepared a library of new Pd-indenyl complexes bearing mono- or bidentate phosphines as spectator ligands, developing specific synthetic strategies. All novel compounds are thoroughly characterized, highlighting that the indenyl ligand presents always a hapticity intermediate between η^3 and η^5 . Secondary amines have been chosen as nucleophiles for the present study and indenyl amination has been monitored by UV-Vis and NMR spectroscopies, deriving a second order rate law, with dependence on both complex and amine concentrations. The rate-determining step of the process is the initial attack of the amine to the coordinated indenyl fragment, and this conclusion has been supported also by DFT calculations. The determination of second order rate constants has allowed us to assess the impact of the phosphine ligands on the kinetics of the process and identify the steric and electronic descriptors most suitable for predicting the reactivity of these systems. Finally, *in vitro* tests have proven that these organometallic compounds promote antiproliferative activity towards ovarian cancer cells better than cisplatin and possibly by adopting a different mechanism of action.

Received 9th June 2022,
Accepted 28th June 2022
DOI: 10.1039/d2dt01821g

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Introduction

The Pd-indenyl organometallic functional group has received relatively less attention, especially when considering the huge number of publications devoted to palladium-allyl derivatives.¹ This is rather odd because some peculiar structural and bonding motifs of indenyl fragments could give these systems a reactivity of great interest. In this context, the central issue con-

cerns the indenyl hapticity that may theoretically be η^1 , η^3 or η^5 , with the further option of intermediate configurations and the possibility of easily switching from one to another. It is well known that electron-poor complexes of early and middle transition metals generally prefer the highest hapticity to maximize the formal electron count and ensure electronic saturation. Conversely, more electron-rich metal centres can accept lower hapticity with consequent slip-fold distortion of the indenyl ligand. The latter feature, which is mainly attributable to benzenoid resonance stabilization, makes the indenyl ligand more versatile than the cyclopentadienyl congener, which hardly changes its η^5 configuration (*indenyl effect*).² An up-to-date overview of the literature on palladium indenyl compounds highlights that these systems seem to generally prefer η^3 hapticity although some notable examples of Pd- η^1 -indenyl complexes are well-known.²⁻⁴ In most cases they are neutral mononuclear or dinuclear Pd(II) complexes, with structures influenced by the nature of ancillary ligands. Another important class of compounds is represented by dinuclear Pd(I) species, in which the indenyl fragment is μ^2 bridging⁴⁻⁷ (Scheme 1).

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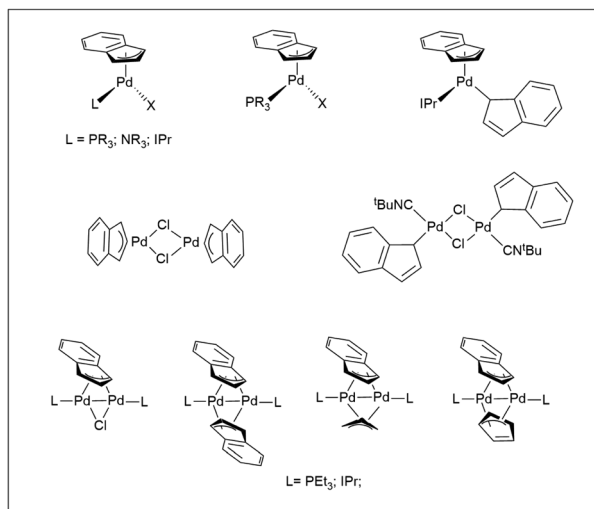
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† Electronic supplementary information (ESI) available. CCDC 2173701–2173705. For ESI and crystallographic data in CIF or other electronic format see DOI: <https://doi.org/10.1039/d2dt01821g>





Scheme 1 Significant examples of Pd(II)- and Pd(I)-indenyl complexes.^{4–7}

Remarkably, all complexes are characterized by a formal electron count of 16 and somehow, this feature ends up determining the hapticity of the indenyl ligand.

However, it should be stressed that the η^3 -binding mode of the complexes reported in Scheme 1 has a certain percentage of η^5 -character, which can be quantified by well-defined structural^{2,8} and spectroscopic⁹ parameters. Generally speaking, an increase of the degree of pentahapto coordination mode is associated with approaching to the planarity of the five-membered ring with its five carbons tending to have very similar distances from the metal centre.

Seminal works of the Zargarian group have shown the capability of some Pd(II)-indenyl derivatives to catalyse a wide range of organic processes such as isomerization, oligomerization and polymerization of olefins and Mizoroki–Heck coupling.^{3,10,11} More recently, Hazari and Nolan have developed a series of precatalysts of the type [PdCl(1-^tBu-indenyl)(L)] (L) (with L = NHC or Buchwald-type phosphine) able to promote a number of challenging cross-coupling processes such as α -arylation of methyl ketones, and Suzuki–Miyaura and Buchwald–Hartwig reactions with deactivated substrates.¹² The key to success of these derivatives stems from the presence of the 1-^tBu-indenyl scaffold which facilitates the selective formation of active Pd(0)-monoligated species disfavoring the production of inactive Pd(I) dimers.

The literature provides much less information on cationic palladium-indenyl species of the type [Pd(indenyl)(L)₂]⁺,^{11,13} and above all no specific studies of their reactivity can be found. This is particularly striking in relation to the large number of publications dealing with their η^3 -allyl counterparts. For example, it is well-known that these derivatives are key-intermediates in the Tsuji–Trost catalytic cycle during which they are subjected to nucleophilic attack on the allyl fragment by many suitable substrates resulting in allyl substituted products.¹⁴ In the past, our research group has studied

the kinetics and reaction mechanisms of palladium-allyl amination, assessing the influence of spectator ligands, allyl substituents, the type of amine and solvent.¹⁵

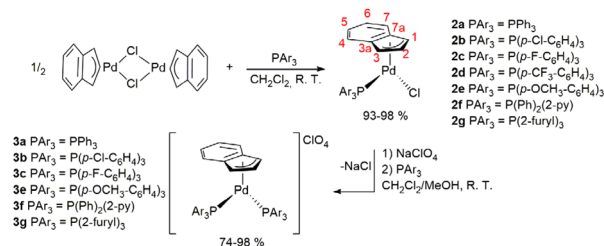
In this work we aim at filling the knowledge gap concerning the nucleophilic attack on palladium(II) coordinated indenyl. The choice of an amine as a nucleophilic agent allows us to take advantage of the information obtained from our previous studies on allyl derivatives and, at the same time, compare the behaviour of these two organometallic systems. In this regard, we have opted to prepare an array of Pd(II)-indenyl complexes equipped with aryl(hetero)phosphines as supporting ligands, since they have been shown to be efficient for promoting allyl amination and also, as is well-known, their steric/electronic features can be easily fine-tuned.

Finally, encouraged by the recent results obtained on the antiproliferative activity of cationic palladium- η^3 -allyl derivatives towards cancer cells,¹⁷ we wanted to see if these promising properties could also be extended to these novel Pd(II)-indenyl derivatives.

Results and discussion

Synthesis of neutral palladium-indenyl complexes

Firstly, we have prepared a series of neutral complexes of the type [PdCl(indenyl)(PAR₃)], adopting the original procedure described by Zargarian and co-workers and based on the dimer [Pd(μ -Cl)(indenyl)]₂ as a starting material.³ The library of the selected phosphines includes *p*-substituted phenyl and heteroaromatic derivatives, as shown in Scheme 2. All reactions proceed smoothly, and even the more deactivated phosphines afforded the target products in good yields. NMR analysis of the compounds confirms unambiguously their identity. The coordination of entering phosphine is validated by the presence of a single peak in ³¹P{¹H} NMR spectra, shifted 30–40 ppm downfield from the free phosphine, with the only noteworthy exception of a 2-trifurylphosphine derivative (in this case $\Delta\delta = -17$ ppm) but the peculiar behaviour of this phosphine has already been observed in complexes with other metal centres.¹⁶ Consistently, the asymmetry induced by the two different ancillary ligands differentiates all the signals ascribable to the protons and carbons of the indenyl fragment. In more detail, it can be noted that carbon C3 and its associated proton H3 (labelling shown in Scheme 2) exhibit lower



Scheme 2 Synthesis of neutral and cationic Pd(II)-indenyl complexes bearing monodentate phosphines.



chemical shifts than C1 and H1, ($\Delta\delta$ of 20 and 2 ppm, respectively). This effect reflects a non-symmetrical Pd–C interaction and can be easily explained by the higher *trans*-influence of phosphine compared to chloride.

Further information about the palladium-indenyl interaction can be deduced by the chemical shift of junction carbons C3a and C7a. As a matter of fact, according to the Baker and Marder empirical protocol,⁹ the magnitude of $\Delta\delta$ (^{13}C) = δ (C3a/7a of M-Ind) – δ (C3a/7a of Na⁺Ind[−]) is correlated with the hapticity of the indenyl fragment in solution. In this series of compounds $\Delta\delta$ (^{13}C) is always between +2.8 and +6.2 ppm, (Table S7 in the ESI[†]) basically indicating intermediate hapticity between η^3 and η^5 .

Synthesis of cationic palladium-indenyl complexes bearing two monodentate phosphines

The simple addition of PAR_3 to a solution of the generic neutral complex $[\text{PdCl}(\text{indenyl})(\text{PAR}_3)]$ does not lead to the formation of the desired cationic derivative $[\text{Pd}(\text{indenyl})(\text{PAR}_3)_2]\text{Cl}$, but induces the reduction of indenyl hapticity to η^1 , affording the intermediate species $[\text{PdCl}(\eta^1\text{-indenyl})(\text{PAR}_3)_2]$ in the first instance. This chemical event triggers cascade reactions with the final production of the bis-indene, $[\text{PdCl}_2(\text{PAR}_3)_2]$ and the dimer $[(\mu\text{-}\eta^3\text{-ind})(\mu\text{-Cl})\text{Pd}_2(\text{PAR}_3)_2]$, which are, in addition to the starting complex, the only palladium species detectable in solution, when the reaction is monitored by NMR spectroscopy (Scheme 3). To overcome this issue, a good option is to preventively remove the chloride ligand from the palladium coordination sphere. As an alternative to the “classical” dehalogenation with silver derivatives,⁶ we have used less expensive sodium salt which, in a 3:1 dichloromethane/methanol mixture, is able to induce the precipitation of sodium chloride after the addition of a second equivalent of phosphine (Scheme 2).

Of course, it is possible to operate without isolating the neutral complex $[\text{PdCl}(\text{indenyl})(\text{PAR}_3)]$, starting directly from $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})_2]$ and adding in succession one equivalent of phosphine, the sodium salt in methanol solution and finally the second equivalent of phosphine. With this synthetic protocol we were able to isolate all the final products, except for the complex bearing two tris(4-trifluoromethylphenyl)phosphine, which decomposes readily during the synthesis.

The only peak exhibited by the $^{31}\text{P}\{^1\text{H}\}$ NMR spectra of the new complexes $[\text{Pd}(\text{indenyl})(\text{PAR}_3)_2]\text{ClO}_4$ is always localised at

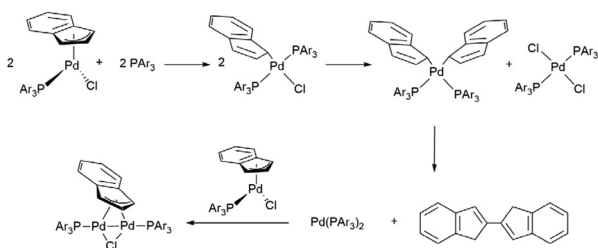
chemical shifts slightly lower than that of the corresponding neutral species $[\text{PdCl}(\text{indenyl})(\text{PAR}_3)]$, (except for the compound coordinating two tri(2-furyl)phosphines). Moreover, the higher symmetry of these complexes greatly simplifies ^1H and $^{13}\text{C}\{^1\text{H}\}$ NMR spectra, where the coincident H1–H3 and C1–C3 resonate at 5.5 and 96 ppm, respectively. From the position of C3a/7a, it is possible to obtain $\Delta\delta$ (^{13}C) values which, being always very close to zero, (see Table S7 in the ESI[†]) indicate that the indenyl ligand exhibits a hapticity intermediate between η^3 and η^5 also in these cationic species. This bonding motif is further confirmed by the structure obtained by single crystal X-ray diffraction of complex **3b** (Fig. 1), from which it was possible to determine the structural parameters $\Delta\text{M-C}$,¹⁸ hinges and fold angles (HA¹⁹ and FA²⁰). Their values (0.298 Å, 13.24° and 11.65°, respectively) are consistent with an intermediate coordination between η^3 and η^5 of the indenyl fragment.²

Finally, the presence of two intense bands at about 1080 and 620 cm^{-1} in the IR spectra, attributable to ν_{ClO_4} and δ_{ClO_4} , respectively, indirectly certifies the cationic nature of the synthesized complexes.

A special case is represented by XPhos (2-dicyclohexylphosphino-2',4',6'-triisopropylbiphenyl) which interacts with the Pd-indenyl fragment in a different way from the other monodentate phosphines investigated so far. This is largely due to its peculiar structure and high steric hindrance.²¹

Indeed, using the same synthetic protocol described above the coordination of only one XPhos molecule can be obtained (Scheme 4). The definitive structure of the final cationic complex $[\text{Pd}(\text{indenyl})(\text{XPhos})]\text{ClO}_4$ (**3h**) was determined by single crystal X-ray analysis and a view of that is shown in Fig. 2.

From this study it emerges that XPhos is anchored to the metal centre with the phosphorus donor atom and *via* a η^2 -



Scheme 3 Cascade reactions in the absence of a dechlorinating agent.

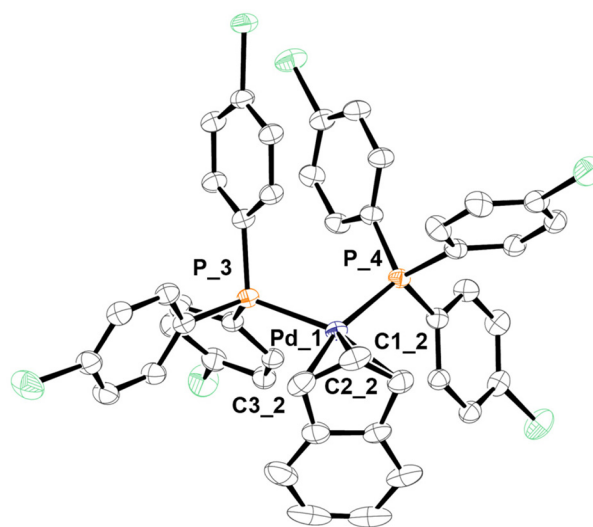
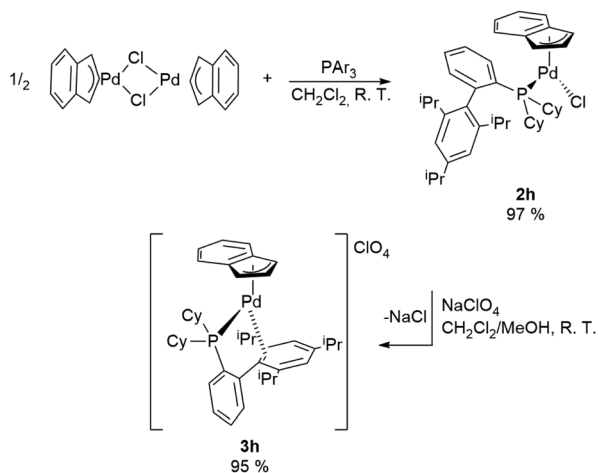


Fig. 1 X-ray molecular structure of **3b**, showing thermal displacement ellipsoids at the 50% probability level with the hydrogen atoms, counterion and solvent molecules omitted for clarity.





Scheme 4 Synthesis of neutral and cationic Pd(II)-indenyl complexes (**2h** and **3h**) bearing XPhos.

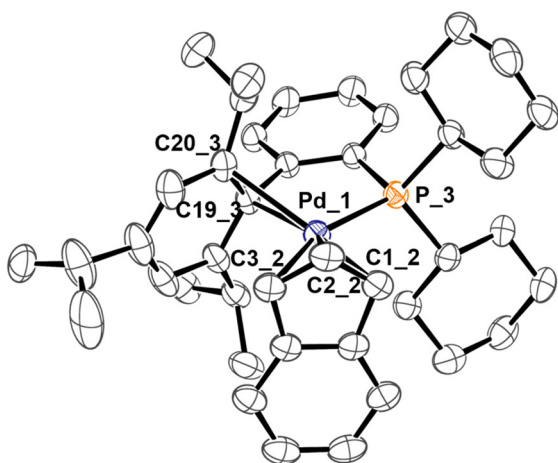


Fig. 2 X-ray molecular structure of **3h**, showing thermal displacement ellipsoids at the 50% probability level with the hydrogen atoms, counterion and solvent molecules omitted for clarity.

coordination of non-phosphine containing an aryl-ring as witnessed by Pd–C_{19_3} and Pd–C_{20_3} distances (2.367 and 2.490 Å, respectively). This binding motif is quite unusual but has already been observed in other palladium(II) complexes.²²

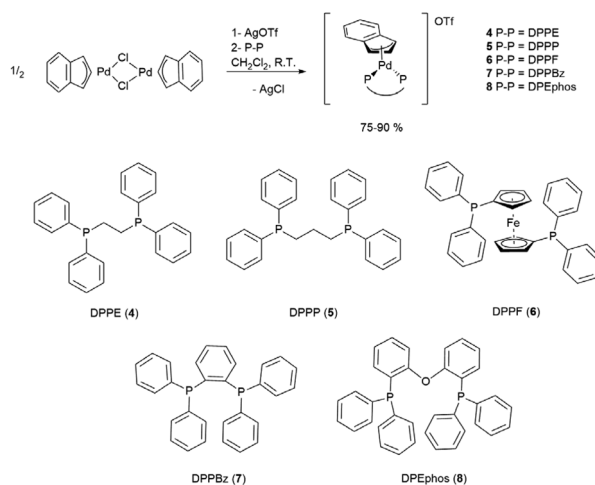
NMR studies confirm that the structure in solution is the same as that in the solid state. As a matter of fact, the integration of the signals in the ¹H NMR spectrum proves that the ratio between the indenyl fragment and coordinated phosphine is 1:1. Moreover, seven and nine different signals attributable to indenyl fragment spectra are traceable in ¹H and ¹³C{¹H} NMR, respectively, as a direct consequence of the described coordination mode of XPhos. Finally, the only signal present in ³¹P{¹H} NMR spectra resonates at a chemical shift significantly higher than that in free XPhos ($\Delta\delta = 65$ ppm), certifying the coordination of the phosphine to a more electron-poor metal centre than those of the bis-phosphine-cationic complexes described above.

Synthesis of cationic palladium-indenyl complexes bearing bidentate phosphines

The same synthetic strategy followed for complexes described in the previous paragraph could not be used for the title compounds. In fact, the presence of two phosphorus donor atoms in the designated ligands activates the unwanted reaction reported in Scheme 3, immediately after the mixing of bidentate phosphine with the dimeric precursor [Pd(μ-Cl)(indenyl)]₂. This undesired side-reaction cannot be avoided even if a solution of NaClO₄ is added in advance. Therefore, we were forced to opt for the preliminary treatment of [Pd(μ-Cl)(indenyl)]₂ with silver triflate that ensured exhaustive dechlorination and the formation *in situ* of [Pd(μ-OTf)(indenyl)]₂. The successive addition of bidentate phosphine, followed by the removal of AgCl, afforded the five desired complexes [Pd(indenyl)(P–P)]OTf, with satisfactory yields (Scheme 5).

As usual, NMR analysis helps us to confirm the identity of the isolated compounds. As a natural consequence of the coordination of a symmetric chelate ligand on the metal centre, all ³¹P{¹H} NMR spectra are characterised by the presence of a single peak resonating at chemical shifts significantly higher than that in the uncoordinated diphosphine. Among the signals detectable in ¹H and ¹³C{¹H} NMR spectra, it is possible to easily distinguish those belonging to the indenyl moiety, among which H1/H3 and C1/C3 can be found at 5.3–6.3 and 90–95 ppm, respectively. Also, in these derivatives the hapticity of the indenyl ligand is intermediate between η³ and η⁵, as inferred from the small values of $\Delta\delta(^{13}\text{C})$ and $\Delta\text{M-C}$, and HA and FA parameters (see Tables S2–7 in the ESI[†]). The latter were derived from the X-ray structures of complexes **4**, **5** and **6** (Fig. 3).

Finally, the single peak present in ¹⁹F{¹H} NMR spectra, detectable at a typical resonance frequency of –78 ppm, certifies the presence of the triflate counterion.



Scheme 5 Synthesis of cationic Pd(II)-indenyl complexes bearing bidentate phosphines.



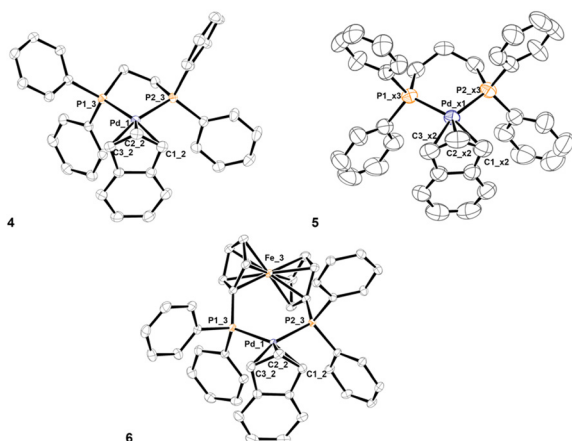
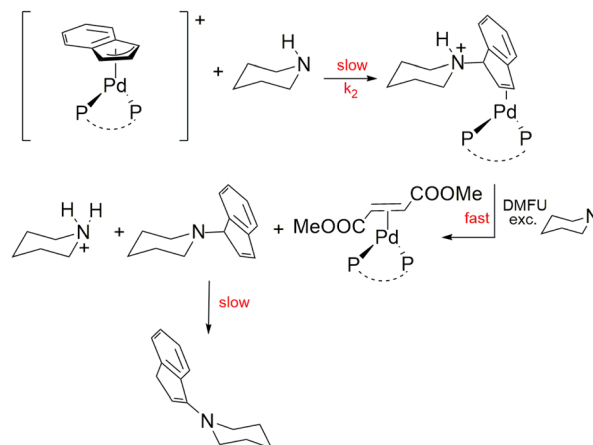


Fig. 3 X-ray molecular structures of **4** (left) and **5** (right) and **6** (down) are presented, showing thermal displacement ellipsoids at the 50% probability level with hydrogen atoms, counterions and solvent molecules omitted for clarity.

Kinetic and computational studies on indenyl amination

Nucleophilic attack on the Pd-indenyl fragment is surprisingly an unexplored topic considering a large amount of information available on the closely related Pd-allyl group. A kinetic and mechanistic study of this reaction could prove especially useful to better understand the distinctive features of the Pd-indenyl organometallic function in view of its use in synthesis, catalysis and medicinal chemistry. Moreover, it can allow us to assess how the electronic and steric characteristics of the phosphine ligands²³ affect this kind of reactivity. A reliable model reaction for probing the susceptibility of the Pd-allyl substrates to nucleophilic attack is typically an amination reaction. In this regard, several studies have amply proved that, in the absence of ligand substitution processes, amine carries out its exclusive attack on the terminal allyl carbon, following a second order kinetic law, with dependence on both complex and amine concentrations.¹⁵ Therefore, in the first instance, a series of preliminary tests have been carried out in order to verify whether this also applied to Pd-indenyl substrates. Piperidine was chosen as a reagent probe for its high reactivity due to its strong nucleophilicity and low steric hindrance. The reaction was studied in excess of amine and in the presence of dimethyl fumarate for stabilizing the resulting palladium(0) derivative. As a matter of fact, after the formation of the Pd- η^2 -indenyl-ammonium complex, the excess of amine promotes its deprotonation and dimethyl fumarate replaces the ensuing indenyl-amine in the Pd(0) coordination sphere. These last two steps are fast and do not affect the reaction rate that is insensitive to the concentration of dimethyl fumarate (Scheme 6). This way of proceeding is particularly useful for preventing the decomposition process that could especially prejudice the spectrophotometric study of the reaction (*vide infra*).

The reaction monitoring by ^1H NMR spectroscopy shows that the initially formed 1-(1*H*-inden-1-yl)piperidine slowly iso-



Scheme 6 Reaction of indenyl amination with piperidine ($[\text{complex}] = 5 \times 10^{-5} - 1 \times 10^{-4} \text{ mol dm}^{-3}$, $[\text{dimethylfumarate}] = 1.5 \times 10^{-4} - 3 \times 10^{-4} \text{ mol dm}^{-3}$, $[\text{piperidine}] \geq 10[\text{complex}]$; in CHCl_3 , $T = 298 \text{ K}$).

merizes to the more thermodynamically stable 1-(1*H*-inden-3-yl)piperidine (using DFT calculations the difference amounts to $9.1 \text{ kcal mol}^{-1}$). This process, which is possibly catalysed by basic conditions²⁴ (excess of piperidine), proceeds, in most cases, much more slowly than the indenyl amination of palladium complexes. It is important to stress that the nature of the final organic product 1-(1*H*-inden-3-yl)piperidine is attested as well as by the presence of its expected signals in the ^1H NMR spectra (in good agreement with those of the simulated spectrum) and by GC-MS analysis of the final reaction mixture (Fig. S96 in the ESI†). The identity of the final complexes $[\text{Pd}(\eta^2\text{-dmfu})(\text{PAR}_3)_2]$ and $[\text{Pd}(\eta^2\text{-dmfu})(\text{P-P})]$ can be unambiguously ascertained by comparing the signals present in the final reaction mixture with those of the ^1H and $^{31}\text{P}\{^1\text{H}\}$ NMR spectra of these Pd(0) derivatives obtained independently in the past by another synthetic route (*e.g.* reacting $[\text{Pd}_2(\text{dba})_3] \cdot \text{CHCl}_3$ with dmfu and the respective phosphine).²⁵

These preliminary studies also provide us with general information about the rate of the process. Under the experimental conditions used (298 K, CDCl_3 , $[\text{Pd-indenyl complex}] = 1 \times 10^{-2} \text{ mol dm}^{-3}$ and $[\text{piperidine}] = 5 \times 10^{-2} \text{ mol dm}^{-3}$), all reactions are completed in a few minutes, with the only exceptions represented by complexes $[\text{Pd}(\text{DPPE})(\text{indenyl})]\text{OTf}$, $[\text{Pd}(\text{DPPP})(\text{indenyl})]\text{OTf}$ and $[\text{Pd}(\text{DPPBz})(\text{indenyl})]\text{OTf}$ which need a few hours to reach completion. Excluding the last cases, for which we have reserved a separate treatment (*vide infra*), a general kinetic study has been carried out taking advantage of UV-Vis spectroscopy. As matter of fact, all our Pd-indenyl derivatives are strongly coloured (deep-orange) and show a strong absorption band at around 400 nm that gradually decreases during the reaction. The presence of an isobestic point, localized in all cases at around 380 nm, suggests the absence of intermediate species at significant concentrations.

For quantitative kinetic measurements, the course of reactions of CHCl_3 solutions of Pd-indenyl complexes (1×10^{-4} or $5 \times 10^{-5} \text{ mol dm}^{-3}$) in the presence of dimethyl fumarate ($3 \times$



10^{-4} or 1.5×10^{-4} mol dm⁻³), upon addition of variable aliquots of a large excess of piperidine, was monitored at a wavelength of 400 nm. Under such *pseudo*-first order conditions the reaction proceeded smoothly to completion in a few minutes, and over this period of time, as proved by previous NMR studies, the isomerization from 1-(1*H*-inden-1-yl)piperidine to 1-(1*H*-inden-3-yl)piperidine can be neglected.

The conversion of Pd-indenyl complexes to the final Pd(0) derivatives appears to obey the customary mono-exponential absorbance (A_t) vs. time (t) relationship (Fig. 4):

$$A_t = A_\infty + (A_0 - A_\infty)\exp(-k_{\text{obs}}t)$$

The *pseudo* first order rate constants k_{obs} were derived by nonlinear regression of absorbance A_t data to time.

For all reactions examined, k_{obs} values fit the simple linear relationship (inset Fig. 4):

$$k_{\text{obs}} = k_2 [\text{piperidine}]$$

Therefore, it can be concluded that the kinetic law governing this process is of the second order, with dependence on both the Pd-indenyl complex and amine concentrations:

$$\text{rate} = k_2 [\text{complex}][\text{piperidine}]$$

This result is in accordance with a mechanism where the initial bimolecular attack of amine to the coordinated indenyl fragment represents the rate-determining step of the process, as already observed for the corresponding Pd-allyl complexes (Scheme 6).

The values of second-order constants relative to complexes bearing monodentate phosphines are listed in Table 1.

In the light of these data, we can propose the following observations:

$$k_{\text{obs}} = k_2 [\text{piperidine}]$$

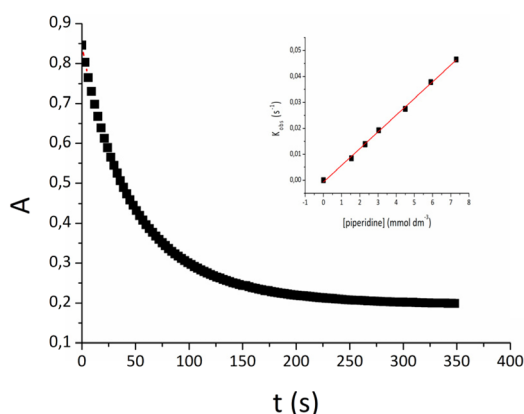


Fig. 4 Fit of absorbance to time for reaction **3e** + piperidine in the presence of dimethyl fumarate ($\lambda = 400$ nm, solvent CHCl₃, $T = 298$ K, $[\mathbf{3e}] = 1 \times 10^{-4}$ mol dm⁻³, $[\text{piperidine}] = 1 \times 10^{-3}$ mol dm⁻³, $[\text{dimethyl fumarate}] = 3 \times 10^{-4}$ mol dm⁻³). The inset shows the linear regression analysis of k_{obs} (s⁻¹) to $[\text{piperidine}]$, for the same reaction.

Table 1 Second order rate constants (k_2 , mol⁻¹ dm³ s⁻¹) for reactions of piperidine with cationic palladium-indenyls coordinating monodentate phosphines in CHCl₃ at 298 K

Compound	k_2 (mol ⁻¹ dm ³ s ⁻¹)
3a	55.5 ± 1.9
3b	420 ± 20
3c	250 ± 20
3e	6.4 ± 0.1
3f	13.3 ± 0.4
3g	410 ± 10
3h	13.1 ± 0.7

(a) Considering the four complexes equipped with monodentate aryl-phosphines **3a–c** and **3e**, the influence of the different substituents in the *para*-position is apparent, with the k_2 values increasing with an increase in the electron-withdrawing nature of the substituents. This observation is in full agreement with the proposed mechanism whereby a more electron-poor metal centre makes the coordinated indenyl fragment more susceptible to nucleophilic attack. This effect can be somewhat systematized recurring to the Hammett equation: fitting log k_2 values vs. tabulated *para*-substituent σ constants, we can obtain the value of the reaction constant ρ amounting to 3.7 ± 0.7 (Fig. S95 in the ESI†).

(b) The particularly high reactivity of [Pd(indenyl)(P(2-furyl)₃)₂]ClO₄ (**3g**) is also due to the high electron-withdrawing nature of hetero-aryl phosphines.

(c) Intriguingly, the complex [Pd(indenyl)(P(Ph)₂(2-py))₂]ClO₄ (**3f**) shows lower reactivity than the very structurally similar [Pd(indenyl)(PPh₃)₂]ClO₄ (**3a**) as opposed to what would be expected considering that the replacement of a phenyl with a more electron withdrawing pyridyl ring should have an activating effect. We can therefore assume that there is some form of stabilization of the starting complex or destabilization of the transition state of the rate-determining step, promoted by pyridyl-nitrogen.

Limited to complex [Pd(indenyl)(PPh₃)₂]ClO₄ (**3a**), we have also determined the rate constants k_2 , when diethylamine and morpholine are used as nucleophiles. The choice of these two different amines was made to better assess the weight of steric and electronic effects on this kind of reaction. As a matter of fact, morpholine has the same steric hindrance as piperidine but less basicity, whereas diethylamine has the same basicity but is more sterically encumbered. Consistently, both resulted in a k_2 value lower than that of piperidine (23 ± 1 and 4.9 ± 0.1 vs. 56 ± 2 , respectively), but the decrease of one order of magnitude recorded for diethylamine seems to indicate that the steric factor plays a more important role than the electronic one.

The complex [Pd(indenyl)(XPhos)]ClO₄ (**3h**), which differs from those examined so far for the presence of an only coordinated phosphine, reacts with piperidine more slowly than expected considering that the indenyl fragment is bound to a more electron-poor metal centre. This fact is possibly due to more steric congestion around the indenyl ligand (see Fig. 2).



For what concerns the complexes equipped with bidentate phosphines, only two of the prepared compounds (**6** and **8**) are reactive enough to determine the relative k_2 rate constant at the concentrations adopted for spectrophotometric studies. For the remaining three complexes (**4**, **5**, and **7**), the reactions have been monitored by $^{31}\text{P}\{^1\text{H}\}$ NMR spectroscopy, which allows working at higher concentrations thus making the process faster. A series of preliminary tests have helped us to define the best conditions to carry out the experiments (see the Experimental section). The two equations used to determine the k_2 rate constants are in this case:

$$d[\text{complex}]/dt = k_2[\text{complex}][\text{piperidine}]$$

$$[\text{piperidine}] = [\text{piperidine}]_0 - 2[\text{complex}]$$

where $[\text{piperidine}]_0$ is the concentration of piperidine added to the initial mixture of an indenyl complex and dimethyl fumarate, and with the second equation we consider that for each amine molecule that attacks the indenyl fragment, another one is needed to deprotonate the resulting indenyl piperidinium intermediate (see Scheme 6). The k_2 values obtained by this approach are also reported in Table 2, together with the corresponding bite-angles of the coordinated diphosphine. From these values, a direct correlation between the amination rate and bite angle may be inferred.

It may finally be interesting to compare the reactivity of Pd-indenyl with that of the Pd-allyl fragment. To this end, we have chosen the two complexes coordinating 1,2 bis(diphenylphosphine)ethane $[\text{Pd}(\text{DPPE})(\text{indenyl})]\text{OTf}$ (**4**) and $[\text{Pd}(\text{DPPE})(\eta^3\text{-allyl})]\text{OTf}$ (**9**). The latter has been synthesized with the same method of its indenyl congener, starting from the dimeric precursor $[\text{Pd}(\mu\text{-Cl})(\eta^3\text{-allyl})]_2$. The first preliminary test carried out in the NMR tube has shown the higher reactivity of the Pd-allyl complex **9** with piperidine compared to **4**, completing the reaction after a few minutes. A more detailed UV-Vis spectrophotometric study has allowed determining the k_2 rate constant relative to this substrate that, amounting to $0.607 \pm 0.002 \text{ mol}^{-1} \text{ dm}^3 \text{ s}^{-1}$, is about one order of magnitude higher than that of the corresponding indenyl derivative **4**. This effect can be attributed to the partial η^5 -character adopted by an indenyl fragment in this type of complex, which strengthens its bond with the palladium centre, and disadvantages the nucleophilic attack.

Table 2 Second order rate constants (k_2 , $\text{mol}^{-1} \text{ dm}^3 \text{ s}^{-1}$) for reactions with piperidine with cationic palladium-indenyl complexes bearing chelate diphosphines in CDCl_3 or CHCl_3 at 298 K and the relative bite angles of the coordinated diphosphines

Compound	k_2 ($\text{mol}^{-1} \text{ dm}^3 \text{ s}^{-1}$)	Bite angle ($^\circ$)
4	$(4.0 \pm 0.2) \times 10^{-2}$	86
5	$(7.1 \pm 0.3) \times 10^{-2}$	91
6	1.81 ± 0.9	99
7	$(2.08 \pm 0.06) \times 10^{-2}$	83
8	8.50 ± 0.4	104
9	0.607 ± 0.002	86

With these data in hand, DFT calculations were performed. X-ray molecular structures (when available) were used as a starting point for geometry optimizations.

Initial efforts were made to compare the trend in the rate constants with the energy barriers of the rate-determining step. To this end we selected the two most active systems, **3b** and **3g**, and the two of the least active ones, **3a** and **3f**. As shown in Fig. 5, the calculated energy barriers for the initial nucleophilic attack of piperidine to the indenyl fragment are 13.0 and 12.7 kcal mol^{-1} for **3b** and **3g** (high reactivity) and 13.8 and 15.4 for **3a** and **3f** (lower reactivity), respectively. This is in agreement with the experimentally observed trend of rate constants.

Having reproduced the experimental trends, we performed an analysis of the steric and electronic properties of the various systems to shed light on the structural parameters impacting most of the reactivity behavior. Focusing on steric features, a comparison of the $\%V_{\text{bur}}$ and the bite angle results in a strong correlation, $R^2 = 0.81$, for 11 Pd-indenyl complexes (**3a–3g** of Table 1, bearing two monodentate phosphines, and **4–8** of Table 2, bearing a bidentate phosphine and an indenyl moiety). This indicates that $\%V_{\text{bur}}$ and the bite angles can be interchanged as descriptors capturing steric pressure on the active center. As for electronic descriptors, we focused on the charge from natural population analysis (NPA) on the indenyl group, the Pd atom, and the ligand. In addition, we also considered the HOMO and LUMO energies.

Considering the small number of systems, 11, that mono- and bidentate phosphines are intrinsically different in flexibility and that the corresponding complexes contain different counterions, attempts to build a statistically robust regression model failed. We will thus provide qualitative analysis of trends for the **6** and **5** complexes bearing a mono- or bidentate phosphine. The R^2 values from univariate linear regression between the single descriptors and the experimental rate constants are reported in Table 3.

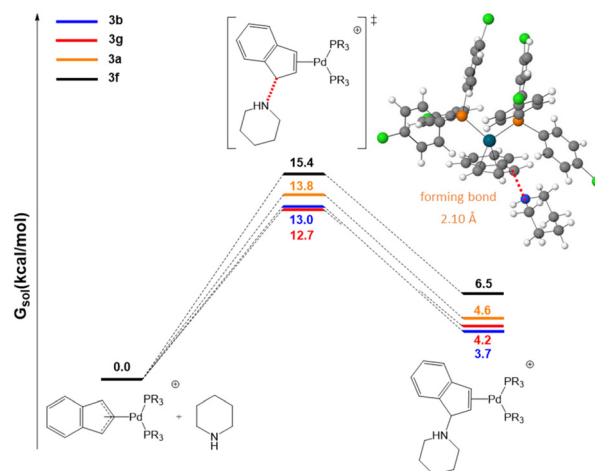


Fig. 5 Free energy profiles in chloroform for piperidine attack on the indenyl moiety. The inset shows the optimized geometry of transition state **3b**.



Table 3 R^2 values from univariate linear regression between the rate constants and a series of molecular descriptors for complexes having mono- and bidentate phosphines

Descriptor	Monodentate	Bidentate
% V_{Bur}	0.17	0.47
Bite angle	0.20	0.52
$E(\text{HOMO})$	0.70	0.09
$E(\text{LUMO})$	0.65	0.01
$E(\text{LUMO}) - E(\text{HOMO})$	0.02	0.09
NPA indenyl	0.59	0.54
NPA Pd	0.38	0.49
NPA Phosphine	0.35	0.66
NPA indenyl-Pd	0.76	0.01
NPA ind – (Pd + ligand)	0.63	0.54
E_{diss} (indenyl)	0.67	0.95

Starting with monodentate systems, poor correlation is observed between the two steric descriptors and the rate constants, with R^2 values of approximately 0.2. This poor correlation could either be due to the flexibility of mono-phosphines, impossible to capture with a single geometry, or the fact that four out of the six mono-phosphines differ by the nature of the *para* substituents. This should minimize differences in steric hindrance while impacting electronic properties. Indeed, a stronger correlation is observed between the rate constants and the HOMO and LUMO energies, as well as the NPA charges, especially of the indenyl ligand, with R^2 values of approximately 0.6–0.7. Higher activity corresponds to less electron-rich indenyls, or complexes having lower energy LUMOs, which would be more inclined to undergo nucleophilic attack. Consistently, the LUMO of **3b** reported in Fig. 6 is clearly localized on these C atoms. A strong correlation is also achieved using the difference between the indenyl and Pd NPA charges, which can be taken as a measure of the polarization of the Pd-indenyl interaction, with an R^2 value of 0.76. Higher rate constants correspond to more polarized Pd-indenyl bonds.

Moving to diphosphine based complexes, a reasonable correlation is observed between the steric descriptors and the rate

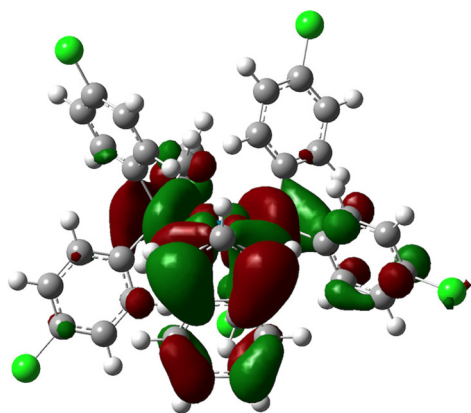


Fig. 6 Graphic representation of the LUMO of **3b**.

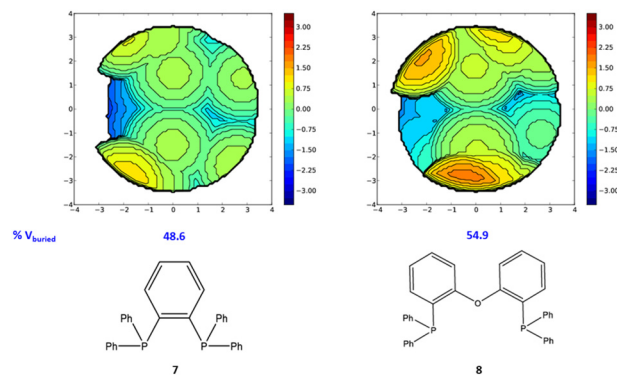


Fig. 7 Steric maps of complexes **7** and **8**.

constants, with R^2 values of approximately 0.5. The stronger impact of steric hindrance is in line with the clearly different steric requirements of the four bidentate phosphines, with larger % V_{Bur} and bite angles corresponding to higher rate constants. The topographic steric maps in Fig. 7 show how the hindrance in **8** results in a smaller reactive pocket compared to that of **7**. A reasonable correlation also results from using NPA charges as electronic descriptors, with R^2 values of approximately 0.5. A reasonable correlation is again calculated with the NPA charge on the indenyl moieties, with higher constant rates again corresponding to more electron deficient indenyls.

A comparison between R^2 values of the considered mono- and bidentate phosphines seems to indicate that the rate constants of complexes with monodentate phosphines are mostly determined by electronic features, whereas those of complexes with bidentate phosphines are determined by both steric and electronic features. Interestingly, the rate constants of both mono- and bidentate-based complexes correlate well with the indenyl dissociation energies. This can be understood considering that dissociation energies are influenced by both steric and electronic effects, and thus could be considered as a versatile descriptor.

Antiproliferative activity towards ovarian cancer and normal cells

At the end of this work, we propose potential for application of this class of compounds, a study of their antiproliferative activity towards cancer cells. In this regard, it is pointed out that in the literature only our recent contribution describes the behaviour of a particular class of Pd-indenyl derivatives in a biological environment,²⁶ and therefore this issue is worth further investigating. The tumour cell lines chosen for *in vitro* assays were ovarian cancer cells OVCAR-5, and A2780 and its cisplatin resistant clone A2780*cis*, all being particularly sensitive to treatment with organopalladium complexes. Our previous studies have highlighted the effectiveness of Pd-allyl derivatives, which present, as stated before, some common characteristics with Pd-indenyl substrates.¹⁷

The list of the tested compounds includes only those complexes that have shown no significant decomposition after



Table 4 Antiproliferative activity towards A2780, A2780*cis*, OVCAR-5 and MRC-5 cell lines

Compound	IC ₅₀ (μM)			
	A2780	A2780 <i>cis</i>	OVCAR-5	MRC-5
Cisplatin	0.9 ± 0.1	32 ± 9	1.3 ± 0.1	3.48 ± 0.09
3a	0.27 ± 0.02	0.26 ± 0.04	0.34 ± 0.08	2.5 ± 0.4
3f	0.26 ± 0.03	0.30 ± 0.04	0.48 ± 0.02	0.9 ± 0.2
3g	0.13 ± 0.05	3.0 ± 0.2	4.6 ± 0.5	2.27 ± 0.09
3h	0.73 ± 0.07	1.2 ± 0.2	1.9 ± 0.2	3.7 ± 0.3
4	0.122 ± 0.007	0.22 ± 0.07	0.10 ± 0.01	0.7 ± 0.1
5	0.12 ± 0.03	0.19 ± 0.08	0.075 ± 0.002	0.51 ± 0.03
6	0.18 ± 0.02	0.17 ± 0.03	0.13 ± 0.02	0.32 ± 0.03
7	0.062 ± 0.009	0.17 ± 0.05	0.15 ± 0.02	0.306 ± 0.003
8	0.29 ± 0.02	0.29 ± 0.07	0.336 ± 0.008	2.4 ± 0.4
9	0.020 ± 0.004	0.051 ± 0.008	0.03 ± 0.02	0.15 ± 0.06

Data after 96 h of incubation. Stock solutions in DMSO for all complexes; stock solutions in H₂O for cisplatin. A2780, cisplatin-sensitive ovarian cancer cells; A2780*cis*, cisplatin-resistant ovarian cancer cells; OVCAR-5, high-grade serous ovarian cancer cells; and MRC-5, normal lung fibroblasts.

24 hours in 1:1 DMSO-d⁶/D₂O solution. Unfortunately, all complexes with *para*-substituted aryl phosphines **3b–e** do not meet this requirement.

To this series of Pd-indenyl compounds we have also added the complex [Pd(DPPE)(η³-allyl)]OTf (**9**), in order to compare, on equal terms, the behaviours of Pd-indenyl and Pd-allyl fragments.

The cytotoxicity data of our compounds on the three ovarian cancer cell lines are summarised in Table 4, in terms of IC₅₀ values (half inhibitory concentrations). In the same table, data referred to a line of normal cells (MRC-5, human lung fibroblasts) are also reported for comparison. Moreover, all the tests have also been extended to cisplatin as a positive control.

On the basis of the obtained data, the following remarks may be proposed:

(a) All complexes show high antiproliferative activity towards all three types of ovarian cancer cells, with IC₅₀ values always significantly lower than cisplatin (often of about one order of magnitude in the cases of cisplatin sensitive A2780 and OVCAR-5 cell lines). In this respect, we emphasize that OVCAR-5 cells are high-grade serous ovarian cancer (HGSOC) cells, the most common and deadly form of the disease.

(b) With the only exception of the complex coordinating the trifurylphosphine (**3g**), all assayed Pd-indenyl derivatives exhibit the same degree of toxicity towards cisplatin-sensitive A2780 and cisplatin-resistant A2780*cis* cell lines. This suggests that our complexes might operate with a mechanism of action different from that of cisplatin (classical DNA metalation). Remarkably, the fact that their IC₅₀ values are lower by about two orders of magnitude than cisplatin qualifies these palladium compounds as potential metallodrugs toward forms of cancer resistant to cisplatin.

(c) The cytotoxicity of all diphosphine Pd-indenyl complexes against tumour cells appears basically independent of the nature of the phosphine used (monodentate or chelate). However, the complex coordinating only one phosphine ([Pd(indenyl)(XPhos)]ClO₄ (**3h**)) seems to be slightly less active.

(d) Unfortunately, in most cases our complexes have the same toxicity on tumour and normal cells (MRC-5). The only noteworthy exceptions have been represented by complexes [Pd(indenyl)(PPh₃)₂]ClO₄ (**3a**) and [Pd(DPEphos)(indenyl)]OTf (**8**) that show significant selectivity (*ca.* one order of magnitude) towards cancer cells.

(e) From the comparison between the IC₅₀ values of [Pd(DPPE)(indenyl)]OTf (**4**) and [Pd(DPPE)(η³-allyl)]OTf (**9**) it is apparent that the latter complex is more cytotoxic than the former on all tested cell lines. This could make us think that the susceptibility of the polyenyl fragment to undergo nucleophilic attack plays a key role in the mechanism of action of this kind of complex.

Conclusions

The results achieved in this work contribute to deepening the knowledge of palladium(II)-indenyl compounds, highlighting some of their features and potential. In particular, our attention has focused on cationic derivatives bearing phosphines as ancillary ligands. First, we have refined the synthetic protocols for the preparation of complexes with mono- and bidentate phosphines, providing detailed information on the structural and spectroscopic characteristics of the products. A characteristic common to all complexes is the hapticity of the indenyl fragment that is always intermediate between η³ and η⁵.

These compounds have proven to be ideal for a systematic study of nucleophilic attacks on coordinated-indenyl, an issue that had never been addressed to date. For the case in which the designed nucleophile is an amine, an in-depth kinetic analysis has allowed us to establish the rate law and the consequent reaction mechanism. DFT calculations have confirmed that the rate-determining step of the process is the initial nucleophilic attack of amine on the coordinated indenyl-fragment. This bimolecular step is favoured by the decrease of electron density on the indenyl-fragment and the steric hindrance of ligands. A systematic study, based on experimental



data, made it possible to select the most suitable steric and electronic descriptors to predict the susceptibility of the Pd-indenyl complexes to nucleophilic attack. Among steric descriptors, the possibility of using the bite angle of bidentate phosphines should be emphasised, in addition to the most predictable % V_{bur} .

An interesting and never before attempted application of these new compounds concerns their behaviour in the biological environment. As a matter of fact, a good number of the synthesized Pd-indenyl complexes have shown significant antiproliferative activity towards three different ovarian cancer cell lines. Their cytotoxicity is nearly always significantly higher than cisplatin, used as a positive control. Furthermore, with their effectiveness being basically the same on cisplatin-sensitive (A2780) and cisplatin-resistant (A2780*cis*) cells, it is reasonable to suppose that these novel complexes present a mechanism of action different from the classical DNA metalation of platinum chemotherapeutic drugs. Complexes **3a** and **8** deserve a special mention, coordinating PPh₃ and DPEphos respectively that exhibit significant selectivity towards tumor cells.

Experimental

Solvents and reagents

All syntheses were carried out under an inert atmosphere using standard Schlenk techniques. The solvent CH₂Cl₂ was distilled over P₂O₅ and stored under a N₂ atmosphere. CHCl₃ was distilled and stored in silver foil. All other solvents and chemicals were commercial grade products and used as purchased. [Pd(μ -Cl)(indenyl)]₂ and Pd(II)-allyl complex **9** were synthesized according to the published protocols.^{27,28}

NMR, UV-Vis and IR measurements

1D-NMR and 2D-NMR spectra were recorded on Bruker 300 or 400 Advance spectrometers. Chemical shift values (ppm) are given relative to TMS (¹H and ¹³C), H₃PO₄ (³¹P) and CCl₃F (¹⁹F).

IR spectra were recorded on a PerkinElmer Spectrum One spectrophotometer and UV-Vis spectra were recorded on a PerkinElmer Lambda 40 spectrophotometer equipped with a PerkinElmer PTP 6 (Peltier temperature programmer) apparatus.

Computational details

Geometries were optimized with the Gaussian09²⁹ package using the PBE0-D₃ functional.^{30–32} The electronic configuration of the system was described with the split-valence SVP basis set³³ for main group atoms (C, H, N, P, F, Cl and O) and the relativistic Stuttgart–Dresden effective core potential with the associated valence triple- ζ basis set for Pd and Fe.³⁴ All geometries were confirmed as the minimum or transition state through frequency calculations. The reported free energies were built through single point energy calculations on the PBE0-D₃ geometries using the M06 functional and the triple- ζ

TZVP basis set for main group atoms.^{35–37} Solvent effects were included with the PCM model using chloroform as the solvent.^{38,39} To this M06/TZVP electronic energy in solvent, thermal corrections were added from the gas-phase frequency calculations at the PBE0-D₃/SVP level.

Crystal structure determination

3b, **3h**, **4**, **5** and **6** crystal data were collected at the XRD1 and XRD2 beamlines of the Elettra Synchrotron, Trieste (Italy),⁴⁰ using a monochromatic wavelength of 0.620 Å, at 100 K (298K for **5**). The data sets were integrated, scaled and corrected for Lorentz, absorption and polarization effects using the XDS package.⁴¹ Data from two random orientations of the same crystals have been merged to obtain complete datasets for the triclinic **3h** crystal form, using the CCP4-Aimless code.^{42,43} The structures were solved by direct methods using the SHELXT program⁴⁴ and refined using full-matrix least-squares implemented in SHELXL-2018/3.⁴⁵ Thermal motions for all non-hydrogen atoms have been treated anisotropically and hydrogens have been included on calculated positions, riding on their carrier atoms. Geometric restraints (SAME, SADI, DFIX, DANG) have been applied to disordered fragments (triflate counterion in **5** and solvent in **3h**). The Coot program was used for structure building.⁴⁶ The crystal data are given in the ESI.† Pictures were prepared using Ortep⁴⁷ and Pymol⁴⁸ software. Crystallographic data have been deposited at the Cambridge Crystallographic Data Centre and allocated the deposition numbers CCDC 2173702, 2173705, 2173701, 2173703, and 2173704 for **3b**, **3h**, **4**, **5**, and **6**, respectively.†

Preliminary studies and kinetic measurements

All the reactions were firstly studied by ¹H and ³¹P{¹H} NMR spectroscopy by preparing a CDCl₃ solution of the selected complex at a concentration range of 0.008–0.010 mol dm⁻³ in the presence of dimethyl fumarate (0.012–0.015 mol dm⁻³), in order to stabilize the final Pd(0) complex formed after a nucleophilic attack at the indenyl fragment. An appropriate amount of piperidine (0.040–0.050 mol dm⁻³) was added and the reaction was followed monitoring the disappearance of the Pd(II) complex and the concomitant appearance of the Pd(0) olefin complex and the final organic product.

In the case of complex **3a** morpholine and diethylamine were also employed as nucleophilic agents.

The kinetic studies of complexes **3a**, **3b**, **3c**, **3e**, **3f**, **3g**, **3h**, **5**, and **8** were carried out by the UV-Vis technique. A series of preliminary tests were performed in order to evaluate the wavelength of the highest absorbance change. The complex of interest was dissolved in CHCl₃ ([complex] = 1 × 10⁻⁴–5.0 × 10⁻⁵ mol dm⁻³) in the presence of dimethyl fumarate (3.0 × 10⁻⁴–5.0 × 10⁻⁵ mol dm⁻³) and was placed in a thermostated cell compartment at 298 K in a spectrophotometer. An adequate excess of piperidine was added ([piperidine] ≥ 10 [complex]) and the reactions were monitored by recording spectra as a function of time at a wavelength interval of 366 to 600 nm. The kinetics of the nucleophile attack was studied by recording spectra with a fixed wavelength for each complex



evaluated from 378 to 398 nm under *pseudo*-first order conditions. To a solution of the complex in 3 mL of CHCl_3 ($[\text{complex}] = 10^{-4}$ – 5.0×10^{-5} mol dm^{-3}) in the presence of dimethyl fumarate (3.0×10^{-4} – 5.0×10^{-5} mol dm^{-3}), known aliquots of piperidine were added using fresh mother solution in CHCl_3 ($[\text{piperidine}] = 0.06$ – 0.14 mol dm^{-3}) (Fig. S80–S91†).

In the case of complexes **4**, **5**, and **7** the reactions were too slow for UV-Vis conditions. Therefore, the kinetics of the nucleophilic attack was studied by ^1H NMR spectroscopy following the integration change of one characteristic peak related to the initial complex. All spectra were recorded at 298 K in CDCl_3 where $[\text{complex}]_0 : [\text{dmfu}] : [\text{piperidine}]_0 = 0.01 : 0.015 : 0.04$ mol dm^{-3} (Fig. S92–S94†).

Synthesis of neutral Pd-indenyl complexes

[PdCl(indenyl)(PPh₃)] (2a). To 0.0500 g (0.0973 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})_2]$ dissolved in 7 mL of anhydrous CH_2Cl_2 , a solution of 0.0510 g (0.1945 mmol) of PPh_3 in 5 mL of anhydrous CH_2Cl_2 was added under an inert atmosphere (N_2). The resulting red solution was stirred for 15 min at room temperature and filtered through a small pad of Celite. From the concentrated dark solution, the title complex was precipitated by the addition of diethyl ether and dried under vacuum. 0.0935 g (yield 93%) of complex **2a** was obtained as a dark orange-red powder.

^1H NMR (300 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 4.58 (ddd, $J = 2.7, 2.0, 0.8$ Hz, 1H, H^3), 6.38 (d, $J = 7.5$ Hz, 1H, H^4), 6.48–6.54 (m, 1H, H^1), 6.71–6.74 (m, 1H, H^2), 6.87 (t, $J = 7.5$ Hz, 1H, H^5), 7.07 (t, $J = 7.5$ Hz, 1H, H^6), 7.26 (d, $J = 7.5$ Hz, 1H, H^7), 7.34–7.52 (m, 15H, Ar-H).

$^{31}\text{P}\{^1\text{H}\}$ NMR (121.5 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 27.5.

[PdCl(indenyl)(P(*p*-Cl-C₆H₄)₃)] (2b). A solution of 0.0718 g (0.1965 mmol) of tris(4-Cl-phenyl)phosphine in 5 mL of anhydrous CH_2Cl_2 was added under an inert atmosphere (N_2) to 0.0505 g (0.0982 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})_2]$ dissolved in 7 mL of anhydrous CH_2Cl_2 . The resulting red solution was stirred for 10 min at room temperature and filtered through a small pad of Celite. The solvent was removed under vacuum and a small amount of diethyl ether was added and the solution was dried at reduced pressure. 0.1204 g (yield 98%) of compound **2b** was obtained as a dark orange solid.

^1H NMR (300 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 4.63–4.65 (m, 1H, H^3), 6.44 (d, $J = 7.5$ Hz, 1H, H^4), 6.54–6.60 (m, 1H, H^1), 6.69–6.72 (m, 1H, H^2), 6.91 (t, $J = 7.5$ Hz, 1H, H^5), 7.10 (t, $J = 7.5$ Hz, 1H, H^6), 7.28 (d, $J = 7.5$ Hz, 1H, H^7), 7.42–7.31 (m, 12H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 80.3 (CH, d, $J_{\text{C-P}} = 3.8$ Hz, C^3), 97.7 (CH, d, $J_{\text{C-P}} = 23.2$ Hz, C^1), 111.4 (CH, d, $J_{\text{C-P}} = 5.8$ Hz, C^2), 116.5 (CH, C^4), 120.0 (CH, C^7), 126.8 (CH, C^5), 127.7 (CH, C^6), 129.3 (CH, d, $J_{\text{C-P}} = 11.5$ Hz, Ar-CH), 129.8 (C, d, $J_{\text{C-P}} = 45.6$ Hz, *ipso*-Ar-C), 134.8 (C, d, $J_{\text{C-P}} = 1.7$ Hz, C^{3a}), 135.1 (CH, d, $J_{\text{C-P}} = 13.6$ Hz, Ar-CH), 135.7 (C, d, $J_{\text{C-P}} = 4.9$ Hz, C^{7a}), 137.9 (C, d, $J_{\text{C-P}} = 2.6$ Hz, *p*-Ar-C).

$^{31}\text{P}\{^1\text{H}\}$ NMR (121.5 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 26.0.

[PdCl(indenyl)(P(*p*-F-C₆H₄)₃)] (2c). Complex **2c** was obtained in the same manner as complex **2b** by employing 0.0503 g (0.0978 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})_2]$ dissolved in 7 mL of

anhydrous CH_2Cl_2 and 0.0619 g (0.1956 mmol) of tris(4-F-phenyl)phosphine in 5 mL of anhydrous CH_2Cl_2 . 0.1085 g (yield 97%) of complex **2c** was obtained as a dark orange solid.

^1H NMR (300 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 4.64 (*pseudo* t, 1H, H^3), 6.44 (d, $J = 7.5$ Hz, 1H, H^4), 6.55–6.59 (m, 1H, H^1), 6.72–6.76 (m, 1H, H^2), 6.92 (t, $J = 7.5$ Hz, 1H, H^5), 7.12 (td, $J = 8.6, 1.6$ Hz, 7H, H^6 , Ar-H), 7.30 (d, $J = 7.5$ Hz, 1H, H^7), 7.42–7.50 (m, 6H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 79.8 (CH, d, $J_{\text{C-P}} = 3.7$ Hz, C^3), 97.3 (CH, d, $J_{\text{C-P}} = 23.6$ Hz, C^1), 111.3 (CH, d, $J_{\text{C-P}} = 5.9$ Hz, C^2), 116.1 (CH, dd, $J_{\text{C-F}}, \text{C-P} = 21.4, 12.0$ Hz, *m*-Ar-CH), 116.4 (CH, C^4), 119.8 (CH, C^7), 126.51 (CH, C^5), 127.3 (C, dd, $J_{\text{C-P}}, \text{C-F} = 47.6, 3.5$ Hz, *ipso*-Ar-C), 127.4 (CH, C^6), 134.7 (C, d, $J_{\text{C-P}} = 1.5$ Hz, C^{3a}), 135.7 (C, d, $J_{\text{C-P}} = 5.0$ Hz, C^{7a}), 135.8 (CH, dd, $J_{\text{C-P}}, \text{C-F} = 14.2, 8.5$ Hz, *o*-Ar-CH), 164.3 (C, dd, $J_{\text{C-F}}, \text{C-P} = 253.5, 2.5$ Hz, *p*-Ar-C).

$^{31}\text{P}\{^1\text{H}\}$ NMR (121.5 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 25.3 (q, $J_{\text{P-F}} = 2.5$ Hz).

$^{19}\text{F}\{^1\text{H}\}$ NMR (377.2 MHz, CDCl_3 , $T = 298$ K, ppm) δ : -108.0 (d, $J_{\text{F-P}} = 2.5$ Hz).

[PdCl(indenyl)(P(*p*-CF₃-C₆H₄)₃)] (2d). Complex **2d** was obtained in the same manner as complex **2b** by employing 0.0510 g (0.0992 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})_2]$ dissolved in 7 mL of anhydrous CH_2Cl_2 and 0.0925 g (0.1984 mmol) of tris(4-CF₃-phenyl)phosphine in 5 mL of anhydrous CH_2Cl_2 . 0.1370 g (yield 95%) of complex **2d** was obtained as a dark red solid.

^1H NMR (300 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 4.71 (*pseudo* t, 1H, H^3), 6.39 (d, $J = 7.4$ Hz, 1H, H^4), 6.60–6.67 (m, 1H, H^1), 6.72 (*pseudo* t, 1H, H^2), 6.92 (t, $J = 7.5$ Hz, 1H, H^5), 7.14 (t, $J = 7.5$ Hz, 1H, H^6), 7.31 (d, $J = 7.4$ Hz, 1H, H^7), 7.54–7.76 (m, 12H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (100.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 80.9 (CH, d, $J_{\text{C-P}} = 3.6$ Hz, C^3), 98.2 (CH, d, $J_{\text{C-P}} = 23.1$ Hz, C^1), 111.6 (CH, d, $J_{\text{C-P}} = 5.9$ Hz, C^2), 116.5 (CH, C^4), 120.2 (CH, C^7), 123.5 (C, d, $J_{\text{C-F}} = 272.8$ Hz, CF₃-C), 125.8–126.0 (CH, *o,m*-Ar-CH), 127.2 (CH, C^5), 128.1 (CH, C^6), 133.4 (C, dd, $J_{\text{C-P}}, \text{C-F} = 32.9, 2.6$ Hz, *p*-Ar-C), 134.3–134.6 (CH, *o,m*-Ar-CH), 134.8 (C, $J_{\text{C-P}} = 1.5$ Hz, C^{3a}), 135.2 (C, d, $J_{\text{C-F}} = 42.5$ Hz, *ipso*-Ar-C), 135.6 (C, d, $J_{\text{C-P}} = 5.0$ Hz, C^{7a}).

$^{31}\text{P}\{^1\text{H}\}$ NMR (121.5 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 28.0.

$^{19}\text{F}\{^1\text{H}\}$ NMR (377.2 MHz, CDCl_3 , $T = 298$ K, ppm) δ : -63.2.

[PdCl(indenyl)(P(*p*-OCH₃-C₆H₄)₃)] (2e). Complex **2e** was obtained in the same manner as complex **2b** by employing 0.0502 g (0.0976 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})_2]$ dissolved in 7 mL of anhydrous CH_2Cl_2 and 0.0688 g (0.1952 mmol) of tris(4-OCH₃-phenyl)phosphine in 5 mL of anhydrous CH_2Cl_2 . 0.1170 g (yield 98%) of complex **2e** was obtained as a dark orange solid.

^1H NMR (300 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 3.82 (s, 9H, OCH₃), 4.57 (*pseudo* t, 1H, H^3), 6.46 (d, $J = 7.8$ Hz, 2H, H^1, H^4), 6.72 (*pseudo* t, 1H, H^2), 6.89 (d, $J = 8.7$ Hz, 7H, H^5 , Ar-H), 7.07 (t, $J = 7.5$ Hz, 1H, H^6), 7.25 (d, $J = 7.8$ Hz, 1H, H^7), 7.38 (dd, $J = 11.3, 8.7$ Hz, 6H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (75.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 55.5 (CH₃, OCH₃), 78.9 (CH, d, $J_{\text{C-P}} = 3.9$ Hz, C^3), 96.9 (CH, d, $J_{\text{C-P}} =$



23.2 Hz, C¹), 111.2 (CH, d, $J_{C-P} = 5.9$ Hz, C²), 114.2 (CH, d, $J_{C-P} = 11.9$ Hz, *o*-Ar-CH), 116.8 (CH, C⁴), 119.6 (CH, C⁷), 123.7 (C, d, $J_{C-P} = 50.9$ Hz, *ipso*-Ar-C), 126.1 (CH, C⁵), 127.0 (CH, C⁶), 135.1 (C, d, $J_{C-P} = 1.9$ Hz, C^{3a}), 135.5 (CH, d, $J_{C-P} = 13.7$ Hz, *m*-Ar-CH), 136.2 (C, d, $J_{C-P} = 4.7$ Hz, C^{7a}), 161.5 (C, d, $J_{C-P} = 2.4$ Hz, *p*-Ar-C).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 23.5.

[PdCl(indenyl)(P(C₆H₄)₂(2-py))] (2f). Complex 2f was obtained in the same manner as complex 2b by employing 0.0502 g (0.0970 mmol) of [Pd(μ -Cl)(indenyl)]₂ dissolved in 7 mL of anhydrous CH₂Cl₂ and 0.0514 g (0.1940 mmol) of diphenyl-2-pyridylphosphine in 5 mL of anhydrous CH₂Cl₂. 0.0914 g (yield 90%) of complex 2f was obtained as a dark orange solid.

¹H NMR (300 MHz, CDCl₃, $T = 298$ K, ppm) δ : 4.82 (*pseudo* t, 1H, H³), 6.39 (d, $J = 7.5$ Hz, 1H, H⁴), 6.52–6.58 (m, 1H, H¹), 6.67–6.70 (m, 1H, H²), 6.84 (t, $J = 7.5$ Hz, 1H, H⁵), 7.06 (t, $J = 7.5$ Hz, 1H, H⁶), 7.25 (d, $J = 7.5$ Hz, 1H, H⁷), 7.28–7.74 (m, 13H, Ar-H, Py-H), 7.79 (d, $J = 4.2$ Hz, 1H, 6-Py-H).

¹³C{¹H} NMR (75.0 MHz, CDCl₃, $T = 298$ K, ppm, *selected peaks*) δ : 78.5 (CH, C³), 97.7 (CH, d, $J_{C-P} = 23.0$ Hz, C¹), 111.4 (CH, d, $J_{C-P} = 5.9$ Hz, C²), 116.9 (CH, C⁴), 119.6 (CH, C⁷), 124.3 (CH, d, $J_{C-P} = 2.4$ Hz, 3-Py-C), 126.4 (CH, C⁵), 127.2 (CH, C⁶), 128.5 (CH, d, $J_{C-P} = 10.8$ Hz, Ar-CH), 128.6 (CH, d, $J_{C-P} = 10.8$ Hz, Ar-CH), 130.8–131.5 (C, CH, Ar-C, Py-CH), 132.1 (C, C^{3a}), 134.2 (CH, d, $J_{C-P} = 12.1$ Hz, Ar-CH), 134.5 (CH, d, $J_{C-P} = 12.1$ Hz, Ar-CH), 134.9 (C, C^{7a}), 136.0 (CH, d, $J_{C-P} = 4.6$ Hz, Py-CH), 136.1 (CH, d, $J_{C-P} = 9.5$ Hz, Py-CH), 150.4 (CH, d, $J_{C-P} = 14.6$ Hz, 6-Py), 156.8 (C, d, $J_{C-P} = 65.4$ Hz, 2-Py).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 29.3.

[PdCl(indenyl)(P(2-furyl)₃)] (2g). Complex 2g was obtained in the same manner as complex 2a by employing 0.0503 g (0.0978 mmol) of [Pd(μ -Cl)(indenyl)]₂ dissolved in 7 mL of anhydrous CH₂Cl₂ and 0.0454 g (0.1957 mmol) of tris(2-furyl) phosphine in 5 mL of anhydrous CH₂Cl₂. 0.0910 g (yield 95%) of the title complex 2g was precipitated and isolated as a red powder and dried under vacuum.

¹H NMR (300 MHz, CDCl₃, $T = 298$ K, ppm) δ : 5.52 (*pseudo* t, 1H, H³), 6.49–6.44 (m, 3H, Furyl-H), 6.64 (bd, 1H, H¹), 6.73–6.68 (m, 2H, H², H⁴), 6.91 (t, $J = 7.5$ Hz, 1H, H⁵), 6.97 (t, $J = 2.9$ Hz, 3H, Furyl-H), 7.07 (t, $J = 7.5$ Hz, 1H, H⁶), 7.25 (d, $J = 6.5$ Hz, 1H, H₇), 7.71 (s, 3H, Furyl-H).

¹³C{¹H} NMR (75.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 76.2 (CH, d, $J_{C-P} = 3.6$ Hz, C³), 98.9 (CH, d, $J_{C-P} = 25.3$ Hz, C¹), 111.4 (CH, d, $J_{C-P} = 5.7$ Hz, C²), 111.6 (CH, d, $J_{C-P} = 8.7$ Hz, Furyl-CH), 118.3 (CH, C⁴), 119.8 (CH, C⁷), 124.7 (CH, d, $J_{C-P} = 23.3$ Hz, Furyl-CH), 126.8 (CH, C⁵), 127.8 (CH, C⁶), 135.2 (C, d, $J_{C-P} = 1.5$ Hz, C^{3a}), 136.3 (C, d, $J_{C-P} = 5.5$ Hz, C^{7a}), 143.5 (C, d, $J_{C-P} = 71.9$ Hz, Furyl-C), 148.7 (CH, d, $J_{C-P} = 5.3$ Hz, Furyl-CH).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : -28.5.

[PdCl(indenyl)(XPhos)] (2h). Complex 2h was obtained in the same manner as complex 2b by employing 0.0502 g (0.0630 mmol) of [Pd(μ -Cl)(indenyl)]₂ dissolved in 7 mL of anhydrous CH₂Cl₂ and 0.0601 g (0.1260 mmol) of XPhos in 5 mL of anhydrous CH₂Cl₂. 0.0894 g (yield 97%) of complex 2h was obtained as a red solid.

¹H NMR (300 MHz, CDCl₃, $T = 298$ K, ppm) δ : 0.94 (d, $J = 6.7$ Hz, 6H, ^{*i*}Pr-CH₃-H), 1.01 (d, $J = 6.7$ Hz, 6H, ^{*i*}Pr-CH₃-H), 0.86–1.12 (m, 2H, Cy-CH-H), 1.28 (dt, $J = 7.0, 4.2$ Hz, 6H, ^{*i*}Pr-CH₃-H), 1.23–1.50 (br, 16H, Cy-CH₂-H), 1.58 (br, 4H, Cy-CH₂-H), 2.46 (p, $J = 6.8$ Hz, 1H, ^{*i*}Pr-CH-H), 2.66 (p, $J = 6.8$ Hz, 1H, ^{*i*}Pr-CH-H), 2.93 (p, $J = 6.8$ Hz, 1H, ^{*i*}Pr-CH-H), 4.98 (bs, 1H, H³), 6.22–6.27 (m, 1H, H¹), 6.59–6.63 (m, 1H, H²), 6.83 (d, $J = 7.4$ Hz, H⁴), 6.91 (t, $J = 7.4$ Hz, H⁵), 6.95–7.10 (m, 4H, H⁶, Ar-H), 7.20 (d, $J = 7.4$ Hz, 1H, H⁷), 7.34–7.39 (m, 2H, Ar-H), 7.79–7.89 (m, 1H, Ar-H).

¹³C{¹H} NMR (75.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 22.4–28.2 (CH₃, CH₂, CH, ^{*i*}Pr-CH₃, Cy-CH₂, Cy-CH), 30.9 (CH, ^{*i*}Pr-CH), 31.0 (CH, ^{*i*}Pr-CH), 34.6 (CH, ^{*i*}Pr-CH), 73.3 (CH, d, $J_{C-P} = 3.7$ Hz, C³), 94.8 (CH, d, $J_{C-P} = 22.5$ Hz, C¹), 111.5 (CH, d, $J_{C-P} = 6.0$ Hz, C²), 117.2 (CH, C⁴), 119.5 (CH, C⁷), 121.2 (CH, d, $J_{C-P} = 3.9$ Hz, Ar-CH), 125.3 (CH, C⁵), 126.0 (CH, $J_{C-P} = 13.8$ Hz, Ar-CH), 126.8 (CH, C⁶), 128.5 (CH, d, $J_{C-P} = 2.4$ Hz, Ar-CH), 132.3 (C, d, $J_{C-P} = 27.1$ Hz, *ipso*-Ar-C), 133.8 (CH, d, $J_{C-P} = 6.4$ Hz, Ar-CH), 136.4 (C, d, $J_{C-P} = 1.5$ Hz, C^{3a}), 136.5–136.6 (C, Ar-C), 137.4 (C, d, $J_{C-P} = 4.7$ Hz, C^{7a}), 137.6–138.1 (CH, Ar-CH), 141.4 (C, Ar-C), 146.4 (C, d, $J_{C-P} = 7.3$ Hz, Ar-C), 149.2 (C, Ar-C).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 57.9.

All neutral complexes [PdCl(indenyl)(PAR₃)] isolated contain always small traces of the respective Pd(II) dimers [(μ - η^3 -ind)(μ -Cl)Pd₂(PAR₃)₂], and for this reason the elemental analyses were not helpful for their characterization.

Synthesis of cationic Pd-indenyl complexes

Complexes with monodentate phosphines. For the synthesis of cationic complexes with monodentate phosphines two consecutive steps were required. The first one is the synthesis of neutral compounds described above and the second one is here reported for each complex. In this case the isolation of the neutral compound was not necessary.

[Pd(indenyl)(PPh₃)₂]ClO₄ (3a). The neutral complex 2a was obtained by the reaction between 0.0200 g (0.0389 mmol) of [Pd(μ -Cl)(indenyl)]₂ and 0.0204 g (0.0778 mmol) of PPh₃ using 4 mL of anhydrous CH₂Cl₂. 0.0208 g (0.1483 mmol) of NaClO₄·H₂O dissolved in 2 mL of methanol and a solution of 0.0204 g (0.0778 mmol) of PPh₃ in 2 mL of CH₂Cl₂ (CH₂Cl₂/MeOH \approx 3/1) were added. The mixture was stirred at room temperature for 15 min and the solvent was removed at low pressure. 5 mL of CH₂Cl₂ was added, the mixture was filtered on Millipore apparatus and the solution was concentrated under vacuum. The title complex is precipitated by the addition of diethyl ether and dried under vacuum. 0.0600 g (yield 96%) was obtained as a dark orange powder.

¹H NMR (400 MHz, CDCl₃, $T = 298$ K, ppm) δ : 5.48–5.51 (m, 2H, H¹, H³), 6.18–6.28 (m, 2H, H⁴, H⁷), 7.06–7.12 (m, 14H, H⁵, H⁶, Ar-H), 7.30 (t, $J = 7.3$ Hz, 13H, H², Ar-H), 7.21 (t, $J = 7.4$ Hz, 6H, *p*-Ar-H).

¹³C{¹H} NMR (100.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 95.6 (CH, t, $J_{C-P} = 11.7$ Hz, C¹, C³), 114.0 (CH, t, $J_{C-P} = 5.3$ Hz, C²), 118.9 (CH, C⁴, C⁷), 127.9 (CH, C⁵, C⁶), 129.1 (CH, t, $J_{C-P} = 5.8$ Hz, Ar-CH), 130.2 (C, dd, $J_{C-P} = 24.0, 23.1$ Hz, *ipso*-Ar-C), 131.4



(CH, *p*-Ar-CH), 132.0 (C, t, $J_{C-P} = 3.2$ Hz, C^{3a}, C^{7a}), 133.6 (CH, t, $J_{C-P} = 6.1$ Hz, Ar-CH).

³¹P{¹H} NMR (162.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 26.4.

IR (KBr pellet, cm⁻¹): $\nu_{ClO} = 1091$, $\delta_{ClO} = 623$.

Elemental analysis calcd (%) for C₄₅H₃₇ClO₄P₂Pd: C, 63.92, H, 4.41; found: C, 63.64, H, 4.58.

HRMS calcd for [C₄₅H₃₇P₂Pd]⁺: 745.1405; found: 745.1440.

[Pd(*indenyl*)(P(*p*-Cl-C₆H₄)₃)₂]ClO₄ (**3b**). The neutral complex **2b** was obtained by the reaction between 0.0504 g (0.0980 mmol) of [Pd(μ -Cl)(*indenyl*)₂] and 0.0717 g (0.1961 mmol) of tris(4-Cl-phenyl)phosphine using 5 mL of anhydrous CH₂Cl₂. The cationic complex **3b** was obtained in the same manner as complex **3a** using 0.0551 g (0.3922 mmol) of NaClO₄·H₂O dissolved in 3 mL of methanol and a solution of 0.0717 g (0.1961 mmol) of tris(4-Cl-phenyl)phosphine in 4 mL of CH₂Cl₂. 0.1733 g (yield 84%) of complex **3b** was obtained as a pale orange powder.

¹H NMR (400 MHz, CDCl₃, $T = 298$ K, ppm) δ : 5.50–5.53 (m, 2H, H¹, H³), 6.47–6.49 (m, 2H, H⁴, H⁷), 7.03–7.09 (m, 12H, *o*-Ar-H), 7.14–7.17 (m, 2H, H⁵, H⁶), 7.33 (d, $J = 8.0$ Hz, 12H, *m*-Ar-H), 7.56 (t, $J = 3.3$ Hz, H²).

¹³C{¹H} NMR (75.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 97.0 (CH, t, $J_{C-P} = 11.7$ Hz, C¹, C³), 116.2 (CH, t, $J_{C-P} = 5.6$ Hz C²), 118.9 (CH, C⁴, C⁷), 128.0 (C, dd, $J_{C-P} = 23.8$, 23.7 Hz, *ipso*-Ar-C), 128.2 (CH, C⁵, C⁶), 129.7 (CH, t, $J_{C-P} = 5.8$ Hz, *m*-Ar-CH), 132.3 (C, C^{3a}, C^{7a}), 134.7 (CH, t, $J_{C-P} = 6.7$ Hz, *o*-Ar-CH), 138.8 (C, *p*-Ar-C).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 24.4.

IR (KBr pellet, cm⁻¹): $\nu_{ClO} = 1089$, $\delta_{ClO} = 622$.

Elemental analysis calcd (%) for C₄₅H₃₁Cl₇O₄P₂Pd: C, 51.37, H, 2.97; found: C, 51.69, H, 2.80.

HRMS calcd for [C₄₅H₃₁Cl₆P₂Pd]⁺: 952.9042; found: 952.9018.

[Pd(*indenyl*)(P(*p*-F-C₆H₄)₃)₂(*indenyl*)]ClO₄ (**3c**). The neutral complex **2c** was obtained by the reaction between 0.0507 g (0.0986 mmol) of [Pd(μ -Cl)(*indenyl*)₂] and 0.0624 g (0.1972 mmol) of tris(4-F-phenyl)phosphine using 6 mL of anhydrous CH₂Cl₂. The cationic complex **3c** was obtained in the same way as complex **3a** using 0.0554 g (0.3944 mmol) of NaClO₄·H₂O dissolved in 3 mL of methanol and a solution of 0.0624 g (0.1972 mmol) of tris(4-F-phenyl)phosphine in 3 mL of CH₂Cl₂. 0.1580 g (yield 80%) of complex **3c** was obtained as a pale orange powder.

¹H NMR (300 MHz, CDCl₃, $T = 298$ K, ppm) δ : 5.49–5.53 (m, 2H, H¹, H³), 6.41–6.44 (m, 2H, H⁴, H⁷), 7.00–7.21 (m, 26H, H⁵, H⁶, Ar-H) 7.54 (t, $J = 3.3$ Hz, H²).

¹³C{¹H} NMR (75.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 96.4 (CH, t, $J_{C-P} = 11.9$ Hz, C¹, C³), 116.0 (CH, d, $J_{C-P} = 4.6$ Hz C²), 116.8 (CH, td, J_{C-F} , $J_{C-P} = 21.6$, 6.2 Hz, *m*-Ar-CH), 118.9 (CH, C⁴, C⁷), 125.9 (C, dd, J_{C-P} , $J_{C-F} = 26.4$, 24.0, 3.4 Hz, *ipso*-Ar-C), 128.0 (CH, C⁵, C⁶), 132.3 (C, t, $J_{C-P} = 3.2$ Hz, C^{3a}, C^{7a}), 135.8 (CH, dt, $J_{C-P} = 7.6$ Hz, $J_{C-F} = 7.6$ Hz, *o*-Ar-CH), 164.6 (C, d, $J_{C-F} = 255.8$ Hz, *p*-Ar-C).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 24.0.

¹⁹F{¹H} NMR (377.2 MHz, CDCl₃, $T = 298$ K, ppm) δ : -106.0

IR (KBr pellet, cm⁻¹): $\nu_{ClO} = 1091$, $\delta_{ClO} = 623$.

Elemental analysis calcd (%) for C₄₅H₃₁ClF₆O₄P₂Pd: C, 56.68, H, 3.28; found: C, 57.01, H, 3.10.

HRMS calcd for [C₄₅H₃₁F₆P₂Pd]⁺: 853.0840; found: 853.0895.

[Pd(*indenyl*)(P(*p*-OCH₃-C₆H₄)₃)₂]ClO₄ (**3e**). The neutral complex **2e** was obtained by the reaction between 0.0495 g (0.0962 mmol) of [Pd(μ -Cl)(*indenyl*)₂] and 0.0679 g (0.1925 mmol) of tris(4-OCH₃-phenyl)phosphine using 4 mL of anhydrous CH₂Cl₂. The cationic complex **3e** was obtained in the same way as complex **3a** using 0.0541 g (0.3848 mmol) of NaClO₄·H₂O dissolved in 3 mL of methanol and a solution of 0.0679 g (0.1925 mmol) of tris(4-OCH₃-phenyl)phosphine in 5 mL of CH₂Cl₂. 0.1940 g (yield 98%) of complex **3e** were obtained as a pale orange powder.

¹H NMR (300 MHz, CDCl₃, $T = 298$ K, ppm) δ : 3.83 (s, 18H, *o*-CH₃-H), 5.37–5.41 (m, 2H, H¹, H³), 6.41–6.44 (m, 2H, H⁴, H⁷), 6.80 (d, $J = 8.1$ Hz, 12H, Ar-H), 6.90–7.00 (m, 12H, Ar-H), 7.11–7.15 (m, 3H, H², H⁵, H⁶).

¹³C{¹H} NMR (75.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 55.7 (CH, OCH₃), 94.5 (CH, t, $J_{C-P} = 12.0$ Hz, C¹, C³), 113.3 (CH, C²), 114.5 (CH, t, $J_{C-P} = 6.0$ Hz, *o*, *m*-Ar-CH), 118.9 (CH, C⁴, C⁷), 122.0 (C, dd, $J_{C-P} = 26.7$, 26.4 Hz, *ipso*-Ar-C), 127.4 (CH, C⁵, C⁶), 132.0 (C, t, $J_{C-P} = 3.1$ Hz, C^{3a}, C^{7a}), 135.1 (CH, t, $J_{C-P} = 6.9$ Hz, *o*, *m*-Ar-CH), 161.8 (C, *p*-Ar-C).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 23.1.

IR (KBr pellet, cm⁻¹): $\nu_{ClO} = 1090$, $\delta_{ClO} = 623$.

Elemental analysis calcd (%) for C₄₅H₄₉ClO₁₀P₂Pd: C, 59.72, H, 4.82; found: C, 59.38, H, 5.04.

HRMS calcd for [C₅₁H₄₉O₆P₂Pd]⁺: 925.2039; found: 925.2135.

[Pd(*indenyl*)(P(Ph)₂(2-py))₂]ClO₄ (**3f**). The neutral complex **2f** was obtained by the reaction between 0.0503 g (0.0978 mmol) of [Pd(μ -Cl)(*indenyl*)₂] and 0.0515 g (0.1957 mmol) of (diphenyl-2-pyridyl)phosphine using 5 mL of anhydrous CH₂Cl₂. The cationic complex **3f** was obtained in the same way as complex **3a** using 0.0550 g (0.3914 mmol) of NaClO₄·H₂O dissolved in 3 mL of methanol and a solution of 0.0515 g (0.1957 mmol) of (diphenyl-2-pyridyl)phosphine in 4 mL of CH₂Cl₂. 0.1605 g (yield 97%) of complex **3f** was obtained as a pale orange powder.

¹H NMR (300 MHz, CDCl₃, $T = 298$ K, ppm) δ : 5.57–6.62 (m, 2H, H¹, H³), 6.19–6.22 (m, 2H, H⁴, H⁷), 6.87–6.98 (m, 3H, H², Py-H), 7.01–7.05 (m, 2H, H⁵, H⁶), 7.07–7.54 (m, 24H, Ar-H, Py-H), 8.25 (d, $J = 4.4$ Hz, 2H, 6-Py-H).

¹³C{¹H} NMR (100.0 MHz, CDCl₃, $T = 298$ K, ppm) δ : 94.6 (CH, t, $J_{C-P} = 11.6$ Hz, C¹, C³), 112.0 (CH, t, $J_{C-P} = 5.2$ Hz C²), 118.7 (CH, C⁴, C⁷), 124.7 (CH, Py-CH), 128.0 (CH, C⁵, C⁶), 128.1–136.7 (CH, C, Ar-CH, Ar-C, Py-CH, Py-C), 131.1 (C, t, $J_{C-P} = 3.2$ Hz, C^{3a}, C^{7a}), 150.2 (CH, t, $J_{C-P} = 9.0$ Hz, 6-Py-CH), 154.7–156.3 (CH, C, Ar-CH, Ar-C, Py-CH, Py-C).

³¹P{¹H} NMR (121.5 MHz, CDCl₃, $T = 298$ K, ppm) δ : 27.5.

IR (KBr pellet, cm⁻¹): $\nu_{ClO} = 1093$, $\delta_{ClO} = 620$.

Elemental analysis calcd (%) for C₄₃H₃₅ClN₂O₄P₂Pd: C, 60.93, H, 4.16, N, 3.31; found: C, 60.65, H, 4.32, N, 3.57.

HRMS calcd for [C₄₃H₃₂N₂P₂Pd]⁺: 747.1310; found: 747.1315.



$[Pd(indenyl)(P(2-furyl)_3)_2]ClO_4$ (**3g**). The neutral complex **2g** was obtained by the reaction between 0.0504 g (0.0980 mmol) of $[Pd(\mu-Cl)(indenyl)]_2$ and 0.0455 g (0.1961 mmol) of tris(2-furyl)phosphine using 4 mL of anhydrous CH_2Cl_2 . The cationic complex **3g** was obtained in the same way as complex **3a** using 0.0551 g (0.3992 mmol) of $NaClO_4 \cdot H_2O$ dissolved in 3 mL of methanol and a solution of 0.0455 g (0.1961 mmol) of tris(2-furyl)phosphine in 5 mL of CH_2Cl_2 . 0.1142 g (yield 74%) of complex **3g** was obtained as a pale brown powder.

1H NMR (400 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 6.47 (dd, $J = 3.5, 1.7$ Hz, 6H, Furyl-H), 6.50–6.58 (m, 8H, H^1, H^3 , Furyl-H), 6.73–6.81 (m, 2H, H^4, H^7), 6.83 (t, $J = 3.2$ Hz, 1H, H^2), 7.15–7.17 (m, 2H, H^5, H^6), 7.61–7.63 (m, 6H, Furyl-H).

$^{13}C\{^1H\}$ NMR (100 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 93.9 (CH, t, $J_{C-P} = 12.1$ Hz, C^1, C^3), 111.1 (CH, t, $J_{C-P} = 6.3$ Hz, C^2), 112.1 (CH, t, $J_{C-P} = 4.0$ Hz, Furyl-C), 120.1 (CH, C^4, C^7), 124.2 (CH, t, $J_{C-P} = 9.7$ Hz, Furyl-C), 128.6 (CH, C^5, C^6), 131.1 (C, t, $J_{C-P} = 3.7$ Hz, C^{3a}, C^{7a}), 140.9–142.6 (C, Furyl-C), 149.5 (CH, t, $J_{C-P} = 3.2$ Hz, Furyl-C).

$^{31}P\{^1H\}$ NMR (121.5 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : -27.1.

IR (KBr pellet, cm^{-1}): $\nu_{ClO} = 1108, \delta_{ClO} = 622$.

Elemental analysis calcd (%) for $C_{33}H_{25}ClO_{10}P_2Pd$: C, 50.47, H, 3.21; found: C, 50.80, H, 3.01.

HRMS calcd for $[C_{33}H_{25}O_6P_2Pd]^+$: 685.0161; found: 685.0206.

$[Pd(indenyl)(XPhos)]ClO_4$ (**3h**). The neutral complex **2h** was obtained by the reaction between 0.0299 g (0.0581 mmol) of $[Pd(\mu-Cl)(indenyl)]_2$ and 0.0555 g (0.1162 mmol) of XPhos using 4 mL of anhydrous CH_2Cl_2 and after 15 min 0.0327 g (0.2328 mmol) of $NaClO_4 \cdot H_2O$ dissolved in 2 mL of methanol was added. The mixture was stirred at room temperature for 15 min and the solvents were removed at low pressure. 5 mL of CH_2Cl_2 was added and the mixture was filtered on Millipore apparatus and the solution was concentrated under vacuum. The title complex is precipitated by the addition of diethyl ether and *n*-pentane and dried under vacuum. 0.0880 g (yield 95%) of complex **3g** was obtained as an orange powder.

1H NMR (400 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 0.55 (d, $J = 6.7$ Hz, 3H, $^iPr-CH_3$), 0.76 (d, $J = 6.7$ Hz, 3H, $^iPr-CH_3$), 0.86 (d, $J = 6.7$ Hz, 3H, $^iPr-CH_3$), 1.13–1.45 (m, 8H, Cy- CH_2), 1.46–1.50 (m, 8H, $^iPr-CH_3$, Cy- CH_2), 1.54 (d, $J = 6.9$ Hz, 3H, $^iPr-CH_3$), 1.65–1.98 (m, 8H, Cy- CH_2), 2.08 (p, $J = 6.7$ Hz, 1H, ^iPr-CH), 2.11–2.25 (m, 3H, ^iPr-CH , Cy- CH_2), 2.33–2.42 (m, 1H, Cy- CH), 2.48–2.57 (m, 1H, Cy- CH), 3.18 (p, $J = 6.9$ Hz, 1H, ^iPr-CH), 4.58–4.63 (m, 1H, H^3), 6.23–6.24 (m, 1H, H^1), 6.62–6.64 (m, 1H, H^2), 6.69 (dd, $J = 7.3, 2.7$ Hz, 1H, Ar-H), 6.83 (d, $J = 7.5$ Hz, 1H, H^7), 6.96 (t, $J = 7.5$ Hz, 1H, H^5), 7.08 (t, $J = 7.5$ Hz, 1H, H^6), 7.20 (s, 1H, Ar-H), 7.28 (d, $J = 7.5$ Hz, 1H, H^4), 7.44 (tt, $J = 7.5, 1.7$ Hz, 1H, Ar-H), 7.50 (s, 1H, Ar-H), 7.53 (tt, $J = 7.5, 1.7$ Hz, 1H, Ar-H), 7.70 (t, $J = 7.1$ Hz, 1H, Ar-H).

$^{13}C\{^1H\}$ NMR (100.0 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 23.6–25.9 (CH, $^iPr-CH_3$), 26.0–30.2 (CH₂, Cy- CH_2), 32.0–34.3 (CH, ^iPr-CH), 37.3 (CH, d, $J_{C-P} = 18.9$ Hz, CH-Cy), 37.6 (CH, d, $J_{C-P} = 21.4$ Hz, CH-Cy), 71.1 (CH, d, $J_{C-P} = 3.3$ Hz, C^3), 113.8 (CH, d, $J_{C-P} = 6.9$ Hz, C^2), 116.9 (CH, d, $J_{C-P} = 21.4$ Hz, C^1), 118.3 (C, d, $J_{C-P} = 4.4$ Hz, Ar-C), 119.4 (CH, C^7), 121.0 (CH, C^4),

125.8 (CH, Ar-CH), 126.4 (CH, Ar-CH), 127.6 (CH, C^5), 128.9 (CH, d, $J_{C-P} = 5.5$ Hz, Ar-CH), 129.3 (CH, C^6), 131.8–132.4 (CH, Ar-CH), 132.9 (C, d, $J_{C-P} = 38.9$ Hz, Ar-C), 134.8 (C, d, $J_{C-P} = 2.2$ Hz, C^{3a}), 138.8 (C, d, $J_{C-P} = 5.1$ Hz, C^{7a}), 145.1 (C, d, $J_{C-P} = 20.3$ Hz, Ar-C), 146.0 (C, Ar-C), 150.3 (C, Ar-C), 152.3 (C, Ar-C).

$^{31}P\{^1H\}$ NMR (121.5 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 54.9.

IR (KBr pellet, cm^{-1}): $\nu_{ClO} = 1089, \delta_{ClO} = 623$.

Elemental analysis calcd (%) for $C_{42}H_{56}ClO_4PPd$: C, 63.24, H, 7.08; found: C, 62.87, H, 7.26.

HRMS calcd for $[C_{42}H_{56}PPd]^+$: 697.3154; found: 697.3209.

Complexes with bidentate phosphines

$[Pd(DPPE)(indenyl)]OTf$ (**4**). To 0.0503 g (0.0978 mmol) of $[Pd(\mu-Cl)(indenyl)]_2$ dissolved in 15 mL of anhydrous CH_2Cl_2 , 0.0528 mg (0.2054 mmol) of AgOTf were added and the mixture was stirred at room temperature for 5 min. A solution of 0.0779 g (0.1956 mmol) of 1,2-bis(diphenylphosphino)ethane in 5 mL of anhydrous CH_2Cl_2 was added under an inert atmosphere (N_2). The resulting red solution was stirred for 30 min at room temperature and filtered through a small pad of Celite to remove the AgCl. From the concentrated dark solution, the title complex was precipitated by the addition of diethyl ether and dried under vacuum. 0.1200 g (yield 80%) was obtained as an orange powder.

1H NMR (300 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 2.50 (d, $J = 19.1$ Hz, 4H, PCH_2), 6.33 (*pseudo* q, 2H, H^1, H^3), 6.60 (t, $J = 3.2$ Hz, 1H, H^2), 6.63–6.70 (m, 2H, H^4, H^7), 7.03–7.11 (m, 2H, H^5, H^6), 6.80–7.80 (bm, 20H, Ar-H).

$^{13}C\{^1H\}$ NMR (75.0 MHz, $CDCl_3$, $T = 298$ K, ppm, *selected peaks*) δ : 28.0 (CH₂, t, $J_{C-P} = 24.3$ Hz, PCH_2), 88.4 (CH, t, $J_{C-P} = 11.9$ Hz, C^1, C^3), 109.5 (CH, bt, C^2), 118.9 (CH, C^4, C^7), 127.1 (CH, C^5, C^6), 128.9 (C, C^{3a}, C^{7a}).

$^{31}P\{^1H\}$ NMR (121.5 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 61.0.

$^{19}F\{^1H\}$ NMR (377.2 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : -78.1.

IR (KBr pellet, cm^{-1}): $\nu_{CF} = 1230, \nu_{SO} = 1026, \delta_{SO} = 633$.

Elemental analysis calcd (%) for $C_{36}H_{31}F_3O_3P_2PdS$: C, 56.22, H, 4.06, S, 4.17; found: C, 56.60, H, 3.91, S, 4.34.

HRMS calcd for $[C_{42}H_{56}P_2Pd]^+$: 619.0936; found: 619.0972.

$[Pd(DPPP)(indenyl)]OTf$ (**5**). Complex **5** was obtained by the same synthetic route of complex **4** using 0.0301 g (0.0586 mmol) of $[Pd(\mu-Cl)(indenyl)]_2$ dissolved in 10 mL of anhydrous CH_2Cl_2 , 0.0316 mg (0.1230 mmol) of AgOTf and a solution of 0.0483 g (0.1172 mmol) of 1,3-bis(diphenylphosphino)propane in 3 mL of anhydrous CH_2Cl_2 . 0.0777 g (yield 85%) was obtained as an orange powder.

1H NMR (400 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 1.50–3.00 (bm, 6H, CH_2), 5.29 (*pseudo* q, 2H, H^1, H^3), 6.81 (t, $J = 3.1$ Hz, 1H, H^2), 6.82–6.86 (m, 2H, H^4, H^7), 6.94 (bs, 4H, Ar-H), 7.13–7.19 (m, 2H, H^5, H^6), 7.33–7.60 (bm, 16H, Ar-H).

$^{13}C\{^1H\}$ NMR (75.0 MHz, $CDCl_3$, $T = 298$ K, ppm, *selected peaks*) δ : 18.5 (CH₂, CH_2), 24.6 (CH₂, t, $J_{C-P} = 17.5$ Hz, PCH_2), 92.5 (CH, t, $J_{C-P} = 12.0$ Hz, C^1, C^3), 111.6 (CH, t, $J_{C-P} = 5.8$ Hz, C^2), 118.5 (CH, C^4, C^7), 126.5 (CH, C^5, C^6), 129.2 (CH, bs, Ar-CH), 129.8 (C, t, $J_{C-P} = 5.8$ Hz, C^{3a}, C^{7a}).

$^{31}P\{^1H\}$ NMR (162.0 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : 8.2.

$^{19}F\{^1H\}$ NMR (377.2 MHz, $CDCl_3$, $T = 298$ K, ppm) δ : -78.0.



IR (KBr pellet, cm^{-1}): $\nu_{\text{CF}} = 1221$, $\nu_{\text{SO}} = 1152$, $\delta_{\text{SO}} = 635$, 512.

Elemental analysis calcd (%) for $\text{C}_{37}\text{H}_{33}\text{F}_3\text{O}_3\text{P}_2\text{PdS}$: C, 56.75, H, 4.25, S, 4.09; found: C, 56.40, H, 4.43, S, 4.48.

HRMS calcd for $[\text{C}_{36}\text{H}_{33}\text{P}_2\text{Pd}]^{\dagger}$: 633.1092; found: 633.1133.

$[\text{Pd}(\text{DPPF})(\text{indenyl})]\text{OTf}$ (6). Complex 6 was obtained by the same synthetic route of complex 4 using 0.0301 g (0.0586 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})]_2$ dissolved in 9 mL of anhydrous CH_2Cl_2 , 0.0316 mg (0.1230 mmol) of AgOTf and a solution of 0.0649 g (0.1172 mmol) of 1,1'-ferrocenediyl-bis(diphenylphosphine) in 2 mL of anhydrous CH_2Cl_2 . 0.0777 g (yield 90%) was obtained as a red powder.

^1H NMR (400 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 4.03 (s, 2H, Fc-H), 4.22 (s, 2H, Fc-H), 4.43 (d, $J = 15.8$ Hz, 4H, Fc-H), 5.45 (*pseudo* q, 2H, H^1 , H^3), 6.16–6.22 (m, 2H, H^4 , H^7), 6.95 (t, $J = 3.1$ Hz, 1H, H^2), 7.00–7.05 (m, 2H, H^5 , H^6), 7.15–7.26 (m, 4H, Ar-H), 7.52 (t, $J = 7.5$ Hz, 4H, *p*-Ar-H), 7.58–7.68 (m, 12H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (100.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 72.5 (C, dd, $J_{\text{C-P}} = 31.4$, 30.2 Hz, Fc-C), 74.2 (CH, t, $J_{\text{C-P}} = 3.9$ Hz, Fc-CH), 74.4 (CH, t, $J_{\text{C-P}} = 3.7$ Hz, Fc-CH), 76.1 (CH, dt, $J_{\text{C-P}} = 12.0$, 5.8 Hz, Fc-CH), 94.2 (CH, t, $J_{\text{C-P}} = 11.8$ Hz, C^1 , C^3), 112.5 (CH, t, $J_{\text{C-P}} = 5.4$ Hz, C^2), 118.5 (CH, C^4 , C^7), 127.9 (CH, C^5 , C^6), 129.2 (CH, t, $J_{\text{C-P}} = 5.5$ Hz, Ar-CH), 129.7 (CH, t, $J_{\text{C-P}} = 5.5$ Hz, Ar-CH), 130.5 (C, t, $J_{\text{C-P}} = 3.1$ Hz, C^{3a} , C^{7a}), 131.6 (C, Ar-C), 132.4–133.5 (CH, Ar-CH), 134.1 (CH, $J_{\text{C-P}} = 6.6$ Hz, Ar-CH).

$^{31}\text{P}\{^1\text{H}\}$ NMR (162.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 30.1.

$^{19}\text{F}\{^1\text{H}\}$ NMR (377.2 MHz, CDCl_3 , $T = 298$ K, ppm) δ : -77.9.

IR (KBr pellet, cm^{-1}): $\nu_{\text{SO}} = 1312$, $\nu_{\text{CF}} = 1224$, 1160, $\nu_{\text{SO}} = 1031$, $\delta_{\text{SO}} = 637$.

Elemental analysis calcd (%) for $\text{C}_{44}\text{H}_{35}\text{F}_3\text{FeO}_3\text{P}_2\text{PdS}$: C, 57.13, H, 3.81, S, 3.47; found: C, 56.84, H, 3.98, S, 3.80.

HRMS calcd for $[\text{C}_{43}\text{H}_{35}\text{FeP}_2\text{Pd}]^{\dagger}$: 775.0598; found: 775.0649.

$[\text{Pd}(\text{DPPBz})(\text{indenyl})]\text{OTf}$ (7). Complex 7 was obtained by the same synthetic route of complex 4 using 0.0319 g (0.0621 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})]_2$ dissolved in 10 mL of anhydrous CH_2Cl_2 , 0.0335 mg (0.1304 mmol) of AgOTf and a solution of 0.0554 g (0.1241 mmol) of 1,2-bis(diphenylphosphino)benzene in 5 mL of anhydrous CH_2Cl_2 . 0.0759 g (yield 75%) was obtained as an orange powder.

^1H NMR (400 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 6.30 (*pseudo* q, 2H, H^1 , H^3), 6.48–6.54 (m, 2H, H^4 , H^7), 6.66 (t, $J = 3.2$ Hz, 1H, H^2), 6.89–6.96 (m, 2H, H^5 , H^6), 6.80–7.68 (bm, 24H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ -NMR (100.0 MHz, CDCl_3 , $T = 298$ K, ppm, *selected peaks*) δ : 89.6 (CH, t, $J_{\text{C-P}} = 11.9$ Hz, C^1 , C^3), 110.0 (bt, C^2), 118.6 (CH, C^4 , C^7), 127.3 (CH, C^5 , C^6), 128.6 (C, t, $J_{\text{C-P}} = 3.1$ Hz, C^{3a} , C^{7a}).

$^{31}\text{P}\{^1\text{H}\}$ NMR (162.0 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 57.7.

$^{19}\text{F}\{^1\text{H}\}$ NMR (377.2 MHz, CDCl_3 , $T = 298$ K, ppm) δ : -78.0.

IR (KBr pellet, cm^{-1}): $\nu_{\text{SO}} = 1313$, $\nu_{\text{CF}} = 1223$, 1149, $\nu_{\text{SO}} = 1030$, $\delta_{\text{SO}} = 636$.

Elemental analysis calcd (%) for $\text{C}_{40}\text{H}_{31}\text{F}_3\text{O}_3\text{P}_2\text{PdS}$: C, 58.80, H, 3.82, S, 3.92; found: C, 58.54, H, 3.99, S, 3.78.

HRMS calcd for $[\text{C}_{39}\text{H}_{31}\text{P}_2\text{Pd}]^{\dagger}$: 667.0936; found: 667.0989.

$[\text{Pd}(\text{DPEphos})(\text{indenyl})]\text{OTf}$ (8). Complex 8 was obtained by the same synthetic route of complex 4 using 0.0296 g

(0.0576 mmol) of $[\text{Pd}(\mu\text{-Cl})(\text{indenyl})]_2$ dissolved in 9 mL of anhydrous CH_2Cl_2 , 0.0311 mg (0.1210 mmol) of AgOTf and a solution of 0.0620 g (0.1241 mmol) of (oxydi-2,1-phenylene)bis(diphenylphosphine) in 2 mL of anhydrous CH_2Cl_2 . 0.0902 g (yield 87%) was obtained as an orange powder.

^1H NMR (400 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 5.59 (*pseudo* q, 2H, H^1 , H^3), 5.72–5.79 (m, 2H, H^4 , H^7), 6.64 (q, $J = 6.8$ Hz, 4H, Ar-H), 6.86–7.02 (m, 9H, H^2 , H^5 , H^6 , Ar-H), 7.27–7.64 (m, 18H, Ar-H).

$^{13}\text{C}\{^1\text{H}\}$ NMR (100.0 MHz, CDCl_3 , $T = 298$ K, *selected peaks*, ppm) δ : 95.0 (CH, C^1 , C^3), 112.6 (CH, t, $J_{\text{C-P}} = 5.6$ Hz, C^2), 118.5 (CH, C^4 , C^7), 128.0 (CH, C^5 , C^6), 131.5 (C, C^{3a} , C^{7a}).

$^{31}\text{P}\{^1\text{H}\}$ NMR (121.5 MHz, CDCl_3 , $T = 298$ K, ppm) δ : 15.8.

$^{19}\text{F}\{^1\text{H}\}$ NMR (377.2 MHz, CDCl_3 , $T = 298$ K, ppm) δ : -77.9.

IR (KBr pellet, cm^{-1}): $\nu_{\text{CF}} = 1222$, 1146, $\nu_{\text{SO}} = 1031$, $\delta_{\text{SO}} = 636$.

Elemental analysis calcd (%) for $\text{C}_{46}\text{H}_{35}\text{F}_3\text{O}_4\text{P}_2\text{PdS}$: C, 60.77, H, 3.88, S, 3.53; found: C, 61.03, H, 3.70, S, 3.39.

HRMS calcd for $[\text{C}_{45}\text{H}_{35}\text{OP}_2\text{Pd}]^{\dagger}$: 759.1198; found: 759.1264.

Cell viability assay

Three cancer cell lines (A2780, A2780*cis*, OVCAR-5) and one normal cell line (MRC-5) were employed and were grown in accordance with the supplier's instructions and maintained at 37 °C in a humidified atmosphere of 5% CO_2 . One thousand cancer cells and eight thousand MRC-5 cells were placed in a 96 well plate and treated after 24 h with six different concentrations of Pd(II) complexes (0.001, 0.01, 0.1, 1, 10, 100 μM). After 96 h of treatment, the cell viability was measured using a CellTiter-Glo assay (Promega, Madison, WI, USA) with a Tecan M1000 instrument. IC_{50} values were calculated from logistical dose-response curves. Averages were obtained from triplicate and error bars are standard deviations.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

Funding: this research was funded by Fondazione AIRC per la Ricerca sul Cancro, IG23566.

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